Level 3 Project Study Plan

2015 Mill Creek Environmental Monitoring

(1) Objectives

In 2015, the Northeast Ohio Regional Sewer District (NEORSD) plans to conduct stream monitoring activities at four sites on Mill Creek, an urbanized tributary to the Cuyahoga River. Mill Creek has a natural waterfall, Mill Creek Falls, which is a fish migration barrier at river mile (RM) 2.80. NEORSD will assess habitat and water chemistry conditions and evaluate the fish and benthic macroinvertebrate communities at each site. The purpose of the 2015 monitoring is to continue the biology of the creek downstream of the falls and to satisfy permit requirements. The four sites, which are along Mill Creek's main branch, are located at RMs 8.30, 2.75, 0.70, and 0.12. These sites were first surveyed in 1995 as part of the Mill Creek Watershed Management Project, and were all surveyed again in 2011 through 2014.

A comparison of the fish and macroinvertebrate communities and the corresponding habitat and water chemistry data will be used to determine the the biological communities and the overall health of the creek segments. Additionally, sampling at RM 0.12 and 8.30 are required by Ohio Environmental Protection Agency (EPA) National Pollutant Discharge Elimination System (NPDES) Permit No. 3PA00002*GD.

The 2015 survey will continue to support several NEORSD capital improvement projects designed to provide wet weather flow relief, stormwater storage capacity, and reduction/elimination of CSOs for several communities in the Mill Creek watershed. The Miles Avenue Relief Sewer (MARS) was completed in June 2010, the Lee Road Relief Sewer (LRRS) was completed in June 2012, and the Phase Three of the Mill Creek Tunnel Project (MCT-3C) was completed in late 2012. The stream monitoring surveys, which are considered post-construction monitoring for LRRS, MARS and MCT-3C, will enable future evaluations of the effectiveness of the capital improvement projects in restoring the chemical and biological health of Mill Creek.

Stream monitoring activities will be conducted at each site by NEORSD Level 3 Qualified Data Collectors certified by Ohio EPA in Fish Community Biology, Benthic Macroinvertebrate Biology, Chemical Water Quality, and Stream Habitat Assessment. Fish and macroinvertebrate community health will be evaluated through the use of appropriate Ohio EPA methods, which may include: Index of Biotic Integrity (IBI); and the Invertebrate Community Index (ICI). An examination of the specific characteristics of the biological communities will be used in conjunction with water quality data, the NEORSD Macroinvertebrate Field Sheet, and Qualitative Habitat Evaluation Index (QHEI) results in order to identify

impacts to the communities. Results will be compared to historic data to show temporal as well as spatial trends. Water chemistry data will also be compared to the Ohio Water Quality Standards to determine attainment of applicable uses (Ohio EPA, 2013a)[1].

(2) Nonpoint/Point Sources

Point Sources	Nonpoint Sources
Combined Sewer Overflows	Urban Runoff
Sanitary Sewer Overflows	Spills
Storm Sewer Outfalls	Sedimentation
Home septic systems	

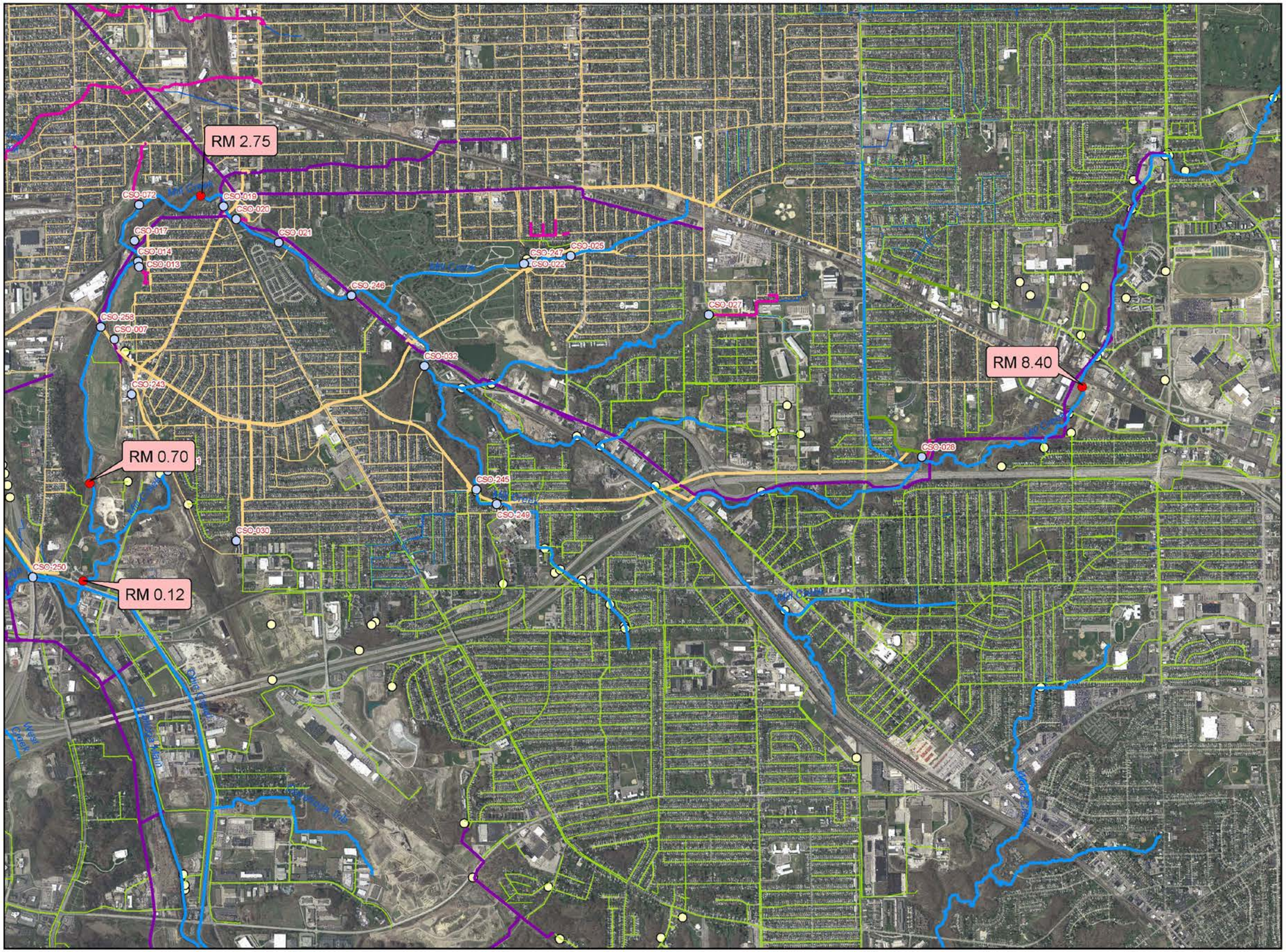
A map has been provided to show point sources that may be influencing the water quality at each sample location. These sources, along with the nonpoint sources listed in the table above, may be impacting the health of the fish and benthic macroinvertebrate communities in Mill Creek. Other factors that may influence ecological conditions during the study include periods of drought or precipitation.

(6) Sampling Locations

The following water chemistry, habitat, electrofishing, and macroinvertebrate sample locations on Mill Creek, listed from upstream to downstream, will be surveyed during the 2015 field season. Benthic macroinvertebrate and water chemistry collection sites are located within each electrofishing zone, indicated by RM. GPS coordinates are recorded at the downstream end of each electrofishing zone.

[1] See appendix H for a list of all references.

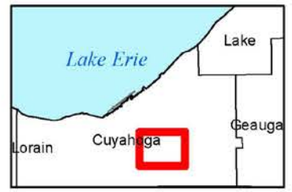
Water Body	Latitude	Longitude	River Mile	Location Information	USGS HUC 8 Number - Name	Purpose
Mill Creek	41.4305	-81.5442	8.30	Upstream of South Miles Road, first site upstream of NEORSD CSOs	04110002 Cuyahoga	Evaluate watershed health, support of Capital Improvement projects. Site required by Ohio EPA NPDES Permit
Mill Creek	41.4451	-81.6271	2.75	Downstream of the Mill Creek Falls	04110002 Cuyahoga	Evaluate watershed health, support of Capital Improvement projects.
Mill Creek	41.4240	-81.6376	0.70	Upstream of the Warner Road Tributary, adjacent to 5000 Warner Road	04110002 Cuyahoga	Evaluate watershed health, support of Capital Improvement projects.
Mill Creek	41.4178	-81.6387	0.12	Upstream of Canal Road	04110002 Cuyahoga	Evaluate watershed health, support of Capital Improvement projects. Site required by Ohio EPA NPDES Permit



Northeast Ohio Regional Sewer District
Protecting Your Health and Environment

Mill Creek Study Plan

Overview Map



Legend

- Monitoring Site
- Regional Drainage
- CSO Outfall
- District Facility
- Outfalls
- NEORSD CSO Combined Sewer
- NEORSD CSO Responsibility Sewer
- NEORSD Intercommunity Relief Sewer
- NEORSD INTERCEPTOR
- Local Combined Sewer
- Local Culverted Stream
- Local Sanitary Sewer
- Local Storm Sewer

N

0.45

Miles

(3) Parameters Covered

Fish specimens will be identified to species level, weighed, counted and examined for the presence of external anomalies including DELTs (deformities, eroded fins, lesions and tumors). An Ohio EPA Fish Data Sheet (Appendix A) will be completed during each assessment. Quantitative fish sampling is expected to be conducted at all locations.

Macroinvertebrate community assemblages will be collected from each location. Third Rock Consultants, LLC will identify and enumerate the specimens collected from each site. All specimens will be identified to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b) [1]. The NEORSD Macroinvertebrate Field Sheet (Appendix A) will be completed at each site during sampler retrieval or when qualitative sampling is conducted.

Stream habitat will be measured by scoring components of the QHEI at all locations, including the substrate, instream cover, channel morphology, riparian zone, bank erosion, pool/glide and riffle/run quality and gradient. The HHEI will be conducted at those sites with drainage areas less than one square mile listed under PSPs with general watershed monitoring. The Lacustuary QHEI (L-QHEI) will be performed at sites that are affected by the water level of Lake Erie. Examples of the Ohio EPA field sheets for the QHEI, L-QHEI and the HHEI can be found in Appendix A.

Water chemistry samples will be collected at each electrofishing/ macroinvertebrate sampling site included in the study. Water chemistry samples will be analyzed by NEORSD's Analytical Services Division. Appendix B lists the parameters to be tested along with the detection limits and practical quantitation limits. Field measurements for dissolved oxygen, pH, temperature, conductivity and turbidity will also be performed. A Surface Water Condition Sampling Field Data Form will be completed at each site during each sampling event (Appendix A).

Benthic and water column chlorophyll *a* samples may be collected from stream locations. Chemical and physical water quality parameters to be measured in conjunction with the chlorophyll *a* samples include total phosphorus, dissolved reactive phosphorus, nitrite, nitrate+nitrite, ammonia, alkalinity, turbidity and suspended solids. In the Cuyahoga River, YSI 6600EDS data sondes may be

[1] See Appendix H for a list of all references.

installed at RMs 16.20, 10.75, 10.10, and 7.00 around the time that this sampling is conducted to more frequently monitor dissolved oxygen, temperature, conductivity, specific conductivity and pH.

(4) Field Collection and Data Assessment Techniques

Field collections for fish will be conducted at all stream locations. Sampling will be conducted using longline, backpack, or boat electrofishing techniques and will consist of shocking all habitat types within a sampling zone. Headwater and wading sites, which are 0.15 and 0.20 kilometers in length, respectively, will be surveyed by moving from downstream to upstream. Boat sites, which are 0.50 kilometers in length, will be surveyed by moving from upstream to downstream. The stunned fish will be collected and placed into a live well for later identification. The longline, backpack, and boat electrofishing zones will be assessed one to three times during the field season (June 15 - October 15).

Fish will be identified to the species level, weighed, counted, and examined for the presence of external anomalies including DELTs. Fish easily identified (commonly collected from year to year) will be returned to the site from which they are collected. Fish species difficult to identify will be brought back to the laboratory for verification by NEORSD Level 3 Fish Qualified Data Collectors (QDC). If necessary, vouchers will be sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Voucher specimens will be collected as described in section (14). Endangered species and those too large for preservation will not be collected as voucher specimens, but will instead be photographed. Photographed vouchers will include features that permit definitive identification of the particular species.

Fish will be preserved in 10 percent formalin in the field, soaked in tap water for 24 to 48 hours after 5 to 7 days, then transferred to solutions of 30 and 50 percent ethanol for 5 to 7 days each and, finally, to 70 percent ethanol for long-term storage. Specimens larger than six inches will be slit along the right side and then soaked in formalin for approximately 10 to 14 days before being transferred to water and solutions of 30, 50 and 70 percent ethanol. Label information will include location (description and coordinates), date, time, collectors' names and sample identification code for each specimen collected.

Macroinvertebrate sampling will be conducted using quantitative and qualitative sampling techniques. Quantitative sampling will be done using a modified Hester-Dendy multi-plate artificial substrate sampler (HD) that is colonized for a six-week period. Multiple HD samplers may be installed at one or all sampling locations in case samplers are lost due to vandalism, burial, etc. and for the purposes of providing a replicate sample. Qualitative sampling will be conducted using a D-frame dip net when HD samplers are retrieved. The NEORSD

Macroinvertebrate Field Sheet will be completed during each HD retrieval. Ronald Maichle of NEORSD, a Level 3 QDC for Benthic Macroinvertebrate Biology, may identify specimens in the replicate sample to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b).

Macroinvertebrate voucher specimens for both quantitative and qualitative sampling will be collected as described in section (14). Macroinvertebrate community assemblages collected will be shipped to Third Rock Consultants, LLC (Lexington, KY) for identification and enumeration. Third Rock Consultants, LLC will identify specimens to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b).

A detailed description of the sampling and analysis methods utilized in the fish community and macroinvertebrate surveys, including calculations of the IBI, MIwb, and ICI, can be found in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volumes II* (1987a) and *III* (1987b). Methods for assessing fish and macroinvertebrate communities in lacustuary zones can be found in Ohio EPA's draft *Biological Criteria for the Protection of Aquatic Life, Volume IV* (1997).

The QHEI, as described in Ohio EPA's *Methods for Assessing Habitat in Flowing Waters: Using the Qualitative Habitat Evaluation Index (QHEI)* (2006) will be used to assess aquatic habitat conditions at each sample location. The L-QHEI will be used where appropriate and will follow Ohio EPA's draft *Methods of Assessing Habitat in Lake Erie Shoreline Waters Using the Qualitative Habitat Evaluation Index (QHEI) Approach (Version 2.1)* (2010).

The HHEI as described in Ohio EPA's *Field Evaluation Manual for Ohio's Primary Headwater Habitat Streams* (2012a) will be used, when necessary, to conduct use attainability analyses and to classify the actual and expected biological conditions in primary headwater habitat streams.

Water chemistry sampling may occur across a variety of flow conditions. Techniques used for water chemistry sampling and chemical analyses will follow the *Surface Water Field Sampling Manual* (Ohio EPA, 2013). Chemical water quality samples from each site will be collected with at least one 4-liter disposable polyethylene cubitainers with disposable polypropylene lids and two 473-mL plastic bottles. Water samples collected for analysis of dissolved reactive phosphorus will be filtered using a 0.45-µm PVDF syringe filter and will be collected in a 125-mL plastic bottle. Bacteriological samples will be collected in a sterile plastic bottle preserved with sodium thiosulfate. All water quality samples

will be collected as grab samples. Field blanks and duplicate samples will each comprise not less than 5% of the total samples collected for this study plan, for a total frequency of quality control samples of not less than 10% of the total samples collected. With the exception of bacteriological duplicate samples, the acceptable percent RPD will be based on the ratio of the sample concentration and detection limit (Ohio EPA, 2013): Acceptable % RPD = $[(0.9465X^{-0.344})*100] + 5$, where X = sample/detection limit ratio. For bacteriological duplicates, duplicate samples more than 5x apart from one another (%RPD > 133.3%) will be rejected in accordance with the Ohio EPA approved method for data validation of bacteriological samples outlined in Section F of the *Ohio 2012 Integrated Water Quality Monitoring and Assessment Report* (Ohio EPA, 2012b). Those RPDs that were higher than acceptable may indicate potential problems with sample collection and, as a result, the data will not be used for comparison to the water quality standards. Acid preservation of the samples, as specified in the NEORSD laboratory's standard operating procedure for each parameter, will occur in the field. Appendix B lists the analytical method, method detection limit and practical quantitation limit for each parameter analyzed. Field analyses include the use of either a YSI-556 MPS Multi-Parameter Water Quality Meter, YSI EXO1 sonde, or YSI 600XL sonde to measure dissolved oxygen (DO), water temperature, conductivity and pH; and when necessary, a Hanna HI 98129 meter to measure pH and a Hach HQ30d meter with LDO101 probe to measure DO. Field turbidity will be measured using either a Hach 2100P Portable Turbidimeter or Hach 2100Q Turbidimeter. Specifications for these meters have been included in Appendix C.

