Level 3 Project Study Plan

2011 Cuyahoga River Environmental Monitoring

(1) Objectives

The lower Cuyahoga River has been designated as one of 42 Great Lakes Areas of Concern (AOC) by the International Joint Commission. Past monitoring indicated impairment of aquatic biota in the river and was the basis of a Total Maximum Daily Load (TMDL) for the Lower Cuyahoga River. The causes of impairment to the river were classified as organic enrichment, toxicity, low dissolved oxygen, nutrients and flow alteration (Ohio EPA, 2003)[1]. Recent monitoring by the Northeast Ohio Regional Sewer District and the Ohio EPA, however, has shown recovery in some reaches of the river. The purposes of this study, therefore, are to determine the attainment status of the river in relation to point and nonpoint sources of pollution and measure the magnitude of some potential causes of impairment.

During the course of the study, fish communities, benthic macroinvertebrate communities, habitat and water chemistry in the Cuyahoga River between River Mile (RM) 16.20 and RM 0.20 will be surveyed. The results from these surveys will be used to characterize the overall fish and macroinvertebrate community health in the river. Fish and macroinvertebrate community health will be evaluated through the use of Ohio EPA's Index of Biotic Integrity (IBI), Modified Index of Well-Being (MIwb), and Invertebrate Community Index (ICI). An examination of the specific characteristics of the biological communities will be used in conjunction with water quality data, the NEORSD Macroinvertebrate Field Sheet, and Qualitative Habitat Evaluation Index (QHEI) results in order to identify impacts to the communities. Results will also be compared to historic data to show temporal as well as spatial trends.

In addition, chlorophyll *a* levels in the river will be measured at four locations in the vicinity of Southerly Wastewater Treatment Center (WWTC) to determine any impacts from nutrients in the effluent on algal production. Data sondes will be installed in the river as part of this sampling to provide a more comprehensive understanding of the relationship among algal production, nutrient levels, and dissolved oxygen diel swings in the river.

Finally, the data collected from this study may be used to support three grants funded as part of the Great Lakes Restoration Initiative (GLRI). One of these grants is the *Cuyahoga River Larval Fish Study* funded by the U.S. Army Corps of Engineers that is being implemented by the Cuyahoga County Planning Commission. The second is a grant funded by the GLRI from the Cuyahoga County Engineer's Office titled *Cuyahoga AOC Urban Riparian Habitat*

[1] See Appendix I for a list of all references.

Restoration. The last one is the Ohio Department of Natural Resource's *Cuyahoga AOC Habitat and Fish Restoration Opportunities* grant. These projects are all focused on restoration of habitat and fish within the Cuyahoga River.

(2) Point/Nonpoint Sources

Point Sources	Nonpoint Sources
Tinkers Creek (RM 16.36)	Urban runoff
Mill Creek (RM 11.49)	Landfills
West Creek (RM 11.05)	Spills
Southerly WWTC (RM 10.57)	Agriculture
Ohio Canal (RM 8.78)	
Big Creek (RM 7.20)	
Combined Sewer Overflows	
Storm Sewer Outfalls	

A map has been provided in Appendix A to show point sources that may be influencing the water quality at each sample location. Other point sources (upstream of RM 16.20) include the Akron Wastewater Treatment Plant (RM 37.45), Combined Sewer Overflows in Akron, and numerous tributaries to the Cuyahoga River (see *Total Maximum Daily Loads for the Lower Cuyahoga River* [Ohio EPA, 2003] for a complete list of tributaries and their locations). These sources, along with the nonpoint sources listed in the table above, may be impacting the health of the fish and benthic macroinvertebrate communities in the Cuyahoga River or may lead to changes in algal and macrophyte production.

(3) Parameters Covered

Fish specimens will be identified to species level, weighed, counted and examined for the presence of external anomalies including DELTs (deformities, eroded fins, lesions and tumors). An Ohio EPA Fish Data Sheet will be completed during each assessment. Quantitative fish sampling is expected to be conducted at all locations, unless boat navigation is impeded by unforeseen manmade or natural barriers.

Cuyahoga River flow will be recorded for all locations during each electrofishing pass utilizing data from the United States Geological Survey (USGS) gage station in Independence, Ohio (Station ID # 04208000).

Macroinvertebrate community assemblages will be collected from each location between RMs 16.20 and 7.00 and transferred to Midwest Biodiversity Institute for identification and enumeration. Midwest Biodiversity Institute will identify the specimens to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b).

The NEORSD Macroinvertebrate Field Sheet (Appendix B) will be completed at each site during sampler retrieval. In addition, stream habitat will be measured by scoring components of the QHEI at all locations, including the substrate, instream cover, channel morphology, riparian zone and bank erosion, pool/glide and riffle/run quality and gradient.

Water chemistry samples will be collected at each electrofishing/ macroinvertebrate site from RMs 16.20 to 0.20 unless otherwise noted in Section 5. Water chemistry samples will be analyzed by NEORSD's Analytical Services. Appendix C lists the parameters to be tested along with the detection limits and practical quantitation limits. Field measurements for dissolved oxygen, pH, temperature, conductivity and turbidity will also be performed. A Surface Water Condition Sampling Field Data Form will be completed at each site during each sampling event (Appendix D).

Benthic and water column chlorophyll *a* samples will be collected at RMs 16.20, 10.75, 10.10, and 7.00. A Chlorophyll Sampling Data Sheet (Appendix J) will be completed for each sample collected. The chemical and physical water quality parameters to be measured in conjunction with the chlorophyll *a* samples include total phosphorus, dissolved reactive phosphorus, nitrate+nitrite, alkalinity, turbidity and suspended solids. In addition, YSI data sondes will be installed at these locations to more frequently monitor dissolved oxygen, temperature, conductivity, and pH.

(4) Field Collection and Data Assessment Techniques

Field collections for fish will be conducted with either a 14-foot Alweld commercial boat or 17-foot Coffelt electrofishing boat, both equipped with a Smith-Root 5.0 GPP Electrofisher. Boat electrofishing will consist of shocking all habitat types within a sampling zone, which is 0.5 kilometers in length, while moving from upstream to downstream. The stunned fish will be collected and put in an on-board live well for later processing. Each boat sampling zone will be electroshocked two or three times during the field season (June 15 - October 15).

Fish will be identified to species level, weighed, counted, and examined for the presence of external anomalies including DELTs. Fish easily identified (commonly collected from year to year) will be returned to the site from which they are collected. Subsamples of difficult to identify species will be brought back to the laboratory for verification by NEORSD Level 3 Fish Qualified Data Collectors (QDC) and, if necessary, sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Voucher specimens will be collected as described in section (14). Endangered species and those too large for preservation will not be collected as

voucher specimens, but will instead be photographed. Photographed vouchers will include features that permit definitive identification of the particular species.

Fish will be preserved in 10 percent formalin in the field, soaked in tap water for 24 to 48 hours after 5 to 7 days, then transferred to solutions of 30 and 50 percent ethanol for 5 to 7 days each and, finally, to 70 percent ethanol for long-term storage. Label information will include location (description and coordinates), date, time, collectors' names and sample identification code for each specimen collected.

Macroinvertebrate sampling will be conducted using quantitative and qualitative sampling techniques. Quantitative sampling will include installation of five replicates of a modified Hester-Dendy multi-plate artificial substrate sampler (HD) that is colonized for a six-week period. Multiple HD samplers will be installed at one or all of the locations in case samplers are lost due to vandalism, burial, etc. Qualitative sampling will be conducted using a D-frame dip net when HD samplers are retrieved. The NEORSD Macroinvertebrate Field Sheet will be completed during each HD retrieval. Voucher specimens will be collected as described in section (14). All other macroinvertebrate community assemblages will be shipped to Midwest Biodiversity Institute for identification and enumeration. Midwest Biodiversity Institute will identify specimens to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b). Voucher specimens will be collected as described in section (14). Stream flow will be measured with a Marsh-McBirney FloMate Model 2000 Portable Flow Meter or an Aquaflow Probe Model 6900, which measure flow in feet per second, when the HD samplers are installed and retrieved.

A detailed description of the sampling and analysis methods utilized in the fish community and macroinvertebrate surveys, including calculations of the IBI, MIwb and ICI, can be found in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volumes II* (1987a) and *III* (1987b).

The QHEI, as described in Ohio EPA's *Methods for Assessing Habitat in Flowing Waters: Using the Qualitative Habitat Evaluation Index (QHEI)* (2006) will be used to assess aquatic habitat conditions at each sample location.

Water chemistry sampling will occur across a variety of flow conditions. Techniques used for water chemistry sampling and chemical analyses will follow the *Manual of Ohio EPA Surveillance Methods and Quality Assurance Practices* (2009a). Chemical water quality samples from each site will be collected with one 4-liter disposable polyethylene cubitainer with disposable polypropylene lid and two 473-mL plastic bottles. Bacteriological samples will be collected in a sterile

plastic bottle preserved with sodium thiosulfate. All water quality samples will be collected as grab samples. One duplicate sample and one field blank will be collected at randomly selected sites, at the frequency not less than 10% of the total samples collected, for this study plan. The acceptable relative percent difference (RPD) for field duplicate samples will be ≤30 percent; results outside this range will trigger further evaluation and investigation into causes for disparities. RPD values above 30 percent, with results less then ten times the practical quantitation limit, will be reviewed on a case-by-case basis to determine if there is any merit for further investigation. Acid preservation of the samples, as specified in the NEORSD laboratory's standard operating procedure for each parameter, will occur in the field. Appendix C lists the analytical method, method detection limit and practical quantitation limit for each parameter analyzed. Field analyses include the use of either a YSI-556 MPS Multi-Parameter Water Quality Meter or YSI 600XL sonde to measure dissolved oxygen (DO), water temperature, conductivity and pH; and when necessary, a Hanna HI 98129 meter to measure pH and a Hach LDO meter to measured DO. Field turbidity will be measured using either a Hach 2100P IS Portable Turbidimeter, a LaMotte 2020 Portable Turbidity Meter, or an Orion AQUAfast AQ4500 Turbidimeter. Specifications for these meters have been included in Appendix E.