Benthic and water column chlorophyll *a* samples may be collected if time and resources allow. Sampling methods will follow those detailed in the NEORSD *Chlorophyll a Sampling and Field Filtering Standard Operating Procedure* (SOP-EA001-00). A Chlorophyll *a* Sampling Field Sheet will be completed for each site (Appendix D). Water chemistry grab samples will be collected at the same time using the methods discussed previously and will be analyzed for nutrients, turbidity, alkalinity and suspended solids. Additionally in the Cuyahoga River, approximately 24-hours prior to each chlorophyll *a* sampling event, YSI 6600 EDS data sondes may be deployed at RMs 16.20, 10.75, 10.10 and 7.00. If installed, each data sonde will record, at fifteen-minute intervals, dissolved oxygen concentration, pH, temperature, and conductivity from the time the data sonde is deployed until the time it is retrieved. These data sondes will be placed in the stream by inserting each one into a 4.5-inch PVC pipe with holes drilled into the sides of the lower third of the pipe to allow water to pass through it. The data sondes will remain in the river for approximately 24-hours or longer following collection of the chlorophyll *a* samples.

Where possible, data assessment will include an analysis of temporal and spatial trends in the collected data. Species assemblages and individual metrics will be

analyzed. Graphs that show current and historic QHEI, L-QHEI, IBI, LIBI, MIwb, ICI, and LICI scores and how these scores compare to attainment status of biocriteria will be prepared. Water chemistry data collected will be compared to Ohio water quality standards to determine whether any excursions from the applicable water quality criteria have occurred. It will also be used to determine any relationships among individual parameters and chlorophyll *a* concentrations. Comparisons between water quality and biological community health will only be made if at least three water quality samples have been collected from that site.

(5) Stream Flow Measurement

Stream flow will be recorded for all locations during each electrofishing pass utilizing data from the United States Geological Survey (USGS) gauge station nearest the stream location, if applicable.

Stream flow will be measured with a Marsh-McBirney FloMate Model 2000 Portable Flow Meter, a HACH FH950 Flow Meter or an Aquaflow Probe Model 6900, which measure flow in feet per second, when HD samplers are installed and retrieved. The specifications for the flow meters can be found in Appendix C.

(7) Schedule

One to three electrofishing surveys will be conducted at each site between June 15 and October 15, 2015. Surveys will be conducted at least three weeks apart. Specific dates have not been scheduled. River flow and weather conditions will be assessed weekly to determine when each electrofishing pass will be conducted.

Artificial substrate samplers will be installed at stream locations between June 15 and August 19, 2015, and retrieved six weeks later. Qualitative macroinvertebrate sampling will be conducted one time at all sites. Specific dates have not been scheduled. River flow and weather conditions will be assessed weekly to determine when the HD sampler installations and retrievals and qualitative sampling will be conducted.

QHEI, and if necessary, HHEI and L-QHEI habitat evaluations will be conducted one time between June 15 and October 15, 2015. QHEI evaluations will be conducted around the same time as one of the electrofishing surveys.

Water chemistry samples will be collected a minimum of three times from stream locations between June 15 and October 15, 2015.

Benthic and water column chlorophyll *a* samples may be collected at least one time from stream locations between June 15 and October 15, 2015. These samples will be collected under low-flow conditions.

(8) QA/QC

Quality assurance and quality control of sampling and analysis methods for habitat, fish, and macroinvertebrate evaluations will follow Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volumes II* (1987a) and *III* (1987b), *Methods for Assessing Habitat in Flowing Waters: Using the Qualitative Habitat Evaluation Index (QHEI)* (2006), *Field Evaluation Manual for Ohio's Primary Headwater Habitat Streams* (2012a), draft *Biological Criteria for the Protection of Aquatic Life: Volume IV: Fish and Macroinvertebrate Indicies for Ohio's Lake Erie Nearshore Waters, Harbors, and Lacustuaries* (1997) and draft *Methods of Assessing Habitat in Lake Erie Shoreline Waters Using the Qualitative Habitat Evaluation Index (QHEI) Approach (Version 2.1)* (2010)

Electrofishing equipment will be used according to the guidelines listed in the operation and maintenance manual provided by Smith-Root, Inc. Malfunctioning equipment will not be used to collect data. Proper steps will be taken to correct any problems as soon as possible, whether by repairing in the field, at the NEORSD Environmental & Maintenance Services Center, or by contacting the supplier or an appropriate service company.

Fish species difficult to identify will be brought back to the laboratory for verification by Level 3 Fish QDC's, and if necessary, sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Voucher specimens will be collected as described in section (14). Endangered species and those too large for preservation will not be collected as voucher specimens, but will instead be photographed. Photographed vouchers will include features that permit definitive identification of the particular species.

All macroinvertebrate community assemblages from stream locations, except for the replicate sample, will be collected and shipped to Third Rock Consultants, LLC for identification and enumeration. All specimens will be identified to the lowest practical taxonomic level as recommended in Ohio EPA's *Biological Criteria for the Protection of Aquatic Life, Volume III* (1987b). All macroinvertebrate specimens will be returned to NEORSD. At least two voucher specimens of each species, when available, will be separated into individual vials and kept as described in section (14). The remaining specimens for each site will be returned in a single container labeled with the site number and collection method and date. All specimens and accompanying chain-of-custody documentation will be retained by NEORSD and stored at the Environmental & Maintenance Services Center for a period not less than ten years.

Water samples obtained for chemical analyses will be collected, preserved (see Section 4), labeled and then placed on ice inside the field truck. The field truck will remain locked at all times when not occupied/visible. Sampling activities, including sample time and condition of surface water sampled, will be entered in a field log book and on the Surface Water Condition Sampling Field Data Form. The samples will then be delivered immediately to the NEORSD Analytical Services cooler, after which the door to the cooler will be locked, and the samples will be transferred to the custody of Analytical Services. The NEORSD Analytical Services Quality Manual and associated Standard Operating Procedures are on file with Ohio EPA. The Quality Assurance Officer at Analytical Services will send updates, revisions and any information on document control to Ohio EPA as needed.

For benthic and water column chlorophyll *a* sampling, three filtrations will be performed for each sample. A field filtration blank will be submitted for every 20 samples.

Calibration of YSI 6600EDS data sondes will be done according to the YSI Environmental Operations Manual. The conductivity will be calibrated first using a 1.413 mS/cm standard. Second, the pH will be calibrated using two different buffers (7 and 10 s.u.). The DO will be calibrated last with an acceptable error of 0.2 mg/L.

Once the sondes are removed from the river, the accuracy of the data that has been collected will be checked by comparing readings taken by the sondes to known standards. If the measurements taken at this time meet quality control goals, all of the data collected since the last calibration will be considered accurate. The acceptable differences for pH and conductivity will be $\pm$0.3 with pH 7 buffer and $\pm$ 10% of the conductivity standard, respectively (EPA New England- Region 1, 2005). The acceptable difference for DO will be $\pm$0.2 mg/L. If the measurements do not meet quality control goals, best professional judgment will be used to decide if any of the data collected during that period may still be accurate. For example, the data collected from the four locations may be plotted on the same graph, and if it appears that the data points are following similar trends, they may be considered accurate. If any data that do not meet quality control goals are used, a rationale for their inclusion will be provided when the data are submitted.

(9) Work Products

Within one year of completion of the project, fish data (species, numbers, weights, pollution tolerances, the incidence of DELT anomalies, IBI or LIBI, MIwb scores), macroinvertebrate data (types and numbers of macroinvertebrates collected and ICI or LICI scores), habitat data (QHEI or L-QHEI raw data and scores) and water chemistry results will be submitted to the Ohio EPA or an Ohio

EPA approved data warehouse. Additionally, reports summarizing, interpreting, graphically presenting and discussing the IBI (LIBI, where applicable), MIwb, ICI (LICI, where applicable) and QHEI (L-QHEI, where applicable) scores, chlorophyll *a* results, and any excursions from water quality standards may be prepared for internal use.

(10) Qualified Data Collectors

The following Level 3 Qualified Data Collectors (QDC) will be involved with this study:

Name	Address	Email Address	Phone Number	QDC Specialty(s)
John W. Rhoades[1]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	rhoadesj@neorsd.org	216-641-6000	QDC - 00008 CWQA/FCB/SHA/ BMB
Cathy Zamborsky	4747 East 49th Street Cuyahoga Hts., Ohio 44125	zamborskyc@neorsd.org	216-641-6000	QDC - 00009 CWQA/SHA
Seth Hothem	4747 East 49th Street Cuyahoga Hts., Ohio 44125	hothems@neorsd.org	216-641-6000	QDC - 00010 CWQA/FCB/SHA/ BMB
Tom Zablotny	4747 East 49th Street Cuyahoga Hts., Ohio 44125	zablotnyt@neorsd.org	216-641-6000	QDC - 00018 CWQA/FCB/SHA
Ron Maichle	4747 East 49th Street Cuyahoga Hts., Ohio 44125	maichler@neorsd.org	216-641-6000	QDC - 00145 CWQA/SHA/BMB
Francisco Rivera	4747 East 49th Street Cuyahoga Hts., Ohio 44125	riveraf@neorsd.org	216-641-6000	QDC - 00262 CWQA/SHA
Jillian Knittle	4747 East 49th Street Cuyahoga Hts., Ohio 44125	knittlej@neorsd.org	216-641-6000	QDC – 00512 CWQA/SHA/BMB
Jonathan Brauer[2]	4747 East 49th Street Cuyahoga Hts., Ohio 44125	brauerj@neorsd.org	216-641-6000	QDC – 00663 SHA
Bert Remley[3]	2526 Regency Road, Suite 180 Lexington, Kentucky 40503	bremley@thirdrockconsultants.com	859-977-2000	QDC – 00837 BMB

[1] NEORSD Lead Project Manager
[2] See acknowledgement letter for conducting QHEIs (Appendix F)
[3] Benthic Macroinvertebrate Identification

The following is a list of persons not qualified as Level 3 QDCs who may be involved in the project. Prior to the start of sampling, the project managers will explain to each individual the proper methods for sampling. Sampling will only be completed under the direct observation of a QDC. The lead project manager will be responsible for reviewing all reports and data analysis prepared by qualified personnel prior to completion.

Name	Address	Email Address	Phone Number
Kelsey Amidon	4747 East 49th Street Cuyahoga Hts., Ohio 44125	amidonk@neorsd.org	216-641-6000
Nick Barille	4747 East 49th Street	barillen@neorsd.org	216-641-6000

Name	Address	Email Address	Phone Number
	Cuyahoga Hts., Ohio 44125		
Mark Colvin	4747 East 49th Street Cuyahoga Hts., Ohio 44125	colvinm@neorsd.org	216-641-6000
Tim Dobriansky	4747 East 49th Street Cuyahoga Hts., Ohio 44125	dobrianskyt@neorsd.org	216-641-6000
Donna Friedman	4747 East 49th Street Cuyahoga Hts., Ohio 44125	friedmand@neorsd.org	216-641-6000
Rae Grant	4747 East 49th Street Cuyahoga Hts., Ohio 44125	grantr@neorsd.org	216-641-6000
Mark Matteson	4747 East 49th Street Cuyahoga Hts., Ohio 44125	mattesonm@neorsd.org	216-641-6000
Mario Meany	4747 East 49th Street Cuyahoga Hts., Ohio 44125	meanym@neorsd.org	216-641-6000
Carrie Millward	4747 East 49th Street Cuyahoga Hts., Ohio 44125	millwardc@neorsd.org	216-641-6000
Denise Phillips	4747 East 49th Street Cuyahoga Hts., Ohio 44125	phillipsd@neorsd.org	216-641-6000
Brandy Reischman	4747 East 49th Street Cuyahoga Hts., Ohio 44125	reischmanb@neorsd.org	216-641-6000
Frank Schuschu	4747 East 49th Street Cuyahoga Hts., Ohio 44125	schuschuf@neorsd.org	216-641-6000
Eric Soehnlen	4747 East 49th Street Cuyahoga Hts., Ohio 44125	soehnlene@neorsd.org	216-641-6000
William Stanford	4747 East 49th Street Cuyahoga Hts., Ohio 44125	standfordw@neorsd.org	216-641-6000
Wolfram von Kiparski	4747 East 49th Street Cuyahoga Hts., Ohio 44125	vonkiparskiw@neorsd.org	216-641-6000
NEORSD Summer Co-op #1	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000
NEORSD Summer Co-op #2	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000
NEORSD Summer Co-op #3	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000
NEORSD Summer Co-op #4	4747 East 49th Street Cuyahoga Hts., Ohio 44125	To Be Determined	216-641-6000

(11) Contract laboratory contact information

All bacteriological and/or chemical sample analysis will be completed by NEORSD's Analytical Services Division. Evidence of NEORSD's Analytical Services current accreditation and method dates can be found in Appendix E. The contact information for NEORSD's Analytical Service Division is:

NEORSD Analytical Services
Mr. Mark Citriglia
4747 E. 49th Street
Cuyahoga Heights, Ohio 44056
citrigliam@neorsd.org
216-641-6000

Any fish that is not positively identified in the field, or at NEORSD, will be sent to The Ohio State University Museum of Biological Diversity for verification by the Curator and/or Associate Curator of Fish. Fish will be identified to the species level.

Dr. Ted Cavender, Curator of Fish / Mr. Marc Kibbey, Associate Curator of Fish
1315 Kinnear Road, Columbus, Ohio 43212
cavender.1@osu.edu / kibbey.3@osu.edu
614-292-7873

Identification of macroinvertebrates for stream locations will be completed by Third Rock Consultants LLC (Lexington, Kentucky) (Appendix F). Benthic macroinvertebrates will be identified to the lowest practical level as recommended by Ohio EPA (1987b). Third Rock Consultants LLC contact information:

Ms. Marcia Wooton
Third Rock Consultants LLC
2526 Regency Road, Suite 180
Lexington, Kentucky 40503
mwooton@thirdrockconsultants.com
859-977-2000

(12) Copy of ODNR collector's permit

See Appendix G.

(13) Digital Catalog Statement

A digital photo catalog of all sampling locations will be maintained for 10 years and will include photos of the specific sampling location(s), the riparian zone adjacent to the sampling location(s) and the general land use in the immediate vicinity of the sampling location(s).

Print/Signature: John W. Rhoades Date: 04/13/15

(14) Voucher Specimen Statement

NEORSD will maintain a benthic macroinvertebrate and fish voucher collection which includes two specimens, or appropriate photo vouchers, of each species or taxa collected during the course of biological sampling from any stream within the NEORSD's service area. When benthic macroinvertebrates from multiple surface waters are collected within the same year and identified by the same QDC, one voucher collection will be created to represent the specimens collected from those

streams. When fish specimens from multiple surface waters are collected within the same year, one voucher collection will be created to represent the specimens collected from those streams. A separate collection for each sampling event will not be maintained.

NEORSD will provide specimens or photo vouchers to the Director upon request. This collection will be stored at the NEORSD laboratory in the Environmental and Maintenance Services Center.

Print/Signature: John W. Rhoades Date: 04/13/15

(15) Sample Location Statement

I attest that I will make available any and all sampling location information, including but not limited to; the name of the water body sampled, sampling location latitude and longitude, sampling location river mile where possible, general location information, the U.S. geological survey HUC 8 number and name, and the purpose for data collection at each sampling location.