Benthic and water column chlorophyll a samples will be collected at least one time under low-flow conditions between June 15th and October 15th, 2011. Sampling methods will follow those detailed in the NEORSD *Chlorophyll a Sampling and Field Filtering Standard Operating Procedure* (SOP-EA001-00). A Chlorophyll *a* Sampling Field Sheet will be completed for each site. Water chemistry grab samples will be collected at the same time using the methods discussed previously and will be analyzed for nutrients, turbidity, alkalinity and suspended solids. In addition, at least 24 hours prior to each chlorophyll *a* sampling event, YSI 6600 EDS data sondes will be deployed at the four locations in the Cuyahoga River. Each data sonde will record, at fifteen-minute intervals, dissolved oxygen concentration, pH, temperature, and conductivity from the time the data sonde is deployed until the time it is retrieved. These data sondes will be placed in the stream by inserting each one into a 4.5-inch PVC pipe with holes drilled into the sides of the lower third of the pipe to allow water to pass through it. The data sondes will remain in the river at least 24 hours following collection of the chlorophyll *a* samples.

Where possible, data assessment will include an analysis of temporal and spatial trends in the collected data. Species assemblages and individual metrics will be analyzed. Graphs that show current and historic QHEI, IBI, MIwb and ICI scores and how these scores compare to attainment status of biocriteria will be prepared. Water chemistry data collected will be compared to Ohio water quality standards to determine whether any excursions from the applicable water quality criteria

have occurred. It will also be used to determine any relationships among individual parameters and chlorophyll *a* concentrations. Comparisons between water quality and biological community health will only be made if at least three water quality samples have been collected from that site.

(5) Sampling Locations

The following electrofishing and macroinvertebrate sample locations, listed from upstream to downstream on the Cuyahoga River, will be surveyed during this study. HD and water chemistry collection sites are located near the mid point of each electrofishing zone, indicated by river mile, unless otherwise noted. GPS coordinates are recorded at the downstream end of each electrofishing zone.

Location	Latitude	Longitude	River Mile	Description	Quadrangle	Purpose
Downstream of Tinkers Creek	N41.3678°	W81.6139°	16.20	Downstream of the confluence with Tinkers Creek near Old Riverview Road	Northfield	Background data for fish, habitat, macroinvertebrates, and chlorophyll *a*
Upstream of Mill Creek	N41.4123° N41.4101°	W81.6364° W81.6346°	12.10[2] 11.95	Upstream of the confluence with Mill Creek (I-480)	Cleveland South	Evaluate Mill Creek discharge on fish, habitat and macroinvertebrates
Downstream of Mill Creek	N41.4179°	W81.6446°	11.30	Downstream of the confluence with Mill Creek	Cleveland South	Evaluate Mill and West Creek discharges on fish, habitat and macroinvertebrates
Upstream of Southerly WWTC	N41.4196°	W81.6547°	10.75	Upstream of Southerly WWTC effluent discharge	Cleveland South	Evaluate West Creek and Southerly WWTC discharges on fish, habitat and macroinvertebrates, and Southerly WWTC discharge on chlorophyll *a* levels.
Downstream of Southerly WWTC	N41.4242°	W81.6638°	10.10	Downstream of Southerly WWTC effluent discharge	Cleveland South	Evaluate Southerly WWTC discharge on fish, habitat, macroinvertebrates, and chlorophyll *a* levels.
Upstream of Big Creek	N41.4381°	W81.6680°	8.60	Upstream of the confluence with Big Creek	Cleveland South	Evaluate Big Creek discharge on fish, habitat and

[2] HD and Water Chemistry Collection Site

Location	Latitude	Longitude	River Mile	Description	Quadrangle	Purpose
						macroinvertebrates
Downstream of Big Creek	N41.4497°	W81.6815°	7.00	Downstream of the confluence with Big Creek/ Upstream of habitat restoration project	Cleveland South	Evaluate Big Creek discharge on fish, habitat and macroinvertebrates; Southerly WWTC discharge on chlorophyll *a* levels; and effectiveness of habitat restoration in navigation channel on fish.
Head of Navigation Channel	N41.4619°	W81.6816°	5.90	Head of navigation channel/Upstream of artificial habitat near ArcelorMittal	Cleveland South	Evaluate effectiveness of habitat restoration in navigation channel on fish.
Abandoned Marina (formerly Scaravelli's)	N41.4881°	W81.6938°	2.75	Mid-navigation channel/Proposed site of GLRI habitat restoration project	Cleveland South	Evaluate effectiveness of habitat restoration in navigation channel on fish.
Cuyahoga River Mouth	N41.5008°	W81.7098°	0.20	Near mouth of river in navigation channel	Cleveland North	Evaluate effectiveness of habitat restoration in navigation channel on fish.

(6) Schedule

Two to three electrofishing surveys will be conducted between June 15 and October 15, 2011, at all locations. Additional qualitative surveys may be conducted at RM 7.00 in April and May of 2011 to determine which species are staging to spawn upstream of the proposed habitat restoration. The qualitative surveys will be used to support the GLRI grants. Surveys will be conducted at least three weeks apart. Specific dates have not been scheduled. River flow and weather conditions will be assessed weekly to determine when each electrofishing pass will be conducted.

Artificial substrate samplers will be installed on the Cuyahoga River once, between June 15 and August 19, 2011, and retrieved six weeks later. Specific dates have not been scheduled. River flow and weather conditions will be assessed weekly to determine when the HD sampler installations and retrievals will be conducted.

QHEI habitat evaluations will be conducted one time between June 15 and October 15, 2011. These evaluations will be conducted around the same time as one of the electrofishing surveys.

Water chemistry samples will be collected a minimum of three times between June 15 and October 15, 2011.

Benthic and water column chlorophyll *a* samples will be collected at least once between June 15 and October 15, 2011. YSI data sondes will be installed at least 24 hours prior to the time that each sample is collected, and retrieved at least 24 hours after the sample is collected.

(7) QA/QC

Quality assurance and quality control of sampling and analysis methods for habitat, fish, and macroinvertebrate evaluations will follow Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volumes II* (1987a) and *III* (1987b) and *Methods for Assessing Habitat in Flowing Waters: Using the Qualitative Habitat Evaluation Index (QHEI)* (2006).

Electrofishing equipment will be used according to the guidelines listed in the operation and maintenance manual provided by Smith-Root, Inc. Malfunctioning equipment will not be used to collect data. Proper steps will be taken to correct any problems as soon as possible, whether by repairing in the field, at the NEORSD Environmental & Maintenance Services Center, or by contacting the supplier or an appropriate service company.

Subsamples of difficult to identify fish species will be brought back to the laboratory for verification by Level 3 Fish Qualified Data Collectors (QDC), and if necessary, sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Voucher specimens will be collected as described in section (14). Endangered species and those too large for preservation will not be collected as voucher specimens, but will instead be photographed. Photographed vouchers will include features that permit definitive identification of the particular species.

All macroinvertebrate community assemblages will be collected and shipped to Midwest Biodiversity Institute for identification and enumeration. Midwest Biodiversity Institute will identify specimens to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b). The Midwest Biodiversity Institute QA/QC manual is attached (Appendix F). All macroinvertebrate specimens will be returned to NEORSD by Midwest Biodiversity Institute. At least two voucher

specimens of each species, when available, will be separated into individual vials and kept as described in section (14). The remaining specimens for each site will be returned in a single container labeled with the site number and collection method and date. All specimens and accompanying chain-of-custody documentation will be retained by NEORSD and stored at the Environmental & Maintenance Services Center for a period not less than ten years.

Water samples obtained for chemical analyses will be collected, preserved (see Section 4), labeled and then placed on ice inside the field truck. The field truck will remain locked at all times when not occupied/visible. Sampling activities, including sample time and condition of surface water sampled, will be entered in a field log book and on the Surface Water Condition Sampling Field Data Form. The samples will then be delivered immediately to the NEORSD Analytical Services cooler, after which the door to the cooler will be locked, and the samples will be transferred to the custody of Analytical Services. The NEORSD Analytical Services Quality Manual and associated Standard Operating Procedures are on file with Ohio EPA. The Quality Assurance Officer at Analytical Service will send updates, revisions and any information on document control to Ohio EPA as needed.

Three filtrations will be completed for each benthic and water column chlorophyll a sample. In addition, a field filtration blank will be submitted for every 20 samples.

Calibration of data sondes will be done according to the YSI Environmental Operations Manual. The conductivity will be calibrated first using a 1.413 mS/cm standard. Second, the pH will be calibrated using two different buffers (7 and 10 s.u.). The DO will be calibrated last. The acceptable error in the DO calibration will be 0.2 mg/L.

Each data sonde will have a separate calibration and maintenance logbook. As the appropriate calibration procedures are completed, the results will be recorded in the logbook along with the name of the person performing the calibration and the date. If difficulty is encountered in calibrating an instrument, or if the instrument will not hold calibration, this information will also be recorded.

Malfunctioning equipment will not be used to collect data. Proper steps will be taken to correct any problems as soon as possible, whether by repairing the sonde in the field or at the Environmental & Maintenance Services Center or by sending it to the supplier or an appropriate service company. All equipment maintenance will be recorded in the logbook.

Once the YSI 6600 EDS data sondes are removed from the river, the accuracy of the data that has been collected will be checked by comparing readings taken by the sondes to known standards. If the measurements taken at this time meet quality control goals, all of the data collected since the last calibration will be considered accurate. The acceptable differences for pH and conductivity will be $\pm$0.3 with pH 7 buffer and $\pm$ 10% of the conductivity standard, respectively (EPA New England- Region 1, 2005). The acceptable difference for DO will be $\pm$0.2 mg/L. If the measurements do not meet quality control goals, best professional judgment will be used to decide if any of the data collected during that period may still be accurate. For example, the data collected from the four locations may be plotted on the same graph, and if it appears that the data points are following similar trends, they may be considered accurate. If any data that do not meet quality control goals are used, a rationale for their inclusion will be provided when the data are submitted.

(8) Work Products

Within one year of completion of the project, fish data (species, numbers, weights, pollution tolerances, the incidence of DELT anomalies, IBI and MIwb scores), macroinvertebrate data (types and numbers of macroinvertebrates collected and ICI scores), habitat data (QHEI raw data and scores) and water chemistry results will be submitted to the Ohio EPA. Additionally, reports summarizing, interpreting, graphically presenting and discussing the IBI, MIwb, ICI and QHEI scores, chlorophyll *a* results, and any excursions from water quality standards may be prepared for internal use.

(9) Qualified Data Collectors

The following Level 3 Qualified Data Collectors (QDC) will be involved with this study:

Name	Address	Email Address	Phone Number	QDC Specialty(s)
John W. Rhoades[1]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	rhoadesj@neorsd.org	216-641-6000	QDC - 00008 CWQA/FCB/SHA/ BMB
Cathy Zamborsky	4747 East 49th Street Cuyahoga Hts., Ohio 44125	zamborskyc@neorsd.org	216-641-6000	QDC - 00009 CWQA/SHA
Seth Hothem[2,3]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	hothems@neorsd.org	216-641-6000	QDC - 00010 CWQA/FCB/SHA
Kathryn Crestani	4747 East 49th Street Cuyahoga Hts., Ohio 44125	crestanik@neorsd.org	216-641-6000	QDC - 00011 CWQA/SHA
Tom Zablotny	4747 East 49th Street Cuyahoga Hts., Ohio 44125	zablotnyt@neorsd.org	216-641-6000	QDC - 00018 CWQA/FCB/SHA
Ron Maichle[4]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	maichler@neorsd.org	216-641-6000	QDC - 00145 CWQA/SHA/BMB
Francisco Rivera[6]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	riveraf@neorsd.org	216-641-6000	QDC - 00262 CWQA/SHA
Kristina Granlund	4747 East 49th Street Cuyahoga Hts., Ohio 44125	granlundk@neorsd.org	216-641-6000	QDC – 00511 CWQA/SHA
Jillian Novak	4747 East 49th Street Cuyahoga Hts., Ohio 44125	novakj@neorsd.org	216-641-6000	QDC – 00512 CWQA/SHA
Martin Knapp[5]	Midwest Biodiversity Institute P.O. Box 21561 Columbus, Ohio 43221	martygator@hotmail.com	614-457-6000	QDC - 300 BMB

[1]Lead Project Manager
[2]Fish Community Biology (FCB) Project Manager
[3]Stream Habitat Assessment (SHA) Project Manager
[4]Benthic Macroinvertebrate Biology (BMB) Project Manager
[5]Benthic Macroinvertebrate Identification
[6]Chemical Water Quality Assessment (CWQA) Project Manager

The following is a list of persons not qualified as QDCs who may be involved in the project. Prior to the start of sampling, the project managers will explain to each individual the proper methods for sampling. Sampling will only be completed under the direct observation of a QDC. The lead project manager will be responsible for reviewing all reports and data analysis prepared by qualified personnel prior to completion.

Name	Address	Email Address	Phone Number
Nick Barille	4747 East 49th Street Cuyahoga Hts., Ohio 44125	barillen@neorsd.org	216-641-6000
Joseph Broz	4747 East 49th Street Cuyahoga Hts., Ohio 44125	brozj@neorsd.org	216-641-6000
Joseph Carbonaro	4747 East 49th Street Cuyahoga Hts., Ohio 44125	carbonaroj@neorsd.org	216-641-6000
Tim Dobriansky	4747 East 49th Street Cuyahoga Hts., Ohio 44125	dobrianskyt@neorsd.org	216-641-6000
Kyle Frantz	4747 East 49th Street Cuyahoga Hts., Ohio 44125	frantzk@neorsd.org	216-641-6000
Rae Grant	4747 East 49th Street Cuyahoga Hts., Ohio 44125	grantr@neorsd.org	216-641-6000
Eric Hinton	4747 East 49th Street Cuyahoga Hts., Ohio 44125	hintone@neorsd.org	216-641-6000
Mark Matteson	4747 East 49th Street Cuyahoga Hts., Ohio 44125	mattesonm@neorsd.org	216-641-6000
Denise Phillips	4747 East 49th Street Cuyahoga Hts., Ohio 44125	phillipsd@neorsd.org	261-641-6000

Name	Address	Email Address	Phone Number
Kevin Roff	4747 East 49th Street Cuyahoga Hts., Ohio 44125	roffk@neorsd.org	216-641-6000
Frank Schuschu	4747 East 49th Street Cuyahoga Hts., Ohio 44125	schuschuf@neorsd.org	216-641-6000
Wolfram von Kiparski	4747 East 49th Street Cuyahoga Hts., Ohio 44125	vonkiparskiw@neorsd.org	216-641-6000
Amy Erzen Summer Co-op	4747 East 49th Street Cuyahoga Hts., Ohio 44125	erzena@neorsd.org	216-641-6000
Summer Co-op #2	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000
Summer Co-op #3	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000
Summer Co-op #4	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000

(10) Documentation of approval of project managers and other personnel as level 3 qualified data collectors

See attached (Appendix G).

(11) Contract laboratory contact information

Any fish that is not positively identified in the field or NEORSD laboratory will be sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Fish will be identified to the species level.

Dr. Ted Cavender, Curator of Fish / Mr. Marc Kibbey, Associate Curator of Fish
1315 Kinnear Road, Columbus, Ohio 43212
cavender.1@osu.edu / kibbey.3@osu.edu
614-292-7873

Identification of macroinvertebrates will be completed by Midwest Biodiversity Institute. Benthic macroinvertebrates will be identified to the lowest practical level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b).

Mr. Chris Yoder
Midwest Biodiversity Institute
P.O. Box 21561
Columbus, Ohio 43221
yoder@rrohio.com
614-457-6000

(12) Copy of ODNR collector's permit

To be submitted electronically when issued by ODNR (Appendix H).

(13) Catalog Statement

A digital photo catalog of all sampling locations will be maintained for 10 years and will include photos of the specific sampling location(s), the riparian zone adjacent to the sampling location(s) and the general land use in the immediate vicinity of the sampling location(s).

Print/Signature: John W. Rhoades / Date: _____

(14) Voucher Specimen Statement

NEORSD will maintain a benthic macroinvertebrate and fish voucher collection which includes two specimens, or appropriate photo vouchers, of each species or taxa collected during the course of biological sampling from any stream within the NEORSD's service area. When benthic macroinvertebrates from multiple surface waters are collected within the same year and identified by the same QDC, one voucher collection will be created to represent the specimens collected from those streams. When fish specimens from multiple surface waters are collected within the same year, one voucher collection will be created to represent the specimens collected from those streams. A separate collection for each sampling event will not be maintained.

NEORSD will provide specimens or photo vouchers to the Director upon request. This collection will be stored at the NEORSD laboratory in the Environmental and Maintenance Services Center.

Print/Signature: John W. Rhoades / Date: _____

(15) Trespassing Statement

I have not been convicted or pleaded guilty to a Violation of section 2911.21 of the Revised Code (criminal trespass) or a substantially similar municipal ordinance within the previous five years.

Print/Signature: John W. Rhoades / _____ Date: _____

Print/Signature: Cathy Zamborsky / _____ Date: _____

Print/Signature: Seth Hothem / _____ Date: _____

Print/Signature: Kathryn Crestani / _____ Date: _____

Print/Signature: Tom Zablotny / _____ Date: _____

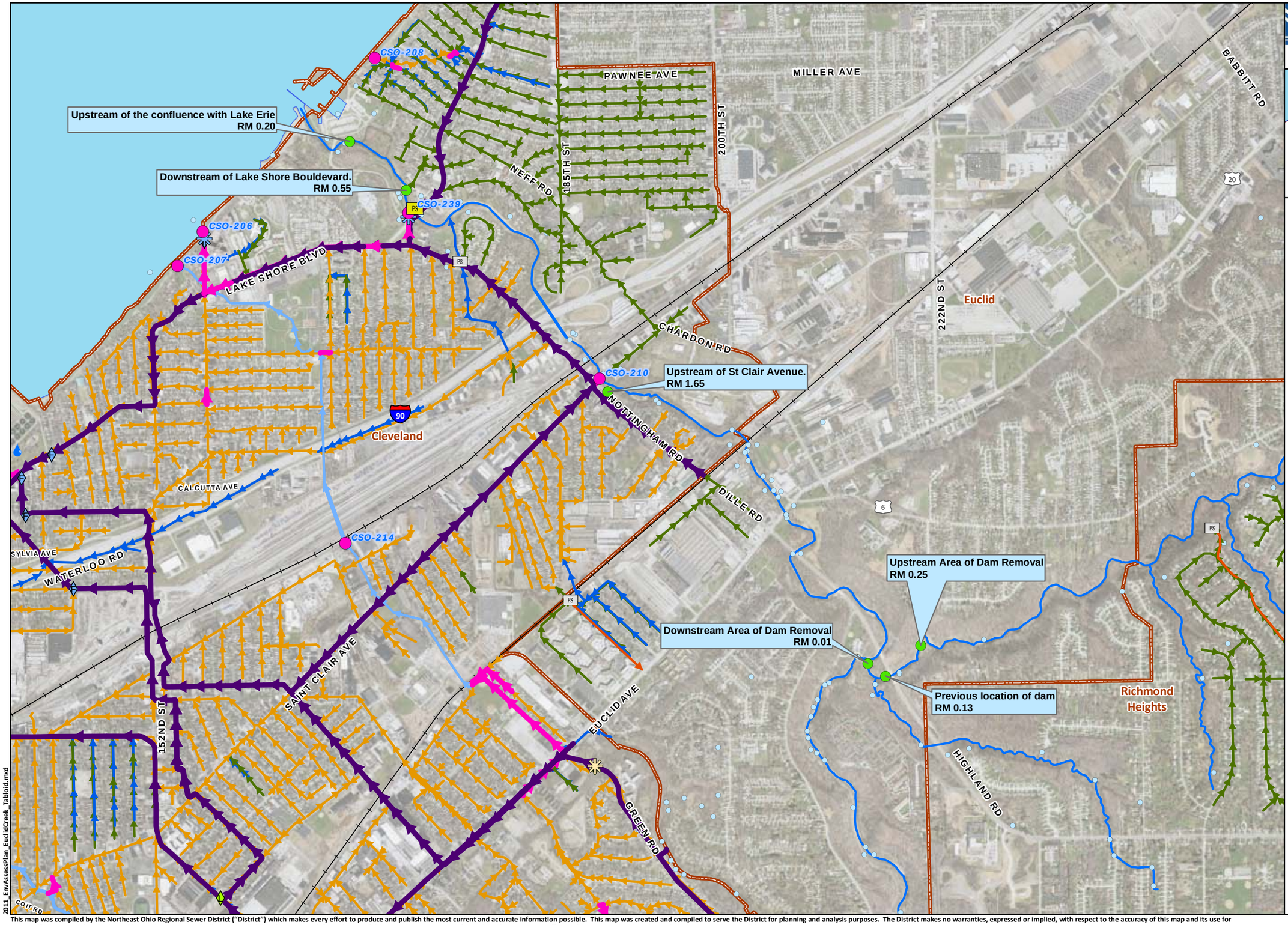
Print/Signature: Ron Maichle / _____ Date: _____