Print/Signature: John W. Rhoades Date: 04/13/15

(16) Additional L3 Data Collector Statement

The Lead Project Manager for all stream locations is approved for all project data types.

Print/Signature: John W. Rhoades Date: 04/13/15

(17) Trespassing Statement

I have not been convicted or pleaded guilty to a Violation of section 2911.21 of the Revised Code (criminal trespass) or a substantially similar municipal ordinance within the previous five years.

Print/Signature: John W. Rhoades / _____ Date: 04/13/15

Print/Signature: Cathy Zamborsky / _____ Date: 4/10/15

Print/Signature: Seth Hothem / _____ Date: 4/10/15

Print/Signature: Tom Zablotny / _____ Date: 4 10 15

Print/Signature: Ron Maichle / _____ Date: 04-10-15

Print/Signature: Jillian Knittle / _____ Date: 4/13/15

Print/Signature: Francisco Rivera / _____ Date: 4/13/15

Print/Signature: Jonathan Brauer / _____ Date: 4/10/15

Appendix A

FISH DATA SHEET

Sheet ID For Office Use Only

New Station (requires lat/long & county) ☐ Mix Zone ☐ Page____of____

Station ID_____ River Code_____ RM_____ Date_____ Time_____

Stream_____ Location _____

Comments _____

Lat _____ Long _____ County _____ ALP _____ Time Fished _____

Crew _____ Netter _____ Others _____ Sampler Type _____

Distance _____ Flow _____ Temp. C _____ Secchi _____ Source _____ Project _____

	Fins Code	Number Weighed	Total Counted	Total Weight	Weights	Counts	DELT ANOMALIES (Deformities, Erosions, Lesions, Tumors; Multiple DELTs on one fish) D	E	L	T	M	*
1	V 10x											
2	V 10x											
3	V 10x											
4	V 10x											
5	V 10x											
6	V 10x											
7	V 10x											
8	V 10x											
9	V 10x											

* A-anchor worm; B-black spot; C-leeches; F-fungus; N-blind; P-parasites; S-emaciated, W-swirled scales Y-popeye; Z-other

	Fins Code		Number Weighed	Total Counted	Total Weight	Weights	Counts		D	E	L	T	M	*
10														
	V	10x												
11														
	V	10x												
12														
	V	10x												
13														
	V	10x												
14														
	V	10x												
15														
	V	10x												
16														
	V	10x												
17														
	V	10x												
18														
	V	10x												
19														
	V	10x												
20														
	V	10x												
21														
	V	10x												

Page _____ of _____

NEORSD Macroinvertebrate Field Sheet

Stream: _____ River Mile: _____ Year: _____

Location: _____ Project: _____

Drainage Area (mi^2): _____ Latitude (°N)/Longitude (°W): _____

Hester-Dendy Deployment Information

Install Date: _____ Crew (QDC Circled): _____

Current at HD (fps): _____ Depth (cm): _____ Pictures Obtained: Yes No

Reinstall Date: _____ Crew (QDC Circled): _____

Current (fps): _____ Depth (cm): _____ Reason: _____

Reinstall Date: _____ Crew (QDC Circled): _____

Current (fps): _____ Depth (cm): _____ Reason: _____

Sampling/Retrieval Information

Sampling Method: Hester-Dendy Dipnet Surber Core Other: _____

Sample ID: HD: _____ Qualitative: _____ Other: _____

Sampling Date: _____ Crew (QDC Circled): _____

HD Condition- Current (fps): _____ Depth (cm): _____ Water Temp: _____ °F / °C

Number of HD Blocks Obtained: _____ Remarks: _____

Disturbed: Yes No Comments: _____

Debris: Yes No Comments: _____

Silt/Solids: None Slight Moderate Heavy

Dipnet- Time Sampled (min): _____ X Number of Crew: _____ = Total (min): _____

Habitats Sampled: Pool Riffle Run Margin Backwater

River Sampling Conditions

Flow Condition:	Flood	Above Normal	Normal	Low	Interstitial	Intermittent	Dry
Current Velocity:	Fast	Moderate	Slow	Non-detect			
Channel Morphology:	Natural	Channelized	Channelized (Recovered)		Impounded		
Bank Erosion:	Extensive	Moderate	Slight	None			
Riffle Development:	Extensive	Moderate	Sparse	Absent			
Riffle Quality:	Good	Fair	Poor		*Embedded:*	Yes	No
Water Clarity:	Clear	Murky	Turbid		Other: _____		
Water Color:	None	Green	Brown	Grey	Other: _____		
Canopy over HD:	Open	75 %	50 %	25 %	Closed		

Comment Section: _____

_____

_____

_____

_____

OEPA Comment Field Codes: _____

Samples Analyzed By: _____ QDC #: _____ Date: _____

Physical Characteristics

Substrate Characteristics

	Pool	Units	Riffle	Units	Run	Units
Bedrock						
Boulder						
Rubble						
Coarse Gravel						
Fine Gravel						
Sand						
Silt						
Clay/Hardpan						
Detritus						
Peat						
Muck						
Other						
Macrophytes						
Algae						
Artifacts						
Compaction (F,M,S)						
Depth (Avg)						
Width (Avg)						

Predominant Land Use (Left, Right or Both)

Forest	Urban	Open Pasture
Shrub	Residential/Park	Closed Pasture
Old Field	Mining/Construction	
Rowcrop	Wetland	
Industrial	Other	

Predominant Riparian Vegetation

Left	Right	Type
_____	_____	Large Trees
_____	_____	Small Trees
_____	_____	Shrubs
_____	_____	Grass/Weeds
_____	_____	None

Margin Habitat

Margin Quality: Good Fair Poor

Undercut Banks	Root Mats	Tree Roots
Grass	Water Willow	Woody Debris
Shallows	Clay/Hardpan	Macrophytes
Rip Rap	Bulkhead	
Other _____		

Biological Characteristics

Riffle:
- Predominant Organism: _____
- Other Common Organisms: _____
- Density: High Moderate Low
- Diversity: High Moderate Low

Run:
- Predominant Organism: _____
- Other Common Organisms: _____
- Density: High Moderate Low
- Diversity: High Moderate Low

Pool:
- Predominant Organism: _____
- Other Common Organisms: _____
- Density: High Moderate Low
- Diversity: High Moderate Low

Margin:
- Predominant Organism: _____
- Other Common Organisms: _____
- Density: High Moderate Low
- Diversity: High Moderate Low

Other Notable Collections: _____

V= Very Abundant; A= Abundant; C= Common; R= Rare

Overall Amount (V=>151; A= 150-101; C= 100-11; R= 10-1)	
/	Porifera, Bryozoa
/ /	Turbellaria, Oligochaeta, Hirudinea
/	Isopoda, Amphipoda
/	Decapoda, Hydracarina
	Ephemeroptera
	Baetidae
/ /	Heptageniidae, Leptohyphidae, Caenidae
	Other _____
/	Zygoptera, Anisoptera
	Plecoptera
	Hemiptera
/	Megaloptera, Neuroptera
	Trichoptera
	Hydropsychidae
/	Hydroptilidae, Leptoceridae
	Other _____
	Coleoptera
	Elimidae
	Other _____
	Diptera
	Chironomidae
	Other _____
/	Gastropoda, Bivalvia
	Other _____

Field Narrative Rating: E VG G MG F P VP

OhioEPA Qualitative Habitat Evaluation Index and Use Assessment Field Sheet

QHEI Score: ____

Stream & Location: _____ **RM:** _ _._ _ **Date:** _ _/ _ _/ _ _

Scorers Full Name & Affiliation: Northeast Ohio Regional Sewer District

River Code: _ _ _-_ _ _-_ _ _ **STORET #:** _ _ _ _ _ _ **Lat./ Long.:** (NAD 83 - decimal °) _ _ . _ _ _ _ /8_ . _ _ _ _ **Office verified location** ☐

1] *SUBSTRATE* Check *ONLY* Two substrate *TYPE BOXES*; estimate % or note every type present

BEST TYPES	POOL	RIFFLE	OTHER TYPES	POOL	RIFFLE
☐☐ BLDR /SLABS [10]	____	____	☐☐ HARDPAN [4]	____	____
☐☐ BOULDER [9]	____	____	☐☐ DETRITUS [3]	____	____
☐☐ COBBLE [8]	____	____	☐☐ MUCK [2]	____	____
☐☐ GRAVEL [7]	____	____	☐☐ SILT [2]	____	____
☐☐ SAND [6]	____	____	☐☐ ARTIFICIAL [0]	____	____
☐☐ BEDROCK [5]	____	____	(Score natural substrates; ignore sludge from point-sources)		

NUMBER OF BEST TYPES: ☐ 4 or more [2] ☐ 3 or less [0]

Check ONE (*Or 2 & average*)

ORIGIN
- ☐ LIMESTONE [1]
- ☐ TILLS [1]
- ☐ WETLANDS [0]
- ☐ HARDPAN [0]
- ☐ SANDSTONE [0]
- ☐ RIP/RAP [0]
- ☐ LACUSTURINE [0]
- ☐ SHALE [-1]
- ☐ COAL FINES [-2]

QUALITY

SILT
- ☐ HEAVY [-2]
- ☐ MODERATE [-1]
- ☐ NORMAL [0]
- ☐ FREE [1]

EMBEDDEDNESS
- ☐ EXTENSIVE [-2]
- ☐ MODERATE [-1]
- ☐ NORMAL [0]
- ☐ NONE [1]

Substrate ____ *Maximum 20*

Comments

2] *INSTREAM COVER* Indicate presence 0 to 3: 0-Absent; 1-Very small amounts or if more common of marginal quality; 2-Moderate amounts, but not of highest quality or in small amounts of highest quality; 3-Highest quality in moderate or greater amounts (e.g., very large boulders in deep or fast water, large diameter log that is stable, well developed rootwad in deep / fast water, or deep, well-defined, functional pools.

- ____ UNDERCUT BANKS [1]
- ____ OVERHANGING VEGETATION [1]
- ____ SHALLOWS (IN SLOW WATER) [1]
- ____ ROOTMATS [1]
- ____ POOLS > 70cm [2]
- ____ ROOTWADS [1]
- ____ BOULDERS [1]
- ____ OXBOWS, BACKWATERS [1]
- ____ AQUATIC MACROPHYTES [1]
- ____ LOGS OR WOODY DEBRIS [1]

AMOUNT Check ONE (*Or 2 & average*)
- ☐ EXTENSIVE >75% [11]
- ☐ MODERATE 25-75% [7]
- ☐ SPARSE 5-<25% [3]
- ☐ NEARLY ABSENT <5% [1]

Cover Maximum 20 ____

Comments

3] *CHANNEL MORPHOLOGY* Check ONE in each category (*Or 2 & average*)

SINUOSITY
- ☐ HIGH [4]
- ☐ MODERATE [3]
- ☐ LOW [2]
- ☐ NONE [1]

DEVELOPMENT
- ☐ EXCELLENT [7]
- ☐ GOOD [5]
- ☐ FAIR [3]
- ☐ POOR [1]

CHANNELIZATION
- ☐ NONE [6]
- ☐ RECOVERED [4]
- ☐ RECOVERING [3]
- ☐ RECENT OR NO RECOVERY [1]

STABILITY
- ☐ HIGH [3]
- ☐ MODERATE [2]
- ☐ LOW [1]

Channel Maximum 20 ____

Comments

4] *BANK EROSION AND RIPARIAN ZONE* Check ONE in each category for *EACH BANK* (*Or 2 per bank & average*)

River right looking downstream

EROSION

L	R	
☐	☐	NONE / LITTLE [3]
☐	☐	MODERATE [2]
☐	☐	HEAVY / SEVERE [1]

RIPARIAN WIDTH

L	R	
☐	☐	WIDE > 50m [4]
☐	☐	MODERATE 10-50m [3]
☐	☐	NARROW 5-10m [2]
☐	☐	VERY NARROW < 5m [1]
☐	☐	NONE [0]

FLOOD PLAIN QUALITY

L	R	
☐	☐	FOREST, SWAMP [3]
☐	☐	SHRUB OR OLD FIELD [2]
☐	☐	RESIDENTIAL, PARK, NEW FIELD [1]
☐	☐	FENCED PASTURE [1]
☐	☐	OPEN PASTURE, ROWCROP [0]

L	R	
☐	☐	CONSERVATION TILLAGE [1]
☐	☐	URBAN OR INDUSTRIAL [0]
☐	☐	MINING / CONSTRUCTION [0]

Indicate predominant land use(s) past 100m riparian.

Riparian Maximum 10 ____

Comments

5] *POOL / GLIDE AND RIFFLE / RUN QUALITY*

MAXIMUM DEPTH Check ONE (*ONLY!*)
- ☐ > 1m [6]
- ☐ 0.7-<1m [4]
- ☐ 0.4-<0.7m [2]
- ☐ 0.2-<0.4m [1]
- ☐ < 0.2m [0]

CHANNEL WIDTH Check ONE (*Or 2 & average*)
- ☐ POOL WIDTH > RIFFLE WIDTH [2]
- ☐ POOL WIDTH = RIFFLE WIDTH [1]
- ☐ POOL WIDTH < RIFFLE WIDTH [0]

CURRENT VELOCITY Check ALL that apply
- ☐ TORRENTIAL [-1]
- ☐ VERY FAST [1]
- ☐ FAST [1]
- ☐ MODERATE [1]
- ☐ SLOW [1]
- ☐ INTERSTITIAL [-1]
- ☐ INTERMITTENT [-2]
- ☐ EDDIES [1]

Indicate for reach - pools and riffles.

Recreation Potential
Primary Contact
Secondary Contact
(circle one and comment on back)

Pool / Current Maximum 12 ____

Comments

Indicate for functional riffles; Best areas must be large enough to support a population of riffle-obligate species: Check ONE (*Or 2 & average*). ☐ NO RIFFLE [metric=0]

RIFFLE DEPTH
- ☐ BEST AREAS > 10cm [2]
- ☐ BEST AREAS 5-10cm [1]
- ☐ BEST AREAS < 5cm [metric=0]

RUN DEPTH
- ☐ MAXIMUM > 50cm [2]
- ☐ MAXIMUM < 50cm [1]

RIFFLE / RUN SUBSTRATE
- ☐ STABLE (e.g., Cobble, Boulder) [2]
- ☐ MOD. STABLE (e.g., Large Gravel) [1]
- ☐ UNSTABLE (e.g., Fine Gravel, Sand) [0]

RIFFLE / RUN EMBEDDEDNESS
- ☐ NONE [2]
- ☐ LOW [1]
- ☐ MODERATE [0]
- ☐ EXTENSIVE [-1]

Riffle / Run Maximum 8 ____

Comments

6] *GRADIENT* (ft/mi)

DRAINAGE AREA (mi²)

- ☐ VERY LOW - LOW [2-4]
- ☐ MODERATE [6-10]
- ☐ HIGH - VERY HIGH [10-6]

%POOL: ____ %GLIDE: ____

%RUN: ____ %RIFFLE: ____

Gradient Maximum 10 ____

A] SAMPLED REACH

Check ALL that apply

Comment RE: Reach consistency/ Is reach typical of steam?, *Recreation/* Observed - Inferred, *Other/* Sampling observations, Concerns, Access directions, etc.

METHOD

- ☐ BOAT
- ☐ WADE
- ☐ L. LINE
- ☐ OTHER

STAGE

1st -sample pass-	2nd
☐ HIGH	☐
☐ UP	☐
☐ NORMAL	☐
☐ LOW	☐
☐ DRY	☐

DISTANCE

- ☐ 0.5 Km
- ☐ 0.2 Km
- ☐ 0.15 Km
- ☐ 0.12 Km
- ☐ OTHER

_____ meters

CLARITY

1st --sample pass--	2nd
☐ < 20 cm	☐
☐ 20-<40 cm	☐
☐ 40-70 cm	☐
☐ > 70 cm/ CTB	☐
☐ SECCHI DEPTH	☐

CANOPY

- ☐ > 85%- OPEN
- ☐ 55%-<85%
- ☐ 30%-<55%
- ☐ 10%-<30%
- ☐ <10%- CLOSED

pass
1st_____ cm
2nd_____ cm

B] AESTHETICS

- ☐ NUISANCE ALGAE
- ☐ INVASIVE MACROPHYTES
- ☐ EXCESS TURBIDITY
- ☐ DISCOLORATION
- ☐ FOAM / SCUM
- ☐ OIL SHEEN
- ☐ TRASH / LITTER
- ☐ NUISANCE ODOR
- ☐ SLUDGE DEPOSITS
- ☐ CSOs/SSOs/OUTFALLS

C] RECREATION

POOL: AREA ☐ >100ft² DEPTH ☐ >3ft

D] MAINTENANCE

Circle some & *COMMENT*

PUBLIC / PRIVATE / BOTH / NA
ACTIVE / HISTORIC / BOTH / NA
YOUNG-SUCCESSION-OLD
SPRAY / SNAG / REMOVED
MODIFIED / DIPPED OUT / NA
LEVEED / ONE SIDED
RELOCATED / CUTOFFS
MOVING-BEDLOAD-STABLE
ARMOURED / SLUMPS
ISLANDS / SCOURED
IMPOUNDED / DESICCATED
FLOOD CONTROL / DRAINAGE

E] ISSUES

WWTP / CSO / NPDES / INDUSTRY
HARDENED / URBAN / DIRT&GRIME
CONTAMINATED / LANDFILL
BMPs-CONSTRUCTION-SEDIMENT
LOGGING / IRRIGATION / COOLING
BANK / EROSION / SURFACE
FALSE BANK / MANURE / LAGOON
WASH H_20 / TILE / H_20 TABLE
ACID / MINE / QUARRY / FLOW
NATURAL / WETLAND / STAGNANT
PARK / GOLF / LAWN / HOME
ATMOSPHERE / DATA PAUCITY

F] MEASUREMENTS

- $\bar{x}$ width
- $\bar{x}$ depth
- max. depth
- $\bar{x}$ bankfull width
- bankfull $\bar{x}$ depth
- W/D ratio
- bankfull max. depth
- floodprone x^2 width
- entrench. ratio

Legacy Tree:

Stream Drawing:

Ohio EPA Primary Headwater Habitat Evaluation Form

HHEI Score (sum of metrics 1, 2, 3) : 1

SITE NAME/LOCATION _____

SITE NUMBER _____ RIVER BASIN _____ DRAINAGE AREA (mi²) _____

LENGTH OF STREAM REACH (ft) _____ LAT. _____ LONG. _____ RIVER CODE _____ RIVER MILE _____

DATE _____ SCORER _____ COMMENTS _____

NOTE: Complete All Items On This Form - Refer to "Field Evaluation Manual for Ohio's PHWH Streams" for Instructions

STREAM CHANNEL MODIFICATIONS: ☐ NONE / NATURAL CHANNEL ☐ RECOVERED ☐ RECOVERING ☐ RECENT OR NO RECOVERY

1. SUBSTRATE (Estimate percent of every type of substrate present. Check *ONLY* two predominant substrate *TYPE* boxes (Max of 32). Add total number of significant substrate types found (Max of 8). Final metric score is sum of boxes A & B.

TYPE		PERCENT	TYPE		PERCENT
☐☐	BLDR SLABS [16 pts]	0%	☐☐	SILT [3 pt]	0%
☐☐	BOULDER (>256 mm) [16 pts]	0%	☐☐	LEAF PACK/WOODY DEBRIS [3 pts]	0%
☐☐	BEDROCK [16 pt]	0%	☐☐	FINE DETRITUS [3 pts]	0%
☐☐	COBBLE (65-256 mm) [12 pts]	0%	☐☐	CLAY or HARDPAN [0 pt]	0%
☐☐	GRAVEL (2-64 mm) [9 pts]	0%	☐☐	MUCK [0 pts]	0%
☐☐	SAND (<2 mm) [6 pts]	0%	☐☐	ARTIFICIAL [3 pts]	0%

Total of Percentages of Bldr Slabs, Boulder, Cobble, Bedrock **0.00%** (A) Substrate Percentage Check 0% (B)

SCORE OF TWO MOST PREDOMINATE SUBSTRATE TYPES: 0 **TOTAL NUMBER OF SUBSTRATE TYPES:** 1

HHEI Metric Points

Substrate Max = 40

1

A + B

2. Maximum Pool Depth ***(Measure the maximum pool depth within the 61 meter (200 ft)*** evaluation reach at the time of evaluation. Avoid plunge pools from road culverts or storm water pipes) (Check *ONLY* one box):

☐	> 30 centimeters [20 pts]	☐	> 5 cm - 10 cm [15 pts]
☐	> 22.5 - 30 cm [30 pts]	☐	< 5 cm [5 pts]
☐	> 10 - 22.5 cm [25 pts]	☐	NO WATER OR MOIST CHANNEL [0 pts]

COMMENTS _____ **MAXIMUM POOL DEPTH (centimeters):** _____

Pool Depth Max = 30

0

3. BANK FULL WIDTH (Measured as the average of 3-4 measurements) **(Check *ONLY* one box):**

☐	> 4.0 meters (> 13') [30 pts]	☐	> 1.0 m - 1.5 m (> 3' 3" - 4' 8") [15 pts]
☐	> 3.0 m - 4.0 m (> 9' 7" - 13') [25 pts]	☐	≤ 1.0 m (<=3' 3") [5 pts]
☐	> 1.5 m - 3.0 m (> 9' 7" - 4' 8") [20 pts]		

COMMENTS _____ **AVERAGE BANKFULL WIDTH (meters):** _____

Bankfull Width Max=30

0

This information must also be completed

RIPARIAN ZONE AND FLOODPLAIN QUALITY ☆NOTE: River Left (L) and Right (R) as looking downstream☆

RIPARIAN WIDTH (Per Bank) L	R	FLOODPLAIN QUALITY (Most Predominant per Bank) L	R		L	R	
☐	☐	Wide >10m	☐	☐	Mature Forest, Wetland	☐	☐	Conservation Tillage
☐	☐	Moderate 5-10m	☐	☐	Immature Forest, Shrub or Old Field	☐	☐	Urban or Industrial
☐	☐	Narrow <5m	☐	☐	Residential, Park, New Field	☐	☐	Open Pasture, Row Crop
☐	☐	None	☐	☐	Fenced Pasture	☐	☐	Mining or Construction

COMMENTS _____

FLOW REGIME *(At Time of Evaluation)* (Check *ONLY* one box):

☐ Stream Flowing
☐ Subsurface flow with isolated pools (Interstitial)
☐ Moist Channel, isolated pools, no flow (Intermittent)
☐ Dry channel, no water (Ephemeral)

COMMENTS _____

SINUOSITY (Number of bends per 61 m (200 ft) of channel) (Check *ONLY* one box):

☐ None ☐ 0.5 ☐ 1.0 ☐ 1.5 ☐ 2.0 ☐ 2.5 ☐ 3.0 ☐ >3

STREAM GRADIENT ESTIMATE

☐ Flat (0.5 ft/100 ft) ☐ Flat to Moderate ☐ Moderate (2 ft/100 ft) ☐ Moderate to Severe ☐ Severe (10 ft/100 ft)

ADDITIONAL STREAM INFORMATION (This Information Must Also be Completed):

QHEI PERFORMED? - ☐ Yes ☐ No QHEI Score _____ (If Yes, Attach Completed QHEI Form)

DOWNSTREAM DESIGNATED USE(S)

☐ WWH Name: _____ Distance from Evaluated Stream _____

☐ CWH Name: _____ Distance from Evaluated Stream _____

☐ EWH Name: _____ Distance from Evaluated Stream _____

MAPPING: ATTACH COPIES OF MAPS, INCLUDING THE ENTIRE WATERSHED AREA. CLEARLY MARK THE SITE LOCATION

USGS Quadrangle Name: _____ NRCS Soil Map Page: _____ NRCS Soil Map Stream Order _____

County: Wyandot Township / City: _____

MISCELLANEOUS

Base Flow Conditions? (Y/N): Y Date of last precipitation: _____ Quantity: 0.00

Photograph Information: _____

Elevated Turbidity? (Y/N): Y Canopy (% open): 0%

Were samples collected for water chemistry? (Y/N): Y (Note lab sample no. or id. and attach results) Lab Number: _____

Field Measures: Temp (°C) _____ Dissolved Oxygen (mg/l) _____ pH (S.U.) _____ Conductivity (µmhos/cm) _____

Is the sampling reach representative of the stream (Y/N) Y If not, please explain: _____

Additional comments/description of pollution impacts: _____

BIOTIC EVALUATION

Performed? (Y/N): Y (If Yes, Record all observations. Voucher collections optional. NOTE: all voucher samples must be labeled with the site ID number. Include appropriate field data sheets from the Primary Headwater Habitat Assessment Manual)

Fish Observed? (Y/N) Y Voucher? (Y/N) Y Salamanders Observed? (Y/N) Y Voucher? (Y/N) Y

Frogs or Tadpoles Observed? (Y/N) Y Voucher? (Y/N) Y Aquatic Macroinvertebrates Observed? (Y/N) Y Voucher? (Y/N) Y

Comments Regarding Biology: _____

DRAWING AND NARRATIVE DESCRIPTION OF STREAM REACH (This must be completed):

Include important landmarks and other features of interest for site evaluation and a narrative description of the stream's location

FLOW ➔

Save as pdf Reset Form

Lake / Lacustuary (Lentic) QHEI Field Sheet

Ohio Environmental Protection Agency

QHEI Score: ☐

RIVERCODE_____ RIVERMILE _____ WATERBODY _____ DISTANCE ASSESSED (m): _____
DATE _____ LOCATION _____
SCORER_____ LAT. _____ LONG. _____ COMMENT _____

1] SUBSTRATE (Check *ONLY* Two Substrate TYPE BOXES; Estimate % or note every type present); LAKE:____ LACUSTUARY:____

TYPE	SHORE	BOTTOM		SHORE	BOTTOM
☐☐-BLDR/SLABS [7]			☐☐-HARDPAN [4]		
☐☐-BOULDER [10]			☐☐-BEDROCK [3]		
☐☐-COBBLE [8]			☐☐-DETRITUS [3]		
☐☐-GRAVEL [7]			☐☐-SILT [2]		
☐☐-SAND [6]			☐☐-MUCK [2]		

NOTE: Ignore sludge that originates from point-sources, score on natural substrates

NUMBER OF SUBSTRATE TYPES: ☐-5 or More [2] ☐-4 or Less [0]

SUBSTRATE ORIGIN Check ONE (or 2 & *AVERAGE*)
☐-LIMESTONE [1] ☐-TILLS [1] ☐-WETLANDS [1] ☐-LACUSTUARINE [1] ☐-SANDSTONE [1] ☐-RIP/RAP [1] ☐-HARDPAN [0] ☐-SHALE [-1] ☐-COAL/ORE [-2]

SILT:

SILT ORIGIN:

SUBSTRATE QUALITY Check ONE (or 2 & *AVERAGE*)
☐-SILT HEAVY [-2] ☐-SILT MODERATE [-1] ☐-SILT NORMAL [0] ☐-SILT FREE [1] ☐-CLAY [-2] ☐-INDUSTRIAL [-1] ☐-ORGANIC [1] ☐-NONE [1]

Substrate ☐ Max 20

COMMENTS: _____

2] COVER TYPES **TYPE:** (Check All That Apply) **AMOUNT**: (Check *ONLY* One or check2 and AVERAGE)

☐-OFF-SHORE SAND BARS [4]	☐-DEEPWATER > 1 M [1]	☐-WETLAND POOLS [1]	☐-EXTENSIVE > 75% [9]
☐-OVERHANGING VEGETATION [1]	☐-ROOTWADS [1]	☐-SUBMERGED AQUATIC VEG. [4]	☐-MODERATE 25-75% [7]
☐-SHALLOWS (ON BEACH) [1]	☐-BOULDERS [1]	☐-LOGS OR WOODY DEBRIS [1]	☐-SPARSE 5-25% [3]
☐-ROOTMATS [1]	☐-SAND BEACH [1]	☐-GRAVEL BEACH [1]	☐-NEARLY ABSENT < 5% [1]

Cover ☐ Max 20

COMMENTS: _____

3] SHORELINE MORPHOLOGY (Check *ONLY* one PER category or check 2 and *AVERAGE*)

SHORE SINUOSITY
☐-HIGH [2] ☐-MODERATE [4] ☐-LOW [3] ☐-NONE [1]

DEVELOPMENT
☐-EXCELLENT [5] ☐-GOOD [5] ☐-FAIR [3] ☐-POOR [1]

MODIFICATION
☐-NONE [7] ☐-RECOVERED [5] ☐-RECOVERING [3] ☐-RECENT OR NO RECOVERY [1]

STABILITY
☐-HIGH [3] ☐-MODERATE [2] ☐-LOW [1]

SHORE to BOTTOM SLOPE MORPHOLOGIES
☐-SLOPE < 15 deg. [0] ☐-SLOPE > 45 deg. [2] ☐-SLOPE < 25 deg. [1] ☐-SLOPE 90 deg. [0] ☐-SLOPE > 25 deg. [3]

AVERAGE DEPTH (of 5 measures)
☐- < 50 cm [0] ☐- > 400 – 500 cm [4] ☐- 50 - <100 cm [1] ☐- > 500 - 900 cm [2] ☐- ≥100 - 200 cm [2] ☐- >900 cm [1] ☐- >200 -4 00 cm [3]

MODIFICATIONS OF SAMPLED SHORELINE
☐- CEMENTED [-1] ☐- STEEL BULKHEADS [-2] ☐-RIP RAPPED [1] ☐-ISLANDS [1] ☐-RAILROAD TIES [-1] ☐-DIKES [-1] ☐-DREDGED [-1] ☐-BANK SHAPING [-1] ☐-TWO SIDE CHANNEL MODIFICATIONS [-1] ☐-WOOD PILINGS [1] ☐-SHIP CHANNEL [-2]

Shore Line ☐ Max 20

COMMENTS: _____

★ Shore Right Looking East or South on Lake ★
★ Shore Right Looking Toward Lake in Lacustuary ★

4] RIPARIAN ZONE AND BANK EROSION (Check *ONE* box PER bank or 2 and *AVERAGE*)

RIPARIAN WIDTH L R (Per Bank)
☐☐-WIDE > 50 m [4] ☐☐-MODERATE 10-50 m [3] ☐☐-NARROW 5-10 m [2] ☐☐-VERY NARROW < 5 m [1] ☐☐-NONE [0]

SHORE LINE QUALITY (PAST 100 FOOT RIPARIAN) L R (Most Predominant Per Bank)
☐☐-FOREST, WETLAND, LAKE [3] ☐☐- SHRUB OR OLD FIELD [2] ☐☐-VINEYARD, ORCHARD [2] ☐☐-FENCED PASTURE [1] ☐☐-RESIDENTIAL, PARK, NEW FIELD [1]
L R
☐☐-CONSERVATION TILLIAGE [1] ☐☐-URBAN OR INDUSTRIAL [0] ☐☐-OPEN PASUTRE, ROWCROP [0] ☐☐-MINING/CONSTRUCTION [0] ☐☐- DIKED WETLAND [0]

BANK EROSION L R (Per Bank)
☐☐-NONE/LITTLE [3] ☐☐-MODERATE [-1] ☐☐-HEAVY/SEVERE [-3]

Riparian ☐ Max 10

COMMENTS: _____

5] AQUATIC VEGETATION QUALITY: PLANT SPECIES OBSERVED (Sum All Scores)
(Score all for observed abundance: ABUNDANT = [3]; COMMON = [5], FEW = [1]; UNCOMMON = [0]) _____NO AQUATIC VEGETATION = 0

____-Pond Lilies (NYMPHAEA) ____-Sedge (CYPERACEAE) ____-Wild Celery (VALLISNERIA)
____-Pond Weed (POTAMOGETON) ____-Bulrush (SCIRPUS) ____-Waterweed (ELODEA) ____-Wild Rice (ZIZANIA)

(Score all for observed abundance: ABUNDANT = [-2]; COMMON = [-1]; FEW = [0])