Print/Signature: Francisco Rivera / _____ Date: _____

Print/Signature: Kristina Granlund / _____ Date: _____

Print/Signature: Jillian Novak / _____ Date: _____

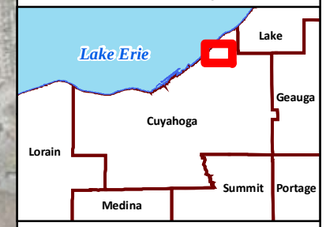
Appendix A



Northeast Ohio Regional Sewer District

Protecting Your Health and Environment

Overview Map



Euclid Creek 2011 Environmental Assessment

- Outfalls
- Stream
- District CSO Permit Point
- District Facility
- Flow Meter
- Level Monitor
- Rain Gauge
- Interceptor
- CSO Responsibility
- Sludge Force Main
- Combined Sewer
- CSO Overflow Sewer
- Culverted Stream
- Sanitary Sewer
- Storm Sewer
- Force Main

Study Sites

- -Upstream of Lake Erie confluence
  -Lat.41.5856 Long. -81.5629
- -Downstream of Lake Shore Blvd.
  -Lat.41.5833 Long. -81.5594
- -Upstream of St. Clair Avenue
  -Lat.41.5738 Long. -81.5470
- -Downstream of dam removal
  - Lat. 41.5610 Long. -81.5310
- -Previous dam location
  -Lat. 41.5604 Long. -81.5299
- -Upstream of dam removal
  -Lat.41.5618 Long. -81.5277

N

1,100

Feet

Date: March 2011

Sources: Orthophotos(2008), Street Centerline (2006), Cuyahoga County GIS; Streams(2005), SSURGO; Environmental Assessment Zones (2009), Collection System, NEORSD GIS;

2011_EnvAssessPlan_EuclidCreek_Tabloid.mxd

Appendix B

NEORSD Macroinvertebrate Field Sheet

Stream: _____ River Mile: _____ Year: _____

Location: _____ Project: _____

Drainage Area (mi^2): _____ Latitude (ºN)/Longitude (ºW): _____

Hester-Dendy Deployment Information

Install Date: _____ Crew Initials (QDC Circled): _____

Current at HD (fps): _____ Depth (cm): _____ Pictures Obtained: Yes No

Reinstall Date: _____ Crew Initials (QDC Circled): _____

Current (fps): _____ Depth (cm): _____ Reason: _____

Reinstall Date: _____ Crew Initials (QDC Circled): _____

Current (fps): _____ Depth (cm): _____ Reason: _____

Sampling/Retrieval Information

Sampling Method: Hester-Dendy Dipnet Surber Grab Other: _____

Sampling ID: HD: _____ Qualitative: _____ Other: _____

Sampling Date: _____ Crew Initials (QDC Circled): _____

HD Condition- Current (fps): _____ Depth (cm): _____ Water Temp: _____ ºF / ºC

Number of HD Blocks Obtained: _____ Remarks: _____

Disturbed: Yes No Comments: _____

Debris: Yes No Comments: _____

Silt/Solids: None Slight Moderate Heavy

Dipnet- Time Sampled (min): _____ X Number of Crew: _____ = Total (min): _____

Habitats Sampled: Pool Riffle Run Margin Backwater

Samples Analyzed By: _____ QDC #: _____ Date: _____

River Sampling Conditions

Flow Condition:	Flood	Above Normal	Normal	Low	Interstitial	Intermittent	Dry
Current Velocity:	Fast	Moderate	Slow	Non-detect			
Channel Morphology:	Natural	Channelized	Channelized (Recovered)		Impounded		
Bank Erosion:	Extensive	Moderate	Slight	None			
Riffle Development:	Extensive	Moderate	Sparse	Absent			
Riffle Quality:	Good	Fair	Poor	*Embedded:*	Yes	No	
Water Clarity:	Clear	Murky	Turbid	Other: _____			
Water Color:	None	Green	Brown	Grey	Other: _____		
Canopy:	Open	75 %	50 %	25 %	Closed		

Comment Section: _____

Physical Characteristics

Substrate Characteristics

	Pool	Units	Riffle	Units	Run	Units
Bedrock						
Boulder						
Rubble						
Coarse Gravel						
Fine Gravel						
Sand						
Silt						
Clay/Hardpan						
Detritus						
Peat						
Muck						
Other						
Macrophytes						
Algae						
Artifacts						
Compaction (F,M,S)						
Depth (Avg)						
Width (Avg)						

Predominant Land Use (Left, Right or Both)

Forest	Urban	Open Pasture
Shrub	Residential/Park	Closed Pasture
Old Field	Mining/Construction	
Rowcrop	Wetland	
Industrial	Other	

Predominant Riparian Vegetation

Left	Right	Type
		Large Trees
		Small Trees
		Shrubs
		Grass/Weeds
		None

Margin Habitat

Margin Quality: Good Fair Poor

Undercut Banks	Root Mats
Grass	Water Willow
Shallows	Clay/Hardpan
Rip Rap	Bulkhead
Other	

Biological Characteristics

Riffle:

Predominant Organism: _____

Other Common Organisms: _____

Density: High Moderate Low

Diversity: High Moderate Low

Run:

Predominant Organism: _____

Other Common Organisms: _____

Density: High Moderate Low

Diversity: High Moderate Low

Pool:

Predominant Organism: _____

Other Common Organisms: _____

Density: High Moderate Low

Diversity: High Moderate Low

Margin:

Predominant Organism: _____

Other Common Organisms: _____

Density: High Moderate Low

Diversity: High Moderate Low

Other Notable Collections: _____

V= Very Abundant; A= Abundant; C= Common; R= Rare

Overall Amount (V= >151; A= 150-101; C= 100-11; R= 10-1)	
/	Porifera, Bryozoa
/ /	Turbellaria, Oligochaeta, Hirudinea
/	Isopoda, Amphipoda
/	Decapoda, Hydracarina
	Ephemeroptera
	Baetidae
	Other _____
/	Zygoptera, Anisoptera
	Plecoptera
	Hemiptera
/	Megaloptera, Neuroptera
	Trichoptera
	Hydropsychidae
	Other _____
	Coleoptera
	Elimidae
	Other _____
	Diptera
	Chironomidae
	Other _____
/	Gastropoda, Bivalvia
	Other _____
	Other _____
	Other _____

Appendix C

Parameter	Additional Name	Test	Minimum Detection Limit	Practical Quantitation Limit
Alkalinity		EPA 310.2	1.5 mg/L	10 mg/L
Chemical Oxygen Demand	COD	EPA 410.4	5 mg/L	10 mg/L
Hexavalent Chromium	Hex Chrome	SM 3500 Cr D. [1]	1 µg/L	5 µg/L
Mercury	Hg	EPA 245.1	0.005 µg/L	0.050 µg/L
Ammonia *	NH_3	EPA 350.1	0.002 mg/L	0.010 mg/L
Nitrite + Nitrate	$NO_2 + NO_3$	EPA 353.2	0.001 mg/L	0.010 mg/L
Nitrite	NO_2	SM 4500-$N0_2^-$ B. [1]	0.002 mg/L	0.010 mg/L
Nitrate	NO_3	EPA 353.2	0.001 mg/L	0.010 mg/L
Soluble Phosphorus	Soluble-P	EPA 365.1	0.004 mg/L	0.010 mg/L
Total Phosphorus	Total-P	EPA 365.1	0.002 mg/L	0.010 mg/L
Chlorophyll *a*	Chlorophyll *a*	EPA 445.0	To be determined	2.0 µg/L
Chloride	Chloride by IC	EPA 300.0	0.057 mg/L	5.000 mg/L
Sulfate	Sulfate by IC	EPA 300.0	0.046 mg/L	5.000 mg/L
Biological Oxygen Demand	BOD	SM 5210 [1]	2 mg/L	5 mg/L
Silver	Ag	EPA 200.7	0.12 µg/L	1.00 µg/L
Aluminum	Al	EPA 200.7	3.7 µg/L	10.0 µg/L
Arsenic	As	EPA 200.7	0.31 µg/L	2.00 µg/L
Barium	Ba	EPA 200.7	0.12 µg/L	2.00 µg/L
Beryllium	Be	EPA 200.7	0.12 µg/L	1.00 µg/L
Calcium	Ca	EPA 200.7	11.2 µg/L	275.0 µg/L
Cadmium	Cd	EPA 200.7	0.022 µg/L	1.00 µg/L
Cobalt	Co	EPA 200.7	0.15 µg/L	1.00 µg/L
Chromium	Cr	EPA 200.7	0.25 µg/L	2.00 µg/L
Copper	Cu	EPA 200.7	0.17 µg/L	1.00 µg/L
Iron	Fe	EPA 200.7	1.5 µg/L	10.00 µg/L
Potassium	K	EPA 200.7	31.4 µg/L	275.0 µg/L
Magnesium	Mg	EPA 200.7	40.9 µg/L	100.0 µg/L
Manganese	Mn	EPA 200.7	0.038 µg/L	1.00 µg/L
Molybdenum	Mo	EPA 200.7	0.31 µg/L	1.00 µg/L
Sodium	Na	EPA 200.7	59.5 µg/L	500.0 µg/L
Nickel	Ni	EPA 200.7	0.17 µg/L	2.00 µg/L
Lead	Pb	EPA 200.7	0.39 µg/L	3.00 µg/L
Antimony	Sb	EPA 200.7	0.61 µg/L	5.00 µg/L
Selenium	Se	EPA 200.7	0.63 µg/L	5.00 µg/L
Tin	Sn	EPA 200.7	13.4 µg/L	50.00 µg/L
Titanium	Ti	EPA 200.7	0.22 µg/L	2.00 µg/L
Thallium	Tl	EPA 200.7	1.10 µg/L	5.00 µg/L
Vanadium	V	EPA 200.7	0.15 µg/L	1.00 µg/L
Zinc	Zn	EPA 200.7	1.6 µg/L	10.00 µg/L
Total Metals	Total Metals (calc.)	EPA 200.7	µg/L =(Cr µg/L)+(Cu µg/L)+(Ni µg/L)+(Zn µg/L)	
Hardness	Hardness (calc.)	SM 2340 B [1]	CaCO3 mg/L =(2.497*Ca mg/L)+(4.118*Mg mg/L)	
Total Solids	TS	SM 2540 B [1]	0.5 mg/L	1.0 mg/L
Total Suspended Solids	TSS	SM 2540 D [1]	0.5 mg/L	1.0 mg/L
Total Dissolved Solids	TDS	SM 2540 C [1]	0.5 mg/L	1.0 mg/L
Turbidity **		EPA 180.1	0.1 NTU	0.2 NTU
Escherichia coli	*E. coli*	EPA 1603 D	1 colony	--
Field Parameter		**Test**	**(Value Reported in)**	
pH		SM 4500H-B [1]	s.u.	
Conductivity		SM 2510A [1]	µs/cm	
Dissolved Oxygen	DO	SM 4500-0 G [1]	mg/L	
Temperature	Temp	SM 2550B [1]	°C	
Turbidity **		EPA 180.1	NTU	

*NOTE: Listed MDL/PQL is for undistilled samples. Any samples that are required to be distilled will have a MDL = 0.044 mg/L, PQL = 0.100 mg/L

** Turbidity will either be completed in the field or at the laboratory.

[1] Standard Methods for the Examination of Water and Wastewater, 19th Edition

Appendix D

NEORSD Surface Water Condition Sampling Field Data Form

Stream: _____ Date: _____ Collectors: _____

Gage Station and ID: _____ Daily Mean Discharge: _____ ft^3/sec

Was this sample taken during or following a wet weather event? YES / NO

Water Quality Meters Used: _____

Time (hrs): _____ River Mile (Site): _____

Weather: Clear Partly Cloudy Overcast Light Rain/Showers Heavy Rain
Steady Rain Heavy Snow Melt Other: _____

Flow: Dry Intermittent Minimal Baseline/Normal Elevated Flood

HD Status: OK Buried Out of Water H-D was Reset
Unknown (river too high) Missing Not Installed Flow: _____ fps

Color: Clear Muddy Tea Milky Other: _____

Odor: Normal Petroleum Anaerobic Sewage Chemical Other: _____

Surface Coating: None Foam Oily Scum Other: _____

Field Parameters: Conductivity (µmhos/cm): _____ Temperature (°C): _____

Dissolved Oxygen (mg/L): _____ pH (s.u.): _____

Turbidity (NTU): _____

Sample ID:

General Comments: _____

Time (hrs): _____ River Mile (Site): _____

Weather: Clear Partly Cloudy Overcast Light Rain/Showers Heavy Rain
Steady Rain Heavy Snow Melt Other: _____

Flow: Dry Intermittent Minimal Baseline/Normal Elevated Flood

HD Status: OK Buried Out of Water H-D was Reset
Unknown (river too high) Missing Not Installed Flow: _____ fps

Color: Clear Muddy Tea Milky Other: _____

Odor: Normal Petroleum Anaerobic Sewage Chemical Other: _____

Surface Coating: None Foam Oily Scum Other: _____

Field Parameters: Conductivity (µmhos/cm): _____ Temperature (°C): _____

Dissolved Oxygen (mg/L): _____ pH (s.u.): _____

Turbidity (NTU): _____

Sample ID:

General Comments: _____

Appendix E

2100P IS Portable Turbidimeter

Specifications

2100P Portable Turbidimeter Specifications:

Ranges:	0-1000 NTU with automatic decimal point placement or manual range selection of 0-9.99, 0-99.9 and 0-1000 NTU selection.
Accuracy:	± 2% of reading plus stray light from 0 to 1000 NTU (stray light: <0.02 NTU)
Repeatability:	± 1% of reading or ± 0.01 NTU, whichever is greater
Resolution:	0.01 NTU on lowest range
Sample Required:	15 mL
Power Requirement:	Four AA alkaline batteries or optional 120 or 230 Vac battery eliminator.
Construction:	High-impact ABS plastic shell
Dimensions:	22.2 x 9.5 x 8.9 cm (8.75 x 3.75 x 3.5")
Shipping Weight:	3.6 kg (8 lb)
Warranty:	Two years

Specifications subject to change.

MAIN PRODUCT PAGE

» 2100P IS Portable Turbidimeter

▲ Go to top

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2020we & 2020wi Portable Turbidity Meters

The Clear Choice for Turbidity Measurement!

Industry-leading precision, sensitivity, and dependability in one of the most innovative **handheld meters** available on the market!

- **Waterproof** to IP67
- Lithium rechargeable battery
- USB port
- 7 languages
- Backlit display
- EPA and ISO versions

2020we Complies with USEPA 180.1 Standard

2020wi Complies with ISO 7027 Standard

Kit supplied with 0, 1, and 10 NTU standard, sample bottle, 4 sample tubes, USB cable, USB computer/wall adapter, and waterproof carrying case.

Order Codes	
1970-EPA	**2020we Kit:** Portable turbidity meter complies with USEPA 180.1 Standard
1970-ISO	**2020wi Kit:** Portable turbidity meter complies with ISO 7027 Standard

Turbidity Specifications Meter Features Order Codes

Turbidity Specifications:	
Unit of Measure:	2020we: NTU, AU, ASBC, EBC 2020wi: FNU, FAU, ASBC, EBC
Range:*	0-4000 NTU/FNU, 0-10,500 ASBC, 0-150 EBC
Resolution:*	0.01 NTU/FNU 10.00-10.99 0.1 NTU/FNU 11.00-109.9 1 NTU/FNU 110-4000
Accuracy:*	From 0-2.5 NTU the accuracy is ±0.05 NTU From 2.5-100 NTU the accuracy is ±2% From 100 NTU the accuracy is ±3%
Detection Limit:	0.05 NTU/FNU
Range Selection:	Automatic
Reproducibility:*	0.02 NTU/FNU or 1%
Light Source:	Tungsten (EPA) complies with EPA 180.1 Standard 860 LED (ISO) complies with ISO 7027

**Over 600 NTU/FNU units expressed as AU/FAU*

Water Quality Turbidity Meter

geotech

Orion AQUAfast AQ4500 Turbidimeter

Thermo Electron introduces the Orion AQ4500 Turbidimeter which offers advanced features not available on any other benchtop or portable turbidimeter. The AQ4500 offers a dual source LED which allows readings that comply with both EPA 180.1 and ISO 7027. Turbidity can be read in the range of 0 - 1000 NTU with a choice of units: NTU, FTU, FNU, ASBC, and EBC. In the range of 0 - 40 NTU the AQ4500 offers a ratiometric range which will give EPA, GLI method 2 equivalent numbers. This portable field unit is truly IP67 waterproof with typical battery life of over 1000 hours on one set of batteries and datalog capacity of 100 points which can later be downloaded to a printer or computer. The AQ4500 accepts 24 mm cuvettes and comes with a two year warranty.

FEATURES

- Nephelometric and Ratiometric measurements with Autoranging
- Data log capacity of up to 100 data points
- Readings in the range of 0 - 1000 NTU with a choice of units:
  NTU, FTU, FNU, ASBC, or EBC
- Includes Turbidity Standards kit, rugged carrying case, and replacement cuvettes
- Orion AQ4500 is truly IP67 waterproof to a depth of 3 meters

SPECIFICATIONS

Type	Turbidity Meter
Principle of Operation	Nepeholmetric
Operating Modes	Automatic
Measurement Modes	Automatic
Ranges	
NTU	0 - 2000
Nephelometric	0 - 4000
EPA	0 - 4000 NTU
ISO - NEPH (7027)	0 - 150 FNU
ISO - ABSB	40 - 4000 FAU
IR RATIO	0 - 4000 NTU
EBC	0 - 24.5
ASBC	0 - 236
Accuracy	± 2% of reading plus 0.01 NTU (0 - 500 NTU)
	± 3% of reading (500 - 1000 NTU)
	± 5% of reading (1000 - 2000 NTU)
Resolution	0.01 NTU (0 - 9.99)
	0.1 NTU (10 -99.9)
	1 NTU (100 - 1000)
Repeatability	± 1% of reading or 0.01 NTU
Response Time	< 8 seconds
Calibration	1, 10, 100, 1000 NTU
Signal Averaging	Yes
Sample Cell Size	24 mm
Sample Size	-12 mL
Display	Custom LED
RTC	Yes
Input/Output	RS-232 Serial Port
Power	Battery - four AA's (2,500 hours Alkaline, 10, 000 lithium)
Environmental Conditions	
Operating Temperature	-40° to 140°F (-40° to -60°C)
Humidity	90% RH at 30.0C max
Light Source	White, IR
Warranty	2 years
Weight	8 lbs (3.63 kg)
Safety Rating	UL, CSA, CE, FCC

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6600EDS Extended Deployment System

Measure over 10 parameters in severe fouling environments Featuring Patented Clean Sweep® Anti-fouling Technology