____-Purple Loosestrife ____-Reed Grass ____-Eurasian Milfoil ____-Cattails ____-Algae (mats) ____-Algae (planktonic)

Vegetation ☐ Max 30

COMMENTS: _____

Is the Sampling Reach Representative of Area Habitat? (Y/N)____ If Not, Explain: _____

Depth measures: | | | | | | |

Zebra Mussel/Quagga Mussel Coverage ☐ >60% ☐ 60->25% ☐ 25->10% ☐ <10->1% ☐ 1–0%

	Gear	Distance	Water Clarity	Wave Height
First Sampling Pass:	_____	_____	_____	_____
Second Sampling Pass:	_____	_____	_____	_____
Third Sampling Pass:	_____	_____	_____	_____

☐ Subjective Rating (1–10)

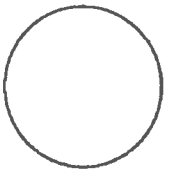
☐ Aesthetic Rating (1–10)

Photos: _____

WATERBODY MEASUREMENTS AVERAGE WIDTH: _____ AVERAGE DEPTH: _____ Maximum Depth: _____

DRAWING OF SITE:

North Arrow:



Enter Site Name or ID _____ Date: _____

PHWH STREAM BIOLOGICAL CHARATERISTICS FIELD SHEET:

1. Fish:

Voucher Specimens Retained? (select) Yes Time Spent (minutes): _____

Sample Method No Evaluation Stream Length Assessed (meters) _____

Species	Number Caught	Notes
Blank	0	
Blank	0	
Blank	0	
Blank	0	
	0	
	0	
	0	
	0	

2. Salamanders:

Voucher Specimens Retained? (circle) N Time Spent (minutes): _____

Sample Method No Evaluation Stream Length Assessed (meters) _____

Species *(Genus)*	# Larvae	# Juveniles/Adults	Total Number
Mountain Dusky (*Desmognathus ochrophaeus)*	0	0	0
Northern Dusky (*Desmognathus fuscus)*	0	0	0
Two-lined *(Eurycea bislineata)*	0	0	0
Long-tailed *(Eurycea longicauda)*	0	0	0
Cave (*Eurycea lucifuga)*	0	0	0
Red *(Pseudotriton ruber)*	0	0	0
Mud *(Pseudotriton montanus)*	0	0	0
Spring *(Gyrinophilus porphyriticus)*	0	0	0
Mole spp. *(Ambystoma spp.)*	0	0	0
Four-toed *(Hemidactylium scutatum)*	0	0	0
Other (name)	0	0	0
Total	0	0	0

Notes on Vertebrates: _____

3. Macroinvertebrate Scoring Sheet:

THE HEADWATER MACROINVERTEBRATE FIELD EVALUATION INDEX (HMFEI) SCORING SHEET

Indicate Abundance of Each Taxa Above each White Box.

Record HMFEI Scoring Value Points Within each Box.

For EPT taxa, also indicate the different taxa present.

Key: **V** = Very Abundant (> 50); **A** = Abundant (10 -50); **C** = Common (3 -9); **R** = Rare (< 3)

Sessile Animals (**Porifera, Cnidaria, Bryozoa**) (HMFEI pts = 1) NA 0	Crayfish (**Decapoda**) (HMFEI pts = 2) NA 0	Fishfly Larvae (**Corydalidae**) (HMFEI pts = 3) NA 0
Aquatic Worms (**Turbellaria, Hirudinea, Oligochaeta**) (HMFEI pts = 1) NA 0	Dragonfly Nymphs (**Anisoptera**) (HMFEI pts = 2) NA 0	Water Penny Beetles (**Psephenidae**) (HMFEI pts = 3) NA 0
Sow Bugs (**Isopoda**) (HMFEI pts = 1) NA 0	Riffle Beetles (**Dryopidae, Elmidae, Ptilodactylidae**) (HMFEI pts = 2) NA 0	Cranefly Larvae (**Tipulidae**) (HMFEI pts = 3) NA 0
Scuds (**Amphipoda**) (HMFEI pts = 1) NA 0	Larvae of other Flies (enter name in comments) (**Diptera**): (HMFEI pts = 1) NA 0	**EPT TAXA*** Total No. EPT Taxa = 0
Water Mites (**Hydracarina**) (HMFEI pts = 1) NA 0	Midges (**Chironomidae**) (HMFEI pts = 1) NA 0	Mayfly Nymphs (**Ephemeroptera**) Taxa Present: 0 [HMFEI pts = No. Taxa (x) 3] NA 0
Damselfly Nymphs (**Zygoptera**) (HMFEI pts = 1) NA 0	Snails (**Gastropoda**) (HMFEI pts = 1) NA 0	
Alderfly Larvae (**Sialidae**) (HMFEI pts = 1) NA 0	Clams (**Bivalvia**) (HMFEI pts = 1) NA 0	Stonefly Nymphs (**Plecoptera**) Taxa Present: 0 [HMFEI pts = No. Taxa (x) 3] NA 0
Other Beetles (**Coleoptera**) (HMFEI pts = 1) NA 0	Other Taxa :	
Other Taxa:	Other Taxa:	Caddisfly Larvae (**Trichoptera**) Taxa Present: 0 [HMFEI pts = No. Taxa (x) 3] NA 0
Other Taxa:	Other Taxa	

*Note: EPT identification based upon Family or Genus level of taxonomy

Voucher Sample ID_____ Time Spent (minutes):_____

Notes on Macroinvertebrates: (Predominant Organisms; Other Common Organisms; Diversity Estimate)

Final HMFEI Calculated Score (Sum of All White Box Scores) = 0

IF Final HMFEI Score is > 19, Then CLASS III PHWH STREAM
IF Final HMFEI Score is 7 to 19, Then CLASS II PHWH STREAM
IF Final HMFEI Score is < 7, Then CLASS I PHWH STREAM

Save as pdf

Reset Form

NEORSD Surface Water Condition Sampling Field Data Form

Stream: _____ Date: _____ Collectors: _____

Gage Station and ID: _____ Daily Mean Discharge: _____ ft^3/sec

Was this sample taken during or following a wet weather event? YES / NO

Water Quality Meters Used: _____

Time (hrs): _____ River Mile (Site): _____

Weather: Clear Partly Cloudy Overcast Light Rain/Showers Heavy Rain
Steady Rain Heavy Snow Melt Other: _____

Flow: Dry Intermittent Minimal Baseline/Normal Elevated Flood

HD Status: OK Other: _____

Color: Clear Muddy Tea Milky Other: _____

Odor: Normal Petroleum Anaerobic Sewage Chemical Other: _____

Surface Coating: None Foam Oily Scum Other: _____

Field Parameters: Conductivity (µmhos/cm): _____ Sp. Cond. (µmhos/cm): _____

Dissolved Oxygen (mg/L): _____ D.O. (%): _____

Temperature (°C): _____ pH (s.u.): _____

Turbidity 1 (NTU): _____ Turbidity 2 (NTU): _____ Average (NTU): _____

Sample ID:

General Comments: _____

Time (hrs): _____ River Mile (Site): _____

Weather: Clear Partly Cloudy Overcast Light Rain/Showers Heavy Rain
Steady Rain Heavy Snow Melt Other: _____

Flow: Dry Intermittent Minimal Baseline/Normal Elevated Flood

HD Status: OK Other: _____

Color: Clear Muddy Tea Milky Other: _____

Odor: Normal Petroleum Anaerobic Sewage Chemical Other: _____

Surface Coating: None Foam Oily Scum Other: _____

Field Parameters: Conductivity (µmhos/cm): _____ Sp. Cond. (µmhos/cm): _____

Dissolved Oxygen (mg/L): _____ D.O. (%): _____

Temperature (°C): _____ pH (s.u.): _____

Turbidity 1 (NTU): _____ Turbidity 2 (NTU): _____ Average (NTU): _____

Sample ID:

General Comments: _____

Appendix B

Parameter	Additional Name	Test	2014 Minimum Detection Limit	2014 Practical Quantitation Limit
Alkalinity	Alkalinity	EPA 310.2	1.6 mg/L	5.0 mg/L
Mercury	Hg	EPA 245.1	0.006 µg/L	0.050 µg/L
Ammonia [1]	NH_3	EPA 350.1	0.002 mg/L	0.020 mg/L
Nitrite	NO2	SM 4500 NO_2^- B [2]	0.001 mg/L	0.020 mg/L
Nitrite + Nitrate	$NO_2 + NO_3$	EPA 353.2	0.003 mg/L	0.020 mg/L
Total Kjeldahl Nitrogen	TKN	EPA 351.2	0.081 mg/L	0.500 mg/L
Dissolved Reactive Phosphorus	DRPhos	EPA 365.1	0.003 mg/L	0.010 mg/L
Total Phosphorus	Total-P	EPA 365.1	0.003 mg/L	0.010 mg/L
Chloride	Chloride by IC	EPA 300.0	1.00 mg/L	5.00 mg/L
Sulfate	Sulfate by IC	EPA 300.0	0.500 mg/L	5.00 mg/L
Silver	Ag	EPA 200.8	0.009 µg/L	0.500 µg/L
Aluminum	Al	EPA 200.8	0.504 µg/L	5.000 µg/L
Arsenic	As	EPA 200.8	0.320 µg/L	1.000 µg/L
Barium	Ba	EPA 200.8	0.033 µg/L	0.500 µg/L
Beryllium	Be	EPA 200.8	0.540 µg/L	0.500 µg/L
Calcium	Ca	EPA 200.8	16.90 µg/L	125.0 µg/L
Cadmium	Cd	EPA 200.8	0.034 µg/L	0.500 µg/L
Cobalt	Co	EPA 200.8	0.056 µg/L	0.500 µg/L
Chromium	Cr	EPA 200.8	0.049 µg/L	0.500 µg/L
Copper	Cu	EPA 200.8	0.073 µg/L	1.000 µg/L
Iron	Fe	EPA 200.8	0.495 µg/L	5.000 µg/L
Potassium	K	EPA 200.8	3.695 µg/L	125.0 µg/L
Magnesium	Mg	EPA 200.8	2.077 µg/L	125.0 µg/L
Manganese	Mn	EPA 200.8	0.057 µg/L	1.000 µg/L
Molybdenum	Mo	EPA 200.8	0.017 µg/L	0.500 µg/L
Sodium	Na	EPA 200.8	13.910 µg/L	125.0 µg/L
Nickel	Ni	EPA 200.8	0.066 µg/L	2.000 µg/L
Lead	Pb	EPA 200.8	0.058 µg/L	0.500 µg/L
Antimony	Sb	EPA 200.8	0.018 µg/L	0.500 µg/L
Selenium	Se	EPA 200.8	0.383 µg/L	2.500 µg/L
Tin	Sn	EPA 200.8	0.018 µg/L	0.500 µg/L
Strontium	Sr	EPA 200.8	0.049 µg/L	0.500 µg/L
Titanium	Ti	EPA 200.8	0.071 µg/L	1.000 µg/L
Thallium	Tl	EPA 200.8	0.007 µg/L	0.500 µg/L
Vanadium	V	EPA 200.8	0.238 µg/L	5.000 µg/L
Zinc	Zn	EPA 200.8	0.241 µg/L	5.000 µg/L
Total Metals	Total Metals (calc.)	EPA 200.8	µg/L =(Cr µg/L)+(Cu µg/L)+(Ni µg/L)+(Zn µg/L)	
Hardness	Hardness (calc.)	SM 2340 [2]	CaCO3 mg/L =(2.497*Ca mg/L)+(4.118*Mg mg/L)	
Escherichia coli	*E. coli*	EPA 1603	1 colony	--
		Colilert QT (SM 9223 B 20th Ed)	1 MPN	1 MPN
Chlorophyll *a*	Chlorophyll *a*	EPA 445.0	0.03 µg/L	0.15 µg/L
Chemical Oxygen Demand	COD	EPA 410.4	0.49 mg/L	10 mg/L
Biological Oxygen Demand	BOD	SM 5210 [2]	2 mg/L	--
Total Solids	TS	SM 2540 B [2]	1.0 mg/L	5.0 mg/L
Total Suspended Solids	TSS	SM 2540 D [2]	0.5 mg/L	1.0 mg/L
Total Dissolved Solids	TDS	SM 2540 C [2]	1.0 mg/L	5.0 mg/L
Turbidity **		EPA 180.1	0.1 NTU	0.2 NTU
Field Parameter		**Test**	**(Value Reported in)**	
pH		EPA 150.1 [2]	s.u.	
Conductivity		SM 2510A [2]	µs/cm	
Specific Conductivity		SM 2510B [2]	µs/cm	
Dissolved Oxygen	DO	SM 4500-O G [2]	mg/L	
Temperature	Temp	EPA 1701.1 [2]	°C	
Turbidity **		EPA 180.1	NTU	

[1] Listed MDL/PQL is for undistilled samples. Any samples that require distillation will have a MDL = 0.020 mg/L, PQL = 0.100 mg/L

[2] Standard Methods for the Examination of Water and Wastewater, Method approved by Standard Methods Committee, 1997. Editorial revisions, 2011

** Turbidity will either be completed in the field or at the laboratory.

Appendix C

Y S I Environmental

YSI 556 Multiparameter System

Versatile, multiparameter handheld instrument

The 556 has multiple language capabilities and graphing!

Rugged and reliable, the YSI 556 MPS (Multiprobe System) combines the versatility of an easy-to-use, easy-to-read handheld unit with all the functionality of a multiparameter system.

- Simultaneously measures dissolved oxygen, pH, conductivity, temperature, and ORP
- Field-replaceable electrodes
- Compatible with EcoWatch® for Windows® data analysis software
- Stores over 49,000 data sets, time and date stamped, interval or manual logging
- Three-year warranty on the instrument; one-year on the probes
- GLP assisting, records calibration data in memory
- Available with 4, 10, and 20-m cable lengths
- IP-67, impact-resistant, waterproof case
- Easy-to-use, screw-on cap DO membranes
- RS-232 interface for PC connection

Options to Fit Your Applications!

- Battery Options – The unit is powered by alkaline batteries or an optional rechargeable battery pack with quick-charge feature.
- Optional Barometer – Internal barometer can be user-calibrated and displayed along with other data, used in dissolved oxygen calibrations, and logged to memory for tracking changes in barometric pressure. (Choose 556-02)
- Optional Flow Cell - The 5083 flow cell can be used for ground water applications or anytime water is pumped for sampling.
- Carrying Case – The instrument comes standard with YSI 5061, a soft-sided carrying case with enough space for the 556, a 20-meter cable, and calibrating supplies. An optional 5080 hard-sided carrying case is also available.
- Confidence Solution® - Quality assurance ensured. Quickly check conductivity, pH, and ORP readings with one solution.

Pure Data for a Healthy Planet.®

A rugged, cost-effective multiparameter handheld system designed for the field!

www.YSI.com/556

+1 937 767 7241
800 897 4151 (US)
www.ysi.com

YSI Environmental
+1 937 767 7241
Fax +1 937 767 9353
environmental@ysi.com

YSI Integrated Systems
+1 508 748 0366
systems@ysi.com

SonTek/YSI
+1 858 546 8327
inquiry@sontek.com

YSI Gulf Coast
+1 225 753 2650
environmental@ysi.com

AMJ Environmental
+1 727 565 2201
amj@ysi.com

YSI Hydrodata (UK)
+44 1462 673 581
europe@ysi.com

YSI Middle East (Bahrain)
+973 1753 6222
halsalem@ysi.com

YSI (Hong Kong) Limited
+852 2891 8154
hongkong@ysi.com

YSI (China) Limited
+86 532 575 3636
beijing@ysi-china.com.