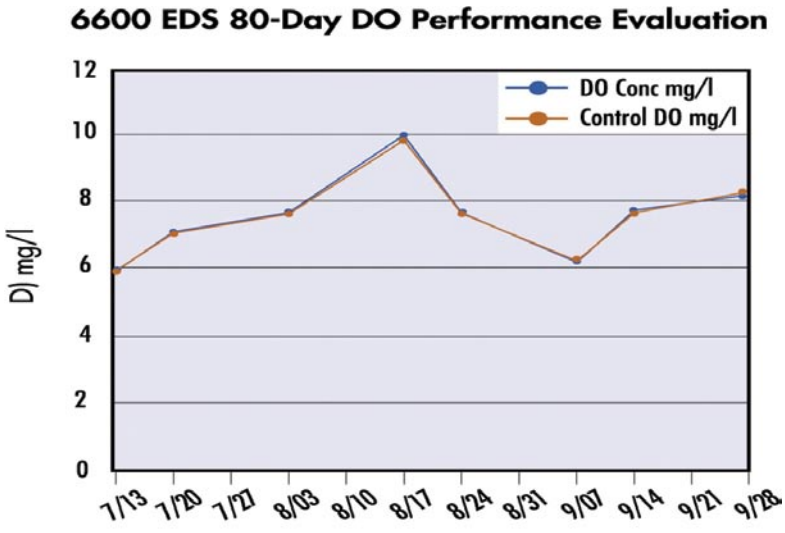
Profile of the 6600EDS depicting (clockwise from bottom) temperature/conductivity, turbidity, Rapid Pulse™ dissolved oxygen, chlorophyll and pH/ORP—all of which (except conductivity) are kept free of fouling by the patented Clean Sweep® universal wiper assembly, as well as individual optical wipers.

Building upon the unprecedented accuracy and reliability of YSI's stirring-independent Rapid Pulse™ dissolved oxygen system, as well as on the improved and proven wiped optical sensors, YSI offers the YSI 6600EDS (Extended Deployment System).

- **Provides unprecedented DO accuracy and longevity in aggressive fouling environments**
- **Patented wiped fouling protection for turbidity, chlorophyll, DO, BGA, pH, and ORP sensors**
- **Ideal for extended, long-term deployments**
- **Virtually maintenance free**
- **Sensors are field-replaceable**
- **Integrates with DCPs (via RS-232 or SDI-12)**

A prototype 6600EDS after continuous deployment for 80 days in Buzzards Bay, MA. The sensor in the foreground is the active DO sensor. The sensor at top-right was used as a non-wiped fouling reference. Note extensive fouling by plant and animal species on the non-wiped sensor.

Initial field studies of the YSI 6600EDS show that the system provides unprecedented DO accuracy and longevity in aggressive fouling environments. The 6600EDS was inspected after 80 days of an ongoing deployment performance evaluation. The Rapid Pulse™ DO sensor performed within specifications throughout this deployment without the need for recalibration or cleaning. During this deployment, the instrument was removed once for battery replacement; none of the sensors was cleaned or recalibrated.



Remarkably close agreement (mean error 0.16mg/l) between the continuously deployed sonde and the control measurements was observed throughout an 80-day deployment.

Pure Data for a Healthy Planet.®

*Sensor Performance verified by the EPA Environmental Technology Verification Program.**

ETV

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ISO 9001
ISO 14001
Yellow Springs, Ohio Facility

Printed in USA 0809 E54-04

CE

*Sensors with listed with the ETV logo were submitted to the ETV program on the YSI 6600EDS. Information on the performance characteristics of YSI water quality sensors can be found at www.epa.gov/etv, or call YSI at 800.897.4151 for the ETV verification report. Use of the ETV name or logo does not imply approval or certification of this product nor does it make any explicit or implied warranties or guarantees as to product performance.

YSI *incorporated*
Who's Minding the Planet?®

Sensor performance verified*

The 6600EDS uses sensor technology that was performance-verified through the US EPA's Environmental Technology Verification Program (ETV). For information on which sensors were performance-verified, look for the ETV logo.

ETV

YSI 6600EDS Sensor Specifications

	Range	Resolution	Accuracy
Dissolved Oxygen• % Saturation ETV 6562 Rapid Pulse™ Sensor*	0 to 500%	0.1%	0 to 200%: ±2% of reading or 2% air saturation, whichever is greater; 200 to 500%: ±6% of reading
Dissolved Oxygen• mg/L ETV 6562 Rapid Pulse™ Sensor*	0 to 50 mg/L	0.01 mg/L	0 to 20 mg/L: ± 0.2 mg/L or 2% of reading, whichever is greater; 20 to 50 mg/L: ±6% of reading
Conductivity•• ETV 6560 Sensor*	0 to 100 mS/cm	0.001 to 0.1 mS/cm (range dependent)	±0.5% of reading + 0.001 mS/cm
Salinity	0 to 70 ppt	0.01 ppt	±1% of reading or 0.1 ppt, whichever is greater
Temperature ETV 6560 Sensor*	-5 to +50°C	0.01°C	±0.15°C
pH ETV 6561 Sensor*	0 to 14 units	0.01 unit	±0.2 unit
ORP	-999 to +999 mV	0.1 mV	±20 mV
Depth — Deep	0 to 656 ft, 200 m	0.001 ft, 0.001 m	±1 ft, ±0.3 m
Depth — Medium	0 to 200 ft, 61 m	0.001 ft, 0.001 m	±0.4 ft, ±0.12 m
Depth — Shallow	0 to 30 ft, 9.1 m	0.001 ft, 0.001 m	±0.06 ft, ±0.02 m
Depth — Vented Level	0 to 30 ft, 9.1 m	0.001 ft, 0.001 m	±0.01 ft, 0.003 m
Turbidity• ETV 6136 Sensor*	0 to 1,000 NTU	0.1 NTU	±2% of reading or 0.3 NTU, whichever is greater**
Rhodamine•	0-200 µg/L	0.1 µg/L	±5% reading or 1 µg/L, whichever is greater

• Maximum depth rating for all standard optical sensors is 200 feet, 61 m. Also available in Deep Depth option: 656 feet, 200 m.
•• Report outputs of specific conductance (conductivity corrected to 25° C), resistivity, and total dissolved solids are also provided. These values are automatically calculated from conductivity according to algorithms found in *Standard Methods for the Examination of Water and Wastewater* (ed 1989).

**In YSI AMCO-AEPA Polymer Standards.

	Range	Detection Limit	Resolution	Linearity
BGA - Phycocyanin•	~0 to 280,000 cells/mL† 0 to 100 RFU	~220 cells/mL§	1 cell/mL 0.1 RFU	$R^2 > 0.9999$**
BGA - Phycoerythrin•	~0 to 200,000 cells/mL† 0 to 100 RFU	~450 cells/mL§§	1 cell/mL 0.1 RFU	$R^2 > 0.9999$***
Chlorophyll• ETV 6025 Sensor*	~0 to 400 µg/L 0 to 100 RFU	~0.1 µg/L§§§	0.1 µg/L Chl 0.1% RFU	$R^2 > 0.9999$****

• Maximum depth rating for all standard optical probes is 200 feet, 61 m. Also available in Deep Depth option 656 ft 200 m.
BGA = Blue-Green Algae
RFU = Relative Fluorescence Units
~ = Approximately

† Explanation of Ranges can be found in the 'Principles of Operation' section of the 6-Series Manual.

§ Estimated from cultures of *Microcystis aeruginosa*.
§§ Estimated from cultures *Synechococcus sp.*
§§§ Determined from cultures of *Isochrysis sp.* and chlorophyll *a* concentration determined via extractions.

**Relative to serial dilution of Rhodamine WT (0-400 ug/L).
***Relative to serial dilution of Rhodamine WT (0-8 µg/L).
****Relative to serial dilution of Rhodamine WT (0-500 ug/L).

YSI 6600EDS Sonde Specifications

Medium		Fresh, sea or polluted water	Software		EcoWatch®
Temperature	Operating	-5 to +50°C	Dimensions	Diameter	3.5 in, 8.9 cm
	Storage	-10 to +60°C		Length, no depth	19.6 in, 34.3 cm
				Length, depth	21.6 in, 54.9 cm
				Weight, depth and batteries	7 lbs, 3.18 kg
Communications		RS-232, SDI-12	Power	External	12 V DC
				Internal	8 C-size alkaline batteries

14.1 Sensor Specifications

Dissolved Oxygen		
Sensor Type		Steady state polarographic
Range:	*% air sat'n*	▪ 0 to 500% air saturation
	mg/L	▪ 0 to 50 mg/L
Accuracy:	*% air sat'n*	▪ 0 to 200% air saturation: ±2% of the reading or 2% air saturation; whichever is greater ▪ 200 to 500% air saturation: ±6% of the reading
	mg/L	▪ 0 to 20 mg/L: ±2% of the reading or 0.2 mg/L; whichever is greater ▪ 20 to 50 mg/L: ±6% of the reading
Resolution:	*% air sat'n*	▪ 0.1% air saturation
	mg/L	▪ 0.01 mg/L
Temperature		
Sensor Type:		YSI Precision™ thermistor
Range:		-5 to 45°C
Accuracy:		±0.15°C
Resolution:		0.01°C
Conductivity		
Sensor Type:		4-electrode cell with auto-ranging
Range:		0 to 200 mS/cm
Accuracy:		±0.5% of reading or ±0.001 mS/cm; whichever is greater–4 meter cable ±1.0% of reading or ±0.001 mS/cm; whichever is greater–20 meter cable
Resolution:		0.001 mS/cm to 0.1 mS/cm (range-dependent)
Salinity		
Sensor Type:		Calculated from conductivity and temperature
Range:		0 to 70 ppt
Accuracy:		±1.0% of reading or 0.1 ppt; whichever is greater
Resolution:		0.01 ppt

YSI 650 Multiparameter Display System

Rugged and Reliable Display and Data Logging System

The YSI 650 Multiparameter Display System

Easily log real-time data, calibrate YSI 6-Series sondes, set up sondes for deployment, and upload data to a PC with the feature-packed YSI 650MDS (Multiparameter Display System). Designed for reliable field use, this versatile display and data logger features a waterproof IP-67, impact-resistant case.