YSI Nanotech (Japan)
+81 44 222 0009
nanotech@ysi.com

YSI India
+91 989 122 0639
sham@ysi.com

YSI Australia
+61 7 390 17223
acorbett@ysi.com

ISO 9001
ISO 14001

5563 MPS Sensor Specifications

Dissolved Oxygen (% saturation)	Sensor Type	Steady state polarographic
	Range	0 to 500% air saturation
	Accuracy	0 to 200% air saturation, ± 2% of the reading or ±2% air saturation, whichever is greater; 200 to 500% air saturation, ± 6% of the reading
	Resolution	0.1% air saturation
Dissolved Oxygen (mg/L)	Sensor Type	Steady state polarographic
	Range	0 to 50 mg/L
	Accuracy	0 to 20 mg/L, ± 2% of the reading or ±0.2 mg/L, whichever is greater; 20 to 50 mg/L, ± 6% of the reading
	Resolution	0.01 mg/L
Temperature	Sensor Type	YSI Temperature Precision™ thermistor
	Range	-5 to 45°C
	Accuracy	± 0.15°C
	Resolution	0.1°C
Conductivity	Sensor Type	4-electrode cell with autoranging
	Range	0 to 200 mS/cm
	Accuracy	± 0.5% of reading or ± 0.001 mS/cm; whichever is greater (4-meter cable) ± 1.0% of reading or ± 0.001 mS/cm; whichever is greater (20-meter cable)
	Resolution	0.001 mS/cm to 0.1 mS/cm (range-dependent)
Salinity	Sensor Type	Calculated from conductivity and temperature
	Range	0 to 70 ppt
	Accuracy	± 1.0% of reading or ±0.1 ppt, whichever is greater
	Resolution	0.01 ppt
pH (optional)	Sensor Type	Glass combination electrode
	Range	0 to 14 units
	Accuracy	±0.2 units
	Resolution	0.01 units
ORP (optional)	Sensor Type	Platinum button
	Range	-999 to +999 mV
	Accuracy	± 20 mV
	Resolution	0.1 mV
Total Dissolved Solids (TDS)	Sensor Type	Calculated from conductivity (variable constant, default 0.65)
	Range	0 to 100 g/L
	Resolution	4 digits
Barometer (optional)	Range	500 to 800 mm Hg
	Accuracy	± 3 mm Hg within ± 10°C temperature range from calibration point
	Resolution	0.1 mm Hg

YSI 556 Instrument Specifications

Size	11.9 cm width x 22.9 cm lenth (4.7 in. x 9 in.)
Weight with batteries	2.1 lbs. (916 grams)
Power	4 alkaline C-cells; optional rechargeable pack
Cables	4-, 10-, and 20-m (13.1, 32.8, 65.6 ft.) lengths
Warranty	3-year instrument; 1-year probes and cables
Communication Port	RS-232 Serial
Data Logger	49,000 data sets, date and time stamp, manual or logging, with user-selectable intervals

556 Ordering Information (Order all items separately)

556-01	Instrument (with 5061 large, soft-sided carrying case)
556-02	Instrument with barometer option (with 5061 carrying case)
5563-4	4-m cable and DO/temp/conductivity
5563-10	10-m cable and DO/temp/conductivity
5563-20	20-m cable and DO/temp/conductivity
5564	pH Probe for any 5563 cable
5565	pH/ORP Probe for any 5563 cable
6118	Rechargeable battery pack kit (includes battery, adapter, charger)
614	Ultra clamp, C-clamp mount
616	Charger, cigarette lighter
4654	Tripod (small tripod for instrument)
5060	Small carrying case, soft-sided (fits instrument and 4-m cable)
5065	Form-fitted carrrier with shoulder strap
5080	Small carrying case, hard-sided (fits instrument, 4-m cable, flow cell, batteries, membrane kit, calibration bottles)
5083	Flow cell
5085	Hands-free harness
5580	Confidence Solution® (insure probe accuracy with a simple field-check for conductivity, pH, and ORP)

The 5080 carrying case with 556, 5563-4 cable, and 5083 flow cell.

YSI 600XL and 600XLM Sondes

The YSI 600XL and 600XLM

Measure multiple parameters simultaneously

The YSI 600XL and YSI 600XLM compact sondes measure eleven parameters simultaneously:

Temperature	TDS
Conductivity	pH
Specific Conductance	ORP
Salinity	Depth or Level
Resistivity	Rapid Pulse™ DO (% and mg/L)

Connect with Data Collection Platforms

Either sonde can easily connect to the YSI 6200 DAS (Data Acquisition System), YSI EcoNet™ or your own data collection platform, via SDI-12 for remote and real-time data acquisition applications.

Economical Logging System

The YSI 600XLM is an economical logging system for long-term, *in situ* monitoring and profiling. It will log all parameters at programmable intervals and store 150,000 readings. At one-hour intervals, the instrument will log data for about 75 days utilizing its own power source. The 600XL can also be utilized in the same manner with user-supplied external power.

- Either sonde fits down 2-inch wells
- Horizontal measurements in very shallow waters
- Stirring-independent Rapid Pulse® dissolved oxygen sensor
- Field-replaceable sensors
- Easily connects to data collection platforms
- Available with detachable cables to measure depth up to 200 feet
- Compatible with YSI 650 Multiparameter Display System
- Use with the YSI 5083 flow cell for groundwater applications

Pure Data for a Healthy Planet.®

Economical, multiparameter sampling or logging in a compact sonde

Sensor performance verified*

The 6820 V2 and 6920 V2 sondes use sensor technology that was verified through the US EPA's Environmental Technology Verification Program (ETV). For information on which sensors were performance-verified, turn this sheet over and look for the ETV logo.

ETV

www.ysi.com

To order, or for more info, contact YSI Environmental.

+1 937 767 7241
800 897 4151 (US)
www.ysi.com

YSI Environmental
+1 937 767 7241
Fax +1 937 767 9353
environmental@ysi.com

Endeco/YSI
+1 508 748 0366
Fax +1 508 748 2543
systems@ysi.com

SonTek/YSI
+1 858 546 8327
Fax +1 858 546 8150
inquiry@sontek.com

YSI Gulf Coast
+1 225 753 2650
Fax +1 225 753 8669
environmental@ysi.com

YSI Hydrodata (UK)
+44 1462 673 581
Fax +44 1462 673 582
europe@ysi.com

YSI Middle East (Bahrain)
+973 1753 6222
Fax +973 1753 6333
halsalem@ysi.com

YSI (Hong Kong) Limited
+852 2891 8154
Fax +852 2834 0034
hongkong@ysi.com

YSI (China) Limited
+86 10 5203 9675
Fax +86 10 5203 9679
beijing@ysi-china.com

YSI Nanotech (Japan)
+81 44 222 0009
Fax +81 44 221 1102
nanotech@ysi.com

ISO 9001
ISO 14001

Yellow Springs, Ohio Facility

Printed in USA 0107 E55-01

CE

*Sensors with labels with the ETV logo were submitted for ETV program on the YSI 6600XL. Information on the performance characteristics of YSI water quality sensors can be found also at www.epa.gov/etv, or call YSI at 800 897 4151 for the ETV verification report. Use of the ETV name or logo does not imply approval or certification of this product nor does it make any explicit or implied warranties or guarantees as to product performance.

YSI incorporated
Who's Minding the Planet?

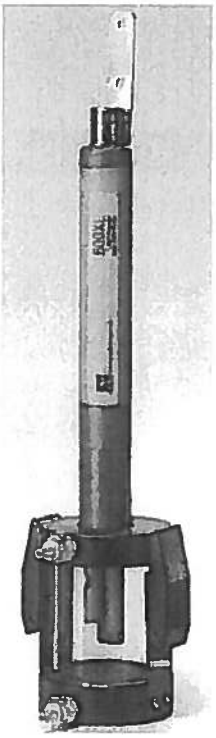
YSI 600XL & 600XLM Sensor Specifications

		Range	Resolution	Accuracy
Dissolved Oxygen % Saturation ETV 6562 Rapid Pulse™ Sensor*		0 to 500%	0.1%	0 to 200%: ±2% of reading or 2% air saturation, whichever is greater; 200 to 500%: ±6% of reading
Dissolved Oxygen mg/L ETV 6562 Rapid Pulse™ Sensor*		0 to 50 mg/L	0.01 mg/L	0 to 20 mg/L: ± 0.2 mg/L or 2% of reading, whichever is greater; 20 to 50 mg/L: ±6% of reading
Conductivity* ETV 6560 Sensor*		0 to 100 mS/cm	0.001 to 0.1 mS/cm (range dependent)	±0.5% of reading + 0.001 mS/cm
Salinity		0 to 70 ppt	0.01 ppt	±1% of reading or 0.1 ppt, whichever is greater
Temperature ETV 6560 Sensor*		-5 to +50°C	0.01°C	±0.15°C
pH ETV 6561 Sensor*		0 to 14 units	0.01 unit	±0.2 unit
ORP		-999 to +999 mV	0.1 mV	±20 mV
Depth & Level	Medium	0 to 200 ft, 61 m	0.001 ft, 0.001 m	±0.4 ft, ±0.12 m
	Shallow	0 to 30 ft, 9.1 m	0.001 ft, 0.001 m	±0.06 ft, ±0.02 m
	Vented Level	0 to 30 ft, 9.1 m	0.001 ft, 0.001 m	±0.01 ft, 0.003 m

* Report outputs of specific conductance (conductivity corrected to 25° C), resistivity, and total dissolved solids are also provided. These values are automatically calculated from conductivity according to algorithms found in *Standard Methods for the Examination of Water and Wastewater* (ed 1989)

YSI 600XL & 600XLM Sonde Specifications

Medium		Fresh, sea or polluted water
Temperature	Operating	-5 to +50°C
	Storage	-10 to +60°C
Communications		RS-232, SDI-12
Software		EcoWatch®
Dimensions 600XL | 600XLM	Diameter	1.65 in, 4.19 cm | 1.65 in, 4.9 cm
	Length	16 in, 40.6 cm | 21.3 in, 54.1 cm
	Weight	1.3 lbs, 0.59 kg | 1.5 lbs, 0.69 kg
Power	External	12 V DC
	Internal (600XLM only)	4 AA size alkaline batteries



YSI model 5083 flow cell and 600XL. This is an ideal combination for groundwater applications.

HI 98129

Combo pH/EC/TDS/Temperature Tester with Low Range EC

Description

The HI 98129 Combo waterproof tester offer high accuracy pH, EC/TDS and temperature measurements in a single tester! No more switching between meters for your routine measurements. The waterproof Combo (it even floats) has a large easy-to-read, dual-level LCD and automatic shut-off. pH and EC/TDS readings are automatically compensated for the effects of temperature (ATC). This technologically advanced tester has a replaceable pH electrode cartridge with an extendable cloth junction as well as an EC/TDS graphite electrode that resists contamination by salts and other substances. This gives these meters a greatly extended life. Your tester no longer needs to be thrown away when the pH sensor is exhausted.

The EC/TDS conversion factor is user selectable as is the temperature compensation coefficient (ß). Fast, efficient, accurate and portable, the Combo pH, EC/TDS and temperature tester brings you all the features you've asked for and more!

Specifications

Range	pH	0.00 to 14.00 pH
Range	EC	0 to 3999 µS/cm
Range	TDS	0 to 2000 ppm
Range	Temperature	0.0 to 60.0°C / 32 to 140.0°F
Resolution	pH	0.01 pH
Resolution	EC	1 µS/cm
Resolution	TDS	1 ppm
Resolution	Temperature	0.1°C / 0.1°F
Accuracy	pH	±0.05 pH
Accuracy	EC/TDS	±2% F.S.
Accuracy	Temperature	±0.5°C / ±1°F
Temperature Compensation		pH: automatic; EC/TDS: automatic with ß adjustable from 0.0 to 2.4% / °C
Calibration	pH	automatic, 1 or 2 points with 2 sets of memorized buffers (pH 4.01 / 7.01 / 10.01 or 4.01 / 6.86 / 9.18)
Calibration	EC/TDS	automatic, 1 point
TDS Conversion Factor		adjustable from 0.45 to 1.00
pH Electrode		HI 73127 (replaceable; included)
Environment		0 to 50°C (32 to 122°F); RH max 100%
Battery Type / Life		4 x 1.5V / approx. 100 hours of continuous use; auto-off after 8 minutes of non-use
Dimensions		163 x 40 x 26 mm (6.4 x 1.6 x 1.0")
Weight		100 g (3.5 oz.)

HQ30d Portable pH, Conductivity, Dissolved Oxygen (DO), ORP, and ISE Multi-Parameter Meter

Product # HQ30D53800000 Quantity
USD Price: $702.00

★★★★★ 5/5

Read 1 Review Write a Review Follow this product

Portable meter measures critical water quality parameters - without the need for multiple instruments
Single input channel for flexible measurement of pH, Conductivity, Dissolved Oxygen (DO), BOD, ORP, Ammonia, Ammonium, Fluoride, Chloride, Sodium, and temperature - any IntelliCAL™ smart probe

Intuitive user interface for simple operation and accurate results
Guided calibration and check standard routines reduce calibration errors. Stabilization alerts and visual measurement lock ensure that you can trust the accuracy of the results

Trust your measurements - IntelliCAL™ smart probes store all calibrations in the probe
Calibration history allows quick and easy change out of probes without re-calibrating. The HQd™ smart system records serial numbers, current calibration data, user ID, sample ID, time, and date automatically in the data log for complete GLP traceability

Designed for demanding conditions
Rugged, waterproof (IP67) meter provides worry-free, reliable operation in lab or field environments

Convenient kit includes everything you need to start testing
Meter kit includes 4 AA batteries, quick-start guide, user manual, and documentation CD

Specifications

AC and USB Operation	optional
Automatic Buffer Recognition	Color-coded: 4.01, 7.00, 10.01 pH IUPAC: 1.679, 4.005, 7.000, 10.012, 12.45 DIN: 1.09, 4.65, 9.23 User-defined custom buffer sets
Barometric Pressure Measurement	For automatic compensation of DO when using an LDO or LBOD probe
Battery Requirements	4 AA
Benchtop	with stand
BOD5/CBOD resolution	Available when used with Hach WIMS BOD Manager software
Cable resistance correction	Digital - not needed
Calibration curves display	Calibration summary data logged and displayed
Calibration Intervals/Alerts/Reminder	2 hours to 7 days
Compliance	CE, WEEE
Conductivity Accuracy	± 0.5 % from (1 µS/cm - 200 mS/cm)
Conductivity measurement	5 different stability modes
Conductivity Measurement Range	0.01 µS/cm to 200 mS/cm
Conductivity resolution	0.01 µS/cm with 2 digits
Custom Calibration Standards	User-defined standard sets
Data Export	Download via USB connection to PC or flash stick Automatically transfer entire data log or as readings are taken
Data Memory	500 results
Digital (intelligent) electrode inputs	2
Dimensions (H x W x D)	7.8 in x 3.7 in x 1.4 in (197 mm x 95 mm x 36 mm)
Display	Display readings from one or two probes Simultaneous readings from two probes (HQ40d only) pH: pH, mV, temperature Conductivity: Conductivity, TDS, salinity, resistivity, temperature LDO: dissolved oxygen, pressure, temperature LBOD: dissolved oxygen, pressure, temperature ORP/Redox: mV, temperature Sodium: Sodium, mV, temperature
Display Lock Function	Continuous measurement or press to read mode available with averaging function for LDO measurement
Display Type	240 x 160 pixel Display readings from one or two probes pH: pH, mV, temperature Conductivity: Conductivity, TDS, salinity, resistivity, temperature LDO: dissolved oxygen, pressure, temperature ORP/Redox: mV, temperature Sodium: Sodium, mV, temperature
DO Measurement Range	0.01 to 20 mg/L (0 to 200%)
DO Resolution	0.01 mg/L
Fixed Buffer Selection	IUPAC standards (DIN 19266) or Technical buffer (DIN 19267) or 4, 7, 10 series or user defined
Inputs	M12 digital (1) for IntelliCAL probes
Interface Languages	13**
Internal Data Storage	500
IP Rating	IP67
Languages	English, French, German, Italian, Spanish, Danish, Dutch, Polish, Portuguese, Turkish, Swedish, Czech, Russian
mV Accuracy	± 0.1 mV
mV Measurement at Stable Reading	5 (auto) stabilization settings
mV Resolution	0.1 mV
Operating Error Messages	Text messages displayed
Operating Humidity	90 % relative humidity (non-condensing)
Operating Interface	Keypad
Operating Temperature	5 to 45 °C
ORP Electrode Calibration	Predefined ORP standards (including Zobell's solution)
Outputs	USB to PC / flash stick
PC Data Transfer Software	Included
pH Measurement at stable reading	5 stabilization settings
Printer	Optional accessory
Salinity Resolution	0.01 ppt
Warranty	3 years
Water Resistance	Meter Casing: 1 meter submersion for 30 minutes (IP67)
Weight	0.74 lbs (0.335 kg)

2100P and 2100P IS Portable Turbidimeter

Features and Benefits

Laboratory Quality in a Portable Unit

The Hach 2100P and 2100P IS Portable Turbidimeters offer a level of performance previously possible only with laboratory instruments. Microprocessor-controlled operation and Hach's unique Ratio™ optics bring great accuracy, sensitivity, and reliability to field and in-plant testing.