- **Compatible with EcoWatch® for Windows® data analysis software**
- **User-upgradable software from YSI's website**
- **Menu-driven, easy-to-use interface**
- **Multiple language capabilities**
- **Graphing feature**
- **Three-year warranty**

Feature-Packed Performance

Battery Life

With the standard alkaline battery configuration of 4 C-cells, the YSI 650 will power itself and a YSI 6600 sonde continuously for approximately 30 hours. Or, choose the rechargeable battery pack option with quick-charge feature.

Optional Barometer

Temperature-compensated barometer readings are displayed and can be used in dissolved oxygen calibration. Measurements can be logged to memory for tracking changes in barometric pressure.

Optional GPS Interface

Designed to NMEA protocol, the YSI 650 MDS will display and log real-time GPS readings with a user supplied GPS interfaced with YSI 6-Series sondes.

Memory Options

Standard memory with 150 data sets, or a high-memory option (1.5 MB) with more than 50,000 data sets; both options with time and date stamp.

Pure Data for a Healthy Planet.®

A powerful logging display for your data collection processes

The 650MDS can be used with YSI sondes for spot sampling as well as short-term data logging.

Supply a GPS with NMEA 0183 protocol, connect with the YSI 6115 kit, and collect GPS data along with water quality data.

Upload data from the 650 to EcoWatch® for instant data viewing.

YSI 650MDS Specifications

Temperature	Operating Storage	-10 to +60°C for visible display -20 to +70°C
Waterproof Rating		IP-67 for both the standard alkaline battery configuration and for the rechargeable battery pack option
Connector		MS-8; meets IP-67 specification
Dimensions	Width Length Weight with batteries	4.7 in, 11.9 cm 9 in, 22.9 cm 2.1 lbs, 0.91 kg
Display		VGA; LCD with 320 by 240 pixels with backlight
Power	Standard Optional	4 alkaline C-cells with detachable battery cover Ni metal hydride battery pack with attached battery cover and 110/220 volt charging system
Communications		RS-232 to all sondes, for data transfer to PC, and for software updates
Optional GPS		NMEA 0183; requires user-supplied GPS and YSI 6115 Y-cable
Backlight		4 LEDs illuminating LCD; user-selectable
Keypad		20 keys, including instrument on/off, backlight on/off, enter, esc, 10 number/letter entry keys, 2 vertical arrow keys, 2 horizontal arrow keys, period key, and minus key
Warranty		3 years

Ordering Information

650-01	Instrument, standard memory
650-02	Instrument, high memory
650-03	Instrument, standard memory, barometer
650-04	Instrument, high memory, barometer
6113	Rechargeable battery pack kit with 110 volt charger and adapter cable
616	Charger, cigarette lighter
4654	Tripod
614	Ultra clamp, C-clamp mount
5081	Carrying case, hard-sided
5085	Hands-free harness
5065	Form-fitted carrying case
6115	Y-cable for interface with user-supplied GPS system

The 650MDS can interface with any YSI sonde for
- *spot sampling*
- *short-term studies*
- *surface and ground water monitoring*
- *water level monitoring*

Packaged together, the 600QS system includes a 600R conductivity sonde, 650MDS, field cable, and additional sensor options such as pH, dissolved oxygen, ORP, and vented level.

To order, or for more information, contact YSI
+1 937 767 7241
800 897 4151 (US)
www.ysi.com

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YSI *incorporated*
Who's Minding the Planet?®

YSI 600XL and 600XLM Sondes

The YSI 600XL and 600XLM

Measure multiple parameters simultaneously

The YSI 600XL and YSI 600XLM compact sondes measure eleven parameters simultaneously:

Temperature	TDS
Conductivity	pH
Specific Conductance	ORP
Salinity	Depth or Level
Resistivity	Rapid Pulse™ DO (% and mg/L)

Connect with Data Collection Platforms

Either sonde can easily connect to the YSI 6200 DAS (Data Acquisition System), YSI EcoNet™ or your own data collection platform, via SDI-12 for remote and real-time data acquisition applications.

Economical Logging System

The YSI 600XLM is an economical logging system for long-term, *in situ* monitoring and profiling. It will log all parameters at programmable intervals and store 150,000 readings. At one-hour intervals, the instrument will log data for about 75 days utilizing its own power source. The 600XL can also be utilized in the same manner with user-supplied external power.

- **Either sonde fits down 2-inch wells**
- **Horizontal measurements in very shallow waters**
- **Stirring-independent Rapid Pulse® dissolved oxygen sensor**
- **Field-replaceable sensors**
- **Easily connects to data collection platforms**
- **Available with detachable cables to measure depth up to 200 feet**
- **Compatible with YSI 650 Multiparameter Display System**
- **Use with the YSI 5083 flow cell for groundwater applications**

Pure Data for a Healthy Planet.®

Economical, multiparameter sampling or logging in a compact sonde

Sensor performance verified*

The 6820 V2 and 6920 V2 sondes use sensor technology that was verified through the US EPA's Environmental Technology Verification Program (ETV). For information on which sensors were performance-verified, turn this sheet over and look for the ETV logo.

ETV

To order, or for more info, contact YSI Environmental.

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*Sensors with listed with the ETV logo were submitted to the ETV program on the YSI 6600EDS. Information on the performance characteristics of YSI water quality sensors can be found at www.epa.gov/etv, or call YSI at 800.897.4151 for the ETV verification report. Use of the ETV name or logo does not imply approval or certification of this product nor does it make any explicit or implied warranties or guarantees as to product performance.

YSI incorporated
Who's Minding the Planet?®

YSI 600XL & 600XLM Sensor Specifications

	Range	Resolution	Accuracy
Dissolved Oxygen % Saturation ETV 6562 Rapid Pulse™ Sensor*	0 to 500%	0.1%	0 to 200%: ±2% of reading or 2% air saturation, whichever is greater; 200 to 500%: ±6% of reading
Dissolved Oxygen mg/L ETV 6562 Rapid Pulse™ Sensor*	0 to 50 mg/L	0.01 mg/L	0 to 20 mg/L: ± 0.2 mg/L or 2% of reading, whichever is greater; 20 to 50 mg/L: ±6% of reading
Conductivity• ETV 6560 Sensor*	0 to 100 mS/cm	0.001 to 0.1 mS/cm (range dependent)	±0.5% of reading + 0.001 mS/cm
Salinity	0 to 70 ppt	0.01 ppt	±1% of reading or 0.1 ppt, whichever is greater
Temperature ETV 6560 Sensor*	-5 to +50°C	0.01°C	±0.15°C
pH ETV 6561 Sensor*	0 to 14 units	0.01 unit	±0.2 unit
ORP	-999 to +999 mV	0.1 mV	±20 mV
Depth & Level Medium Shallow Vented Level	0 to 200 ft, 61 m 0 to 30 ft, 9.1 m 0 to 30 ft, 9.1 m	0.001 ft, 0.001 m 0.001 ft, 0.001 m 0.001 ft, 0.001 m	±0.4 ft, ±0.12 m ±0.06 ft, ±0.02 m ±0.01 ft, 0.003 m

• Report outputs of specific conductance (conductivity corrected to 25° C), resistivity, and total dissolved solids are also provided. These values are automatically calculated from conductivity according to algorithms found in *Standard Methods for the Examination of Water and Wastewater* (ed 1989).

YSI 600XL & 600XLM Sonde Specifications

Medium		Fresh, sea or polluted water
Temperature	Operating Storage	-5 to +50°C -10 to +60°C
Communications		RS-232, SDI-12
Software		EcoWatch®
Dimensions 600XL | 600XLM	Diameter Length Weight	1.65 in, 4.19 cm | 1.65 in, 4.9 cm 16 in, 40.6 cm | 21.3 in, 54.1 cm 1.3 lbs, 0.59 kg | 1.5 lbs, 0.69 kg
Power	External Internal (600XLM only)	12 V DC 4 AA-size alkaline batteries

YSI model 5083 flow cell and 600XL. This is an ideal combination for groundwater applications.

HI 98129

Combo pH/EC/TDS/Temperature Tester with Low Range EC

Description

The HI 98129 Combo waterproof tester offer high accuracy pH, EC/TDS and temperature measurements in a single tester! No more switching between meters for your routine measurements. The waterproof Combo (it even floats) has a large easy-to-read, dual-level LCD and automatic shut-off. pH and EC/TDS readings are automatically compensated for the effects of temperature (ATC). This technologically advanced tester has a replaceable pH electrode cartridge with an extendable cloth junction as well as an EC/TDS graphite electrode that resists contamination by salts and other substances. This gives these meters a greatly extended life. Your tester no longer needs to be thrown away when the pH sensor is exhausted.

The EC/TDS conversion factor is user selectable as is the temperature compensation coefficient (ß). Fast, efficient, accurate and portable, the Combo pH, EC/TDS and temperature tester brings you all the features you've asked for and more!

Specifications

Range	pH	0.00 to 14.00 pH
Range	EC	0 to 3999 µS/cm
Range	TDS	0 to 2000 ppm
Range T	emperature	0.0 to 60.0°C / 32 to 140.0°F
Resolution pH		0.01 pH
Resolution EC		1 µS/cm
Resolution T	DS	1 ppm
Resolution	Temperature	0.1°C / 0.1°F
Accuracy pH		±0.05 pH
Accuracy EC/T	DS	±2% F.S.
Accuracy	Temperature	±0.5°C / ±1°F
Temperature Compensation		pH: automatic; EC/TDS: automatic with ß adjustable from 0.0 to 2.4% / °C
Calibration	pH	automatic, 1 or 2 points with 2 sets of memorized buffers (pH 4.01 / 7.01 / 10.01 or 4.01 / 6.86 / 9.18)
Calibration	EC/TDS	automatic, 1 point
TDS Conversion Factor		adjustable from 0.45 to 1.00
pH Electrode		HI 73127 (replaceable; included)
Environment		0 to 50°C (32 to 122°F); RH max 100%
Battery Type / Life		4 x 1.5V / approx. 100 hours of continuous use; auto-off after 8 minutes of non-use
Dimensions		163 x 40 x 26 mm (6.4 x 1.6 x 1.0")
Weight		100 g (3.5 oz.)

Appendix H

DIVISION OF WILDLIFE

Ohio Department of Natural Resources

Division of Wildlife Headquarters
2045 Morse Road, Bldg. G
Columbus, Ohio 43229-6693
1-800-WILDLIFE

WILD ANIMAL PERMIT: 12-108

SCIENTIFIC COLLECTION

Vicki J. Mountz
Acting Chief, Division of Wildlife

DATE ISSUED
3/14/2011

Others authorized on permit

YES (SEE ATTACHMENT)

JOHN W. RHOADES
NEORSD
4747 EAST 49TH ST.
CUYAHOGA HEIGHTS, OH 44125-1

SOCIAL SECURITY NUMBER: XXX-XX-7681

is hereby granted permission to take, possess, and transport at any time and in any manner specimens of wild animals, subject to the conditions and restrictions listed below or any documents accompanying this permit.

This permit, unless revoked earlier by the Chief, Division of Wildlife, is effective from: 3/16/2011 to: 3/15/2012

This permit must be carried while collecting wild animals and be exhibited to any person on demand.

THIS PERMIT IS RESTRICTED TO THE FOLLOWING

1. Permittee may collect fish and aquatic macro-invertebrates for survey and inventory purposes. All endangered species are to be released at site of capture.
2. Permittee must consult with Wildlife's Stream Conservation and Environmental Assessment Unit (SCEA) prior to conducting any wild animal work associated with compliance requirements of the Clean Water Act (CWA) Section 401 and/or 404. Contact the unit at 614/265-6346 (John Navarro)) or 614/265-6631 (Becky Jenkins).
3. 24 hours prior to setting trap nets or gillnets, contact must be made with the local wildlife officer or nearest district office to advise location and duration of sampling. All vouchers are to be deposited at NEORSD.
4. Collection is prohibited in Big Darby, Little Darby, Chagrin river and Fish Creek (Williams County) without explicit written permission from the Division of Wildlife.
5. Permittee must provide an annual report of collecting activities to the Division of Wildlife. Report shall provide species, quantity and locations of collection.

Locations of Collecting

STATEWIDE WITH NOTED EXCEPTIONS

Equipment and method used in collection:

SEINES, TRAP NETS AND ELECTROSHOCKER.

Name and number of each species to be collected:

FISH AND MACROINVERTEBRATES AS REQUIRED. ALL FISH (EXCEPT VOUCHER SPECIES) MUST BE RELEASED AT THE COLLECTION SITE.

RESTRICTIVE DOCUMENTS ACCOMPANYING THIS PERMIT? NO

This permit is not valid for collecting migratory birds, their nests, or eggs unless a current permit from the U.S. Fish and Wildlife Service has been obtained.

NO ENDANGERED SPECIES MAY BE TAKEN WITHOUT WRITTEN PERMISSION FROM THE CHIEF

ATTACHMENT

This attachment to Scientific Collecting Permit #12-108 authorizes the following persons to conduct the activities listed on the permit, within the conditions and restrictions set forth. Each person must carry and exhibit upon request, a copy of the permit and this attachment when conducting any of the listed activities. The person named on the permit assumes full responsibility for the actions of the persons on this list and for completing and submitting all required reports.

Name	SSN or Driver License
SETH HOTHEM	XXX-XX-6166
THOMAS ZABLOTNY	XXX-XX-6448
CATHERINE ZAMBORSKY	XXX-XX-6550
FRANCISO RIVERA	XXX-XX-5886
JILLIAN NOVAK	SA294701
KATHRYN CRESTANI	XXX-XX-1565
RON MAICHLE	XXX-XX-8924
KRISTINA GRANLUND	SJ501394
ADDITIONAL ON FILE	

STANDARD CONDITIONS FOR SCIENTIFIC COLLECTING AND EDUCATION PERMITS (ORC 1533.08 AND 1533.09)

The standard conditions listed below apply to all permit holders unless otherwise stated on an issued permit. The standard conditions below are in addition to the provisions listed on the permit. Failure to comply with the conditions of the permit may result in the suspension or termination of your permit. If you need an amendment to your permit, or have questions regarding these conditions, contact the Division of Wildlife Permit Coordinator at (614)265-6315. Please allow a minimum of two weeks for amendments.

1. When collecting or sampling you and any subpermittees must carry a copy of your permit and present to any officer upon request.
2. Only persons listed on the permit may conduct permitted activities.
3. Collection on all Department of Natural Resources properties is prohibited without authorization from the appropriate landholding division.
4. Collection is prohibited in the Little Darby Creek, Big Darby Creek, Killbuck Creek, Fish Creek (Williams County) and the upper portions of the Grand River watershed without written authorization from the Chief.
5. The collection and possession of state endangered and threatened species is prohibited without prior approval from the Chief.
6. The possession of Aquatic Nuisance Species(ANS) for educational or scientific purposes is prohibited without authorization from the Chief.
7. A migratory bird permit issued by the United States Fish and Wildlife Service may be required for all persons collecting or in possession of migratory birds.
8. Twenty-four hours prior to all stream collection, the permit holder must contact the local wildlife officer or nearest district office to advise the location and duration of sampling. Messages are acceptable.
9. All voucher specimens must be ascensioned to the Cleveland Museum of Natural History, The Ohio State University, Museum of Biological Diversity or the Cincinnati Museum of Natural History.
10. Traps and nets must be checked and all animals removed every twenty-four hours.
11. Traps and nets must bear a durable waterproof tag bearing the name and address of the user in English letters, legible at all times.
12. Unless otherwise provided, all specimens must be released at the point of capture.
13. When sampling on public properties or over water, non-toxic shot shall be used.
14. Newly discovered Aquatic Nuisance Species (ANS) must be reported to the Division of Wildlife within twenty-four hours of capture.
15. All Starlings, house sparrows and aquatic nuisance species collected for laboratory use must be euthanized upon completion of project.

Appendix I. References

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Ohio Environmental Protection Agency. (2009a). *Ohio EPA manual of surveillance methods and quality assurance practices*. Columbus, OH: Divisions of Surface Water and Environmental Services.

Ohio Environmental Protection Agency. (2009b). State of Ohio Water Quality Standards *Ohio Administrative Code* Chapter 3745-1 (Revision: Adopted July 9, 2009; Effective October 9, 2009). Columbus, OH: Division of Surface Water, Standards and Technical Support Section.

Appendix J. NEORSD Chlorophyll *a* Sampling Field Sheet

Stream:_____ Collectors:_____
Location:_____ Date:_____
RM:_____ Time:_____
Lat/Long:_____

Number of Rocks:_____ Total Area Scraped:_____cm^2

Diameter of individual scrape	Area of individual scrape
1_____	1_____
2_____	2_____
3_____	3_____
4_____	4_____
5_____	5_____
6_____	6_____
7_____	7_____
8_____	8_____
9_____	9_____
10_____	10_____
11_____	11_____
12_____	12_____
13_____	13_____
14_____	14_____
15_____	15_____
16_____	16_____
17_____	17_____
18_____	18_____
19_____	19_____
20_____	20_____
21_____	21_____
22_____	22_____
23_____	23_____
24_____	24_____
25_____	25_____
	Total:_____

Diameter to Area Conversion

Diameter (cm)	Area (cm2)
1.6	2.011
1.7	2.27
1.8	2.545
1.9	2.835
2.0	3.142
2.1	3.464
2.2	3.801
2.3	4.155

Total Sample Volume_____ml

Filter 1 LABLynx ID_____
Vol _____ml

Filter 2 LABLynx ID_____
Vol _____ml

Filter 3 LABLynx ID_____
Vol _____ml

Water Column Chlorophyll Sample

Filter 1 LABLynx ID_____
Vol _____ml

Filter 2 LABLynx ID_____
Vol _____ml

Filter 3 LABLynx ID_____
Vol _____ml

Flow: None Low Normal Elevated High

Turbidity: Clear Low Moderate* High*

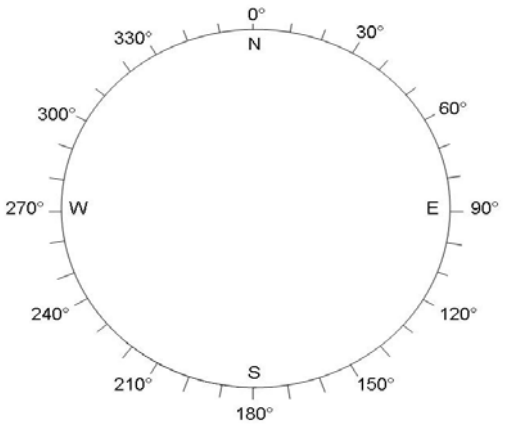
*Explain_____

Sky: Overcast Cloudy Partly Cloudy Mostly Clear Clear

Canopy: Open Mostly Open Partly Closed Closed

Riparian None Narrow L R Moderate L R Wide L R

Downstream Channel Direction



Clinometer

Left Bank _____°
Right Bank _____°

Left Bank _____°
Right Bank _____°

Left Bank _____°
Right Bank _____°

Stream Widths

_____m _____m _____m

Record two most predominate substrates with an X, and check all present.

	Riffle	Run	Reach
Boulder/Slabs	_____	_____	_____
Bedrock	_____	_____	_____
Boulder/Slabs	_____	_____	_____
Cobble	_____	_____	_____
Gravel	_____	_____	_____
Sand	_____	_____	_____
Silt _____		_____	_____
Hardpan	_____	_____	_____
Detritus	_____	_____	_____
Artificial	_____	_____	_____

Substrate Origin

____Limestone ____Tills ____Rip-rap
____Sandstone ____Shale ____Wetlands
____Lacustrine ____Hardpan ____Coal Fines

Silt

____Heavy ____Moderate ____Normal ____None

Embeddedness

____Extensive ____Moderate ____Normal ____None

Notes:_____

_____

Length of Reach: _____m

Stream Drawing