Two Models for Specific Requirements

- **2100P Turbidimeter**—*Get fast, accurate turbidity testing in the field or the lab, over a wide range of samples. Compliant with USEPA Method 180.1 design criteria.*
- **2100P IS Turbidimeter**—*Designed to meet international standards that mandate measurement using an LED light source.*

Two-detector Optical System

The two-detector optical system compensates for color in the sample, light fluctuation, and stray light, enabling analysts to achieve laboratory-grade performance on a wide range of samples, even under difficult, onsite conditions.

The Hach 2100P and 2100P IS Portable Turbidimeters bring laboratory-level performance on-site, offering fast, accurate results and the ease-of-use analysts demand in the field. With a measurement range of 0 to 1000 NTU and a resolution of 0.01 NTU, the 2100P turbidimeter is ideal for regulatory monitoring, process control or field studies.

Specifications*

	2100P	2100P IS
Measurement Method	Nephelometric Ratio	
Regulatory	Meets EPA Method 180.1	Meets EN ISO 7027
Light Source	Tungsten lamp	Light-emitting diode (LED) @ 860 nm
Range		
Automatic Range Mode	0 to 1000 NTU	0 to 1000 FNU
Manual Range Selection	0 to 9.99, 0 to 99.9 and 0 to 1000 NTU	0 to 9.99, 0 to 99.9 and 0 to 1000 FNU
Accuracy	±2% of reading plus stray light	
Repeatability	±1% of reading, or 0.01 NTU, whichever is greater	±1% of reading, or 0.01 FNU, whichever is greater
Resolution	0.01 on lowest range	
Signal Averaging	Selectable on/off	
Power Requirement	4 AA alkaline batteries or optional battery eliminator	
Battery Life, Typical	300 tests with signal average mode off 180 tests with signal average mode on	
Operating Temperature	0 to 50°C (32 to 122°F)	
Sample Required	15 mL (0.5 oz.)	
Sample Cells	60 x 25 mm (2.36 x 1 in.) borosilicate glass with screw caps	
Dimensions	22.2 x 9.5 x 7.9 cm (8.75 x 3.75 x 3.12 in.)	
Weight	0.5 kg (1.1 lb.); shipping weight 2.7 kg (6 lb.)	
Warranty	2 years	

**Specifications subject to change without notice.*

DW = drinking water WW = wastewater municipal PW = pure water / power
IW = industrial water E = environmental C = collections FB = food and beverage

HACH®

Be Right™

2100Q and 2100Q *is* Portable Turbidimeter

The Hach 2100Q and 2100Q is Portable Turbidimeters offer unsurpassed ease of use and accuracy in turbidity measurement. Only Hach offers this unique combination of advanced features, such as assisted calibration and simplified data transfer, and measurement innovation, giving you accurate results every time.

Features and Benefits

Easy Calibration and Verification

Hach 2100Q and 2100Q *is* Portable Turbidimeters provide confidence your measurements are right every time. On-screen assisted calibration and verification save you time and ensure accuracy. With an easy-to-follow interface, complicated manuals are not needed to perform routine calibrations. Single-standard RapidCal™ calibration offers a simplified solution for low level measurements.

Simple Data Transfer

Data transfer with the optional USB + Power Module is simple, flexible, and doesn't require additional software. All data can be transferred to the module and easily downloaded to your computer with a USB connection, providing superior data integrity and availability. With two different module options, you can customize connectivity and power to meet your unique needs.

Accurate for Rapidly Settling Samples

The Hach 2100Q Portable Turbidimeter incorporates an innovative Rapidly Settling Turbidity™ mode to provide accurate, repeatable measurements for difficult to measure, rapidly settling samples. An exclusive algorithm that calculates turbidity based on a series of automatic readings eliminates redundant measurements and estimating.

Convenient Data Logging

Up to 500 measurements are automatically stored in the instrument for easy access and backup. Stored information includes: date and time, operator ID, reading mode, sample ID, sample number, units, calibration time, calibration status, error messages and the result.

Optical System for Precision in the Field

The two-detector optical system compensates for color in the sample, light fluctuation, and stray light, enabling analysts to achieve laboratory-grade performance on a wide range of samples, even under difficult site conditions.

Two Models for Specific Requirements

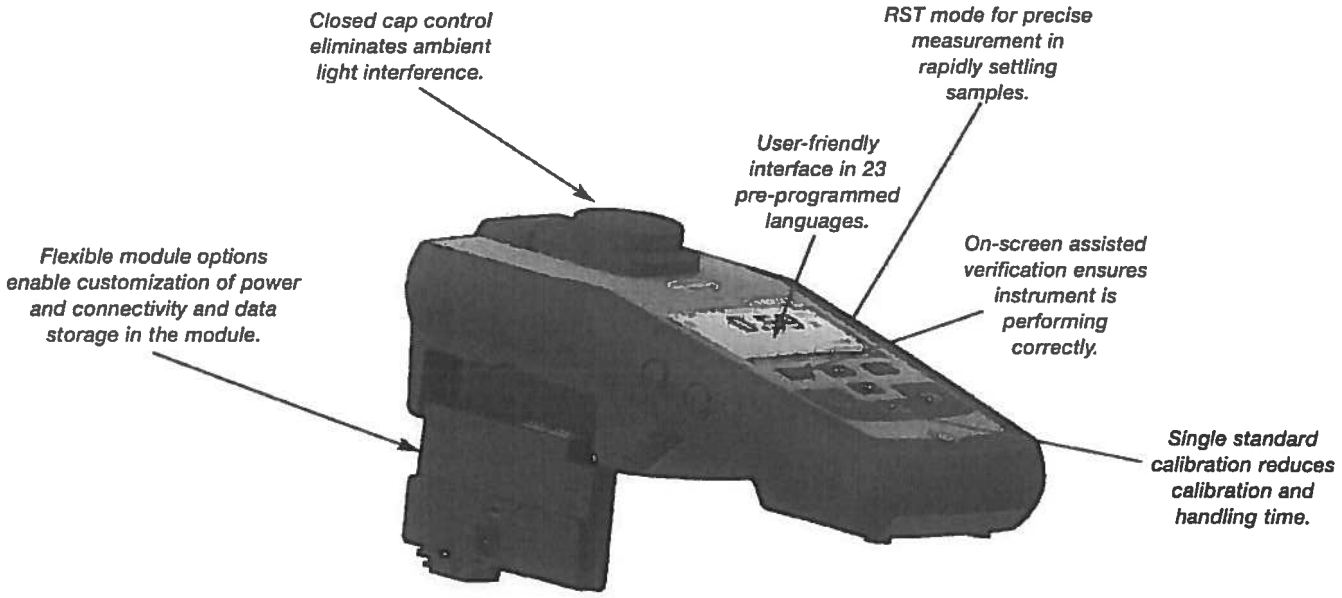
- **2100Q Turbidimeter**—*Compliant with USEPA Method 180.1 design criteria.*
- **2100Q *is* Turbidimeter**—*Compliant with ISO 7027 design criteria.*

DW = drinking water WW = wastewater municipal PW = pure water / power
IW = industrial water E = environmental C = collections FB = food and beverage

HACH®

Be Right™

Key Features



Specifications*

Measurement Method
Ratio turbidimetric determination using a primary nephelometric light scatter signal (90°) to the transmitted light scatter signal.

Regulatory
2100Q: Meets EPA Method 180.1
2100Q *is:* Meets ISO 7027

Light Source
2100Q: Tungsten filament lamp
2100Q *is:* Light-emitting diode (LED) @ 860 nm

Range
0 to 1000 NTU (FNU)

Accuracy
±2% of reading plus stray light from 0 to 1000 NTU

Repeatability
±1% of reading, or 0.01 NTU (FNU), whichever is greater

Resolution
0.01 NTU on lowest range

Stray Light
<0.02 NTU (FNU)

Signal Averaging
Selectable on/off

Detector
Silicon photovoltaic

Reading Modes (user selectable)
Normal (Push to Read)
Signal Averaging
Rapidly Settling Turbidity

Data Logger
500 records

Power Requirement
110-230 Vac, 50/60 Hz (with Power or USB+Power Module)
4 AA alkaline batteries
Rechargeable NiMH (for use with USB+Power Module)

Operating Conditions
Temperature: 0 to 50°C (32 to 122°F)
Relative Humidity: 0 to 90% @ 30°C,
0 to 80% @ 40°C, 0 to 70% @ 50°C, noncondensing

Storage Conditions
-40 to 60°C (-40 to 140°F), instrument only

Languages
English, French, German, Italian, Spanish, Portuguese (BR), Portuguese (PT), Bulgarian, Chinese, Czech, Danish, Dutch, Finnish, Greek, Hungarian, Japanese, Korean, Polish, Romanian, Russian, Slovenian, Swedish, Turkish

Interface
Optional USB

Instrument Enclosure Rating
IP67 (closed lid, battery compartment excluded)

Protection Class
Power Supply: Class II

Certification
CE certified

Sample Required
15 mL (0.3 oz.)

Sample Cells
60 x 25 mm (2.36 x 1 in.) borosilicate glass with screw cap

Dimensions
22.9 x 10.7 x 7.7 cm (9.0 x4.2 x 3.0 in.)

Weight
527 g (1.16 lb) without batteries
618 g (1.36 lb) with four AA alkaline batteries

Warranty
1 year

**Specifications subject to change without notice.*

Appendix D

NEORSD Chlorophyll *a* Sampling Field Sheet

Stream:_____ Collectors:_____
Location:_____ Date:_____
RM:_____ Time:_____
Lat/Long:_____

Number of Rocks:_____ Total Area Scraped:_____cm^2

Diameter of individual scrape	Area of individual scrape
1_____	1_____
2_____	2_____
3_____	3_____
4_____	4_____
5_____	5_____
6_____	6_____
7_____	7_____
8_____	8_____
9_____	9_____
10_____	10_____
11_____	11_____
12_____	12_____
13_____	13_____
14_____	14_____
15_____	15_____
16_____	16_____
17_____	17_____
18_____	18_____
19_____	19_____
20_____	20_____
21_____	21_____
22_____	22_____
23_____	23_____
24_____	24_____
25_____	25_____
	Total:_____

Diameter to Area Conversion

Diameter (cm)	Area (cm2)
1.6	2.011
1.7	2.27
1.8	2.545
1.9	2.835
2.0	3.142
2.1	3.464
2.2	3.801
2.3	4.155

Total Sample Volume_____ml

Filter 1 LABLynx ID_____
Vol _____ml

Filter 2 LABLynx ID_____
Vol _____ml

Filter 3 LABLynx ID_____
Vol _____ml

Water Column Chlorophyll Sample

Filter 1 LABLynx ID_____
Vol _____ml

Filter 2 LABLynx ID_____
Vol _____ml

Filter 3 LABLynx ID_____
Vol _____ml

Flow: None Low Normal Elevated High

Turbidity: Clear Low Moderate* High*

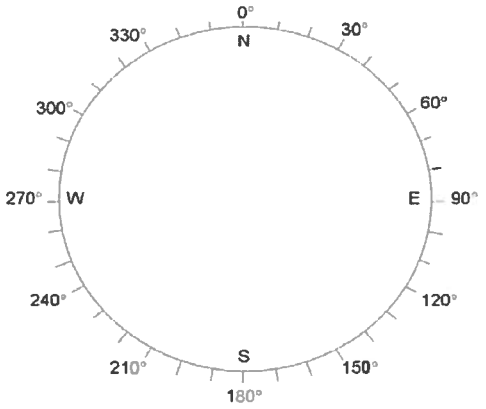
*Explain_____

Sky: Overcast Cloudy Partly Cloudy Mostly Clear Clear

Canopy: Open Mostly Open Partly Closed Closed

Riparian None Narrow L R Moderate L R Wide L R

Downstream Channel Direction



Clinometer

Left Bank _____°
Right Bank _____°

Left Bank _____°
Right Bank _____°

Left Bank _____°
Right Bank _____°

Stream Widths

_____m _____m _____m

Record two most predominate substrates with an X, and check all present.

	Riffle	Run	Reach
Boulder/Slabs			
Bedrock			
Boulder/Slabs			
Cobble			
Gravel			
Sand			
Silt			
Hardpan			
Detritus			
Artificial			

Substrate Origin

____Limestone ____Tills ____Rip-rap
____Sandstone ____Shale ____Wetlands
____Lacustrine ____Hardpan ____Coal Fines

Silt

____Heavy ____Moderate ____Normal ____None

Embeddedness

____Extensive ____Moderate ____Normal ____None

Notes:_____

_____

Length of Reach: _____m

Stream Drawing

Appendix E

COMMONWEALTH OF PENNSYLVANIA
DEPARTMENT OF ENVIRONMENTAL PROTECTION

BUREAU OF LABORATORIES

LABORATORY ACCREDITATION PROGRAM

pennsylvania
DEPARTMENT OF ENVIRONMENTAL PROTECTION

Certifies That

68-03670

Northeast Ohio Regional Sewer District Analytical Services

4747 East 49th Street, Cuyahoga Heights, OH 44125

Having duly met the requirement of
The act of June 29, 2002 (P.L. 596, No. 90)
dealing with Environmental Laboratories Accreditation
(27 Pa. C.S. §§4104-4113) and the
National Environmental Laboratory Accreditation Program Standard

is hereby approved as an

Accredited Laboratory

As more fully described in the attached Scope of Accreditation

Expiration Date: **11/30/2015**

Certificate Number: **008**

Aaren S. Alger, Chief
Laboratory Accreditation Program
Bureau of Laboratories

Continued accreditation status depends on successful ongoing participation in the program
Certificate not transferable Surrender upon revocation
To be conspicuously displayed at the Laboratory
Not valid unless accompanied by a valid Scope of Accreditation
Shall not be used to imply endorsement by the Commonwealth of Pennsylvania
Customers are urged to verify the laboratory's current accreditation status
PA DEP is a NELAP recognized accreditation body

pennsylvania
DEPARTMENT OF ENVIRONMENTAL PROTECTION

Laboratory Scope of Accreditation

Attached to Certificate of Accreditation 008-001 expiration date November 30, 2015. This listing of accredited analytes should be used only when associated with a valid certificate of accreditation.

DEP Laboratory ID: 68-03670 EPA Lab Code: OH00300 TNI Code: (216) 641-6000

Northeast Ohio Regional Sewer District Analytical Services
4747 East 49th Street
Cuyahoga Heights, OH 44125

Matrix: Non-Potable Water

Method	Revision	Analyte	Accreditation Type	Primary	Effective Date
ASTM D4839-03		Total organic carbon (TOC)	NELAP	PA	11/25/2014
EPA 1000.0		Pimephales promelas	NELAP	PA	1/8/2009
EPA 1002.0		Ceriodaphnia dubia	NELAP	PA	1/8/2009
EPA 160.4		Residue, volatile	NELAP	PA	10/22/2008
EPA 1600		Enterococci	NELAP	PA	11/22/2010
EPA 1603		E. coli (Enumeration)	NELAP	PA	11/29/2007
EPA 1631	E	Mercury	NELAP	PA	3/31/2008
EPA 180.1		Turbidity	NELAP	PA	12/31/2007
EPA 200.7	4.4	Aluminum	NELAP	PA	11/29/2007
EPA 200.7	4.4	Antimony	NELAP	PA	11/29/2007
EPA 200.7	4.4	Arsenic	NELAP	PA	11/29/2007
EPA 200.7	4.4	Barium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Beryllium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Cadmium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Calcium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Chromium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Cobalt	NELAP	PA	11/29/2007
EPA 200.7	4.4	Copper	NELAP	PA	12/31/2007
EPA 200.7	4.4	Iron	NELAP	PA	11/29/2007
EPA 200.7	4.4	Lead	NELAP	PA	11/29/2007
EPA 200.7	4.4	Magnesium	NELAP	PA	11/17/2010
EPA 200.7	4.4	Manganese	NELAP	PA	11/29/2007
EPA 200.7	4.4	Molybdenum	NELAP	PA	11/29/2007
EPA 200.7	4.4	Nickel	NELAP	PA	11/29/2007
EPA 200.7	4.4	Potassium	NELAP	PA	12/31/2007
EPA 200.7	4.4	Selenium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Silver	NELAP	PA	11/29/2007
EPA 200.7	4.4	Sodium	NELAP	PA	12/31/2007
EPA 200.7	4.4	Thallium	NELAP	PA	4/15/2014
EPA 200.7	4.4	Tin	NELAP	PA	11/29/2007
EPA 200.7	4.4	Titanium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Vanadium	NELAP	PA	11/29/2007
EPA 200.7	4.4	Zinc	NELAP	PA	12/31/2007
EPA 245.1	3.0	Mercury	NELAP	PA	11/29/2007
EPA 300.0	2.1	Bromide	NELAP	PA	11/22/2010
EPA 300.0	2.1	Chloride	NELAP	PA	11/22/2010
EPA 300.0	2.1	Fluoride	NELAP	PA	11/22/2010
EPA 300.0	2.1	Nitrate as N	NELAP	PA	11/22/2010
EPA 300.0	2.1	Orthophosphate as P	NELAP	PA	11/22/2010
EPA 300.0	2.1	Sulfate	NELAP	PA	11/22/2010

The Pennsylvania Department of Environmental Protection Laboratory Accreditation Program is a NELAP recognized Accreditation Body. Customers are urged to verify the laboratory's current accreditation standing.

pennsylvania
DEPARTMENT OF ENVIRONMENTAL PROTECTION

Laboratory Scope of Accreditation

Attached to Certificate of Accreditation 008-001 expiration date November 30, 2015. This listing of accredited analytes should be used only when associated with a valid certificate of accreditation.

DEP Laboratory ID: 68-03670 EPA Lab Code: OH00300 TNI Code: (216) 641-6000

Matrix: Non-Potable Water

Method	Revision	Analyte	Accreditation Type	Primary	Effective Date
EPA 3005	A	Preconcentration under acid	NELAP	PA	11/29/2007
EPA 3010	A	Hot plate acid digestion (HNO3 + HCl)	NELAP	PA	11/29/2007
EPA 3015		Microwave-assisted acid digestion	NELAP	PA	11/29/2007
EPA 310.2		Alkalinity as CaCO3	NELAP	PA	9/20/2012
EPA 350.1		Ammonia as N	NELAP	PA	11/29/2007
EPA 351.2		Kjeldahl nitrogen, total (TKN)	NELAP	PA	11/17/2010
EPA 353.2		Nitrate as N	NELAP	PA	11/29/2007
EPA 353.2		Total nitrate-nitrite	NELAP	PA	11/17/2010
EPA 365.1		Phosphorus, total	NELAP	PA	10/22/2008
EPA 410.4		Chemical oxygen demand (COD)	NELAP	PA	11/29/2007
EPA 420.4		Total phenolics	NELAP	PA	11/17/2010
EPA 445		Chlorophyll A	NELAP	PA	11/22/2010
EPA 6010		Aluminum	NELAP	PA	11/29/2007
EPA 6010		Antimony	NELAP	PA	11/29/2007
EPA 6010		Arsenic	NELAP	PA	11/29/2007
EPA 6010		Barium	NELAP	PA	11/29/2007
EPA 6010		Beryllium	NELAP	PA	11/29/2007
EPA 6010		Cadmium	NELAP	PA	11/29/2007
EPA 6010		Calcium	NELAP	PA	11/29/2007
EPA 6010		Chromium	NELAP	PA	11/29/2007
EPA 6010		Cobalt	NELAP	PA	11/29/2007
EPA 6010		Copper	NELAP	PA	12/31/2007
EPA 6010		Iron	NELAP	PA	11/29/2007
EPA 6010		Lead	NELAP	PA	11/29/2007
EPA 6010		Magnesium	NELAP	PA	11/29/2007
EPA 6010		Manganese	NELAP	PA	11/29/2007
EPA 6010		Molybdenum	NELAP	PA	11/29/2007
EPA 6010		Nickel	NELAP	PA	11/29/2007
EPA 6010		Potassium	NELAP	PA	12/31/2007
EPA 6010		Selenium	NELAP	PA	11/29/2007
EPA 6010		Silver	NELAP	PA	11/29/2007
EPA 6010		Sodium	NELAP	PA	12/31/2007
EPA 6010		Thallium	NELAP	PA	4/15/2014
EPA 6010		Tin	NELAP	PA	11/29/2007
EPA 6010		Titanium	NELAP	PA	11/29/2007
EPA 6010		Vanadium	NELAP	PA	11/29/2007
EPA 6010		Zinc	NELAP	PA	12/31/2007
EPA 7470		Mercury	NELAP	PA	11/29/2007
Enterolert		Enterococci (Enumeration)	NELAP	PA	11/22/2010
OIA 1677		Available cyanide	NELAP	PA	11/29/2007
SM 2540 B		Residue, total	NELAP	PA	11/29/2007
SM 2540 C		Residue, filterable (TDS)	NELAP	PA	11/29/2007
SM 2540 D		Residue, nonfilterable (TSS)	NELAP	PA	11/29/2007
SM 2540 F		Residue, settleable	NELAP	PA	11/29/2007
SM 2550 B		Temperature, deg. C	NELAP	PA	10/22/2008
SM 3500-Cr B	20-22	Chromium VI	NELAP	PA	11/29/2007
SM 4500-CN- C/E		Total cyanide	NELAP	PA	11/29/2007

The Pennsylvania Department of Environmental Protection Laboratory Accreditation Program is a NELAP recognized Accreditation Body. Customers are urged to verify the laboratory's current accreditation standing.

pennsylvania
DEPARTMENT OF ENVIRONMENTAL PROTECTION

Laboratory Scope of Accreditation

Attached to Certificate of Accreditation 008-001 expiration date November 30, 2015. This listing of accredited analytes should be used only when associated with a valid certificate of accreditation.

DEP Laboratory ID: 68-03670 EPA Lab Code: OH00300 TNI Code: (216) 641-6000

Matrix: Non-Potable Water

Method	Revision	Analyte	Accreditation Type	Primary	Effective Date
SM 4500-CN- G		Amenable cyanide	NELAP	PA	11/29/2007
SM 4500-Cl E		Total residual chlorine	NELAP	PA	11/29/2007
SM 4500-Cl- C		Chloride	NELAP	PA	11/19/2012
SM 4500-H+ B		pH	NELAP	PA	11/29/2007
SM 4500-NO2- B		Nitrite as N	NELAP	PA	11/29/2007
SM 4500-Norg B		Kjeldahl nitrogen, total (TKN)	NELAP	PA	10/22/2008
SM 4500-P B		Preliminary treatment of phosphate samples	NELAP	PA	11/13/2013
SM 4500-P E		Orthophosphate as P	NELAP	PA	11/13/2013
SM 5210 B		Biochemical oxygen demand (BOD)	NELAP	PA	11/29/2007
SM 5210 B		Carbonaceous BOD (CBOD)	NELAP	PA	11/29/2007
SM 9222 D		Fecal coliform (Enumeration)	NELAP	PA	11/29/2007
SM 9223 Colilert MPN or QT		E. coli (Enumeration)	NELAP	PA	11/29/2007
SM 9223 Colilert MPN or QT		Total coliform (Enumeration)	NELAP	PA	11/22/2010

Matrix: Solid and Chemical Materials

Method	Revision	Analyte	Accreditation Type	Primary	Effective Date
EPA 245.1	3.0	Mercury	NELAP	PA	11/22/2010
EPA 3051		Microwave digestion of solids (HNO3 only)	NELAP	PA	11/17/2010
EPA 6010		Aluminum	NELAP	PA	11/22/2010
EPA 6010		Antimony	NELAP	PA	11/13/2013
EPA 6010		Arsenic	NELAP	PA	11/22/2010
EPA 6010		Barium	NELAP	PA	11/22/2010
EPA 6010		Beryllium	NELAP	PA	11/22/2010
EPA 6010		Cadmium	NELAP	PA	11/22/2010
EPA 6010		Calcium	NELAP	PA	11/22/2010
EPA 6010		Chromium	NELAP	PA	11/22/2010
EPA 6010		Cobalt	NELAP	PA	11/22/2010
EPA 6010		Copper	NELAP	PA	11/22/2010
EPA 6010		Iron	NELAP	PA	11/22/2010
EPA 6010		Lead	NELAP	PA	11/22/2010
EPA 6010		Magnesium	NELAP	PA	11/22/2010
EPA 6010		Manganese	NELAP	PA	11/22/2010
EPA 6010	B	Metals by ICP/AES	NELAP	PA	1/22/2013
EPA 6010		Molybdenum	NELAP	PA	11/22/2010
EPA 6010		Nickel	NELAP	PA	11/22/2010
EPA 6010		Potassium	NELAP	PA	11/22/2010
EPA 6010		Selenium	NELAP	PA	11/22/2010
EPA 6010		Silver	NELAP	PA	11/22/2010
EPA 6010		Sodium	NELAP	PA	11/22/2010
EPA 6010		Thallium	NELAP	PA	11/22/2010
EPA 6010		Tin	NELAP	PA	4/15/2013
EPA 6010		Titanium	NELAP	PA	11/22/2010
EPA 6010		Vanadium	NELAP	PA	11/22/2010
EPA 6010		Zinc	NELAP	PA	11/22/2010

The Pennsylvania Department of Environmental Protection Laboratory Accreditation Program is a NELAP recognized Accreditation Body. Customers are urged to verify the laboratory's current accreditation standing.

pennsylvania
DEPARTMENT OF ENVIRONMENTAL PROTECTION

Laboratory Scope of Accreditation

Attached to Certificate of Accreditation 008-001 expiration date November 30, 2015. This listing of accredited analytes should be used only when associated with a valid certificate of accreditation.

DEP Laboratory ID: 68-03670 EPA Lab Code: OH00300 TNI Code: (216) 641-6000

The Pennsylvania Department of Environmental Protection Laboratory Accreditation Program is a NELAP recognized Accreditation Body. Customers are urged to verify the laboratory's current accreditation standing.

Appendix F

Northeast Ohio Regional Sewer District
Protecting Your Health and Environment

February 4, 2015

Mr. John Rhoades
Supervisor of Environmental Assessment
Northeast Ohio Regional Sewer District
4747 East 49th Street
Cuyahoga Heights, Ohio 44125

Dear Mr. Rhoades:

This letter is to acknowledge that I am responsible for assisting the Northeast Ohio Regional Sewer District's Water Quality and Industrial Surveillance Division in conducting stream habitat assessments using the Qualitative Habitat Evaluation Index for the 2015 Big Creek, Cuyahoga River, Euclid Creek, Hemlock Creek, Mill Creek, and West Creek Environmental Monitoring Project Study Plans.

It is understood that an Ohio Environmental Protection Agency Level 3 Qualified Data Collector Certification for Stream Habitat Assessment is required to perform these tasks and that I am responsible for maintaining my Level 3 QDC Certification during the term of these Study Plans.

In addition, I have not been convicted nor pleaded guilty to a Violation of Section 2911.21 of the Revised Code (criminal trespass) or a substantially similar municipal ordinance within the previous five years.

Sincerely,

Jonathan Brauer
Stormwater Inspector
Northeast Ohio Regional Sewer District
4747 East 49th Street
Cuyahoga Heights, Ohio 44125

THIRD ROCK CONSULTANTS

April 14, 2015

Mr. John Rhoades
Manager of Water Quality and Industrial Surveillance
Northeast Ohio Regional Sewer District
4747 East 49th Street
Cuyahoga Heights, Ohio 44125

Re: *2015 Benthic Services*
PO 15001329

Dear Mr. Rhoades:

This letter is to acknowledge that I am responsible for the identification of benthic macroinvertebrates for the following Northeast Ohio Regional Sewer District Study Plans:

- 2015 Big Creek Environmental Monitoring
- 2015 Cuyahoga River Environmental Monitoring
- 2015 Euclid Creek Environmental Monitoring
- 2015 Hemlock Creek Environmental Monitoring
- 2015 Mill Creek Environmental Monitoring
- 2015 West Creek Environmental Monitoring

It is understood that an Ohio Environmental Protection Agency Level 3 Qualified Data Collector Certification for Benthic Macroinvertebrate, with the specialty of identification, is required to perform these tasks and that I am responsible for maintaining my Level 3 QDC Certification during the term of these Study Plans.

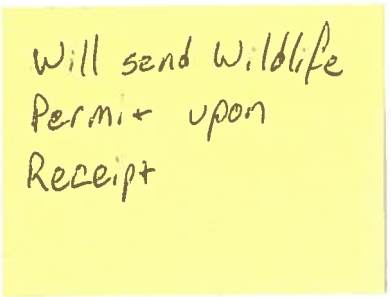
In addition, I have not been convicted nor pleaded guilty to a Violation of section 2911.21 of the Revised Code (criminal trespass) or a substantially similar municipal ordinance within the previous five years.

Sincerely,

Albert W. Remley II

Bert Remley
Senior Taxonomist
Bremley@thirdrockconsultants.com

Appendix G



Appendix H

References

Chlorophyll a Sampling and Field Filtering Standard Operating Procedure (SOP-EA001-00)

EPA New England- Region 1. (2005). *Standard operating procedure for calibration and field measurement procedures for the YSI Model 6-Series Sondes and Data Logger (Including: temperature, pH, specific conductance, turbidity, dissolved oxygen, chlorophyll, rhodamine WT, ORP, and barometric pressure)* (7th Revision). North Chelmsford, MA: The Office of Environmental Measurement and Evaluation, Ecosystem Assessment- Ecology Monitoring Team.

Ohio Environmental Protection Agency. (1987a). *Biological criteria for the protection of aquatic life: Volume II. Users manual for biological field assessment of Ohio surface waters* (Updated January 1988; September 1989; November 2006; August 2008). Columbus, OH: Division of Water Quality Monitoring and Assessment.

Ohio Environmental Protection Agency. (1987b). *Biological criteria for the protection of aquatic life: Volume III. Standardized biological field sampling and laboratory methods for assessing fish and macroinvertebrate communities* (Updated September 1989; March 2001; November 2006; and August 2008). Columbus, OH: Division of Water Quality Monitoring and Assessment.

Ohio Environmental Protection Agency. (1997). Draft. *Biological Criteria for the Protection of Aquatic Life: Volume IV: Fish and Macroinvertebrate Indicies for Ohio's Lake Erie Nearshore Waters, Harbors, and Lacustuaries*. Columbus, OH: Division of Surface Water, Ecological Assessment Unit.

Ohio Environmental Protection Agency. (2006). *Methods for assessing habitat in flowing waters: using the Qualitative Habitat Evaluation Index (QHEI)*. (Ohio EPA Technical Bulletin EAS/2006-06-1). Columbus, OH: Division of Surface Water; Division of Ecological Assessment Section.

Ohio Environmental Protection Agency. (2010). Draft. *Methods of Assessing Habitat in Lake Erie Shoreline Waters Using the Qualitative Habitat Evaluation Index (QHEI) Approach (Version 2.1)*. Columbus, OH: Division of Surface Water.

Ohio Environmental Protection Agency. (2011). *State of Ohio Water Quality Standards Ohio Administrative Code Chapter 3745-1* (Revision: March 16, 2011; Effective June 16, 2011). Columbus, OH: Division of Surface Water; Standards and Technical Support Section.

Ohio Environmental Protection Agency. (2012a). *Field Evaluation Manual for Ohio's Primary Headwater Habitat Stream*. Columbus, OH: Division of Surface Water; Division of Ecological Assessment Section.

Ohio Environmental Protection Agency. (2012b). *Ohio 2012 Integrated Water Quality Monitoring and Assessment Report.* Columbus, Ohio: Division of Surface Water.

Ohio Environmental Protection Agency. (2013). *Surface Water Field Sampling Manual for water chemistry, bacteria, and flows.* Columbus, OH: Division of Surface Water